In Vitro Activity of Telithromycin against Spanish Streptococcus pneumoniae Isolates with Characterized Macrolide Resistance Mechanisms

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The susceptibilities to telithromycin of 203 Streptococcus pneumoniae isolates prospectively collected during 1999 and 2000 from 14 different geographical areas in Spain were tested and compared with those to erythromycin A, clindamycin, quinupristin-dalfopristin, penicillin G, cefotaxime, and levofloxacin. Telithromycin was active against 98.9% of isolates (MICs, ≤0.5 µg/ml), with MICs at which 90% of isolates are inhibited being 0.06 µg/ml, irrespective of the resistance genotype. The corresponding values for erythromycin were 61.0% (MICs, ≤0.25 µg/ml) and >64 µg/ml. The erm(B) gene (macrolide-lincosamide-streptogramin B resistance phenotype) was detected in 36.4% (n = 74) of the isolates, which corresponded to 93.6% of erythromycin-intermediate and -resistant isolates, whereas the mef(A) gene (M phenotype [resistance to erythromycin and susceptibility to clindamycin and spiramycin without blunting]) was present in only 2.4% (n = 5) of the isolates. One of the latter isolates also carried erm(B). Interestingly, in one isolate for which the erythromycin MIC was 2 µg/ml, none of these resistance genes could be detected. Erythromycin MICs for S. pneumoniae erm(B)-positive isolates were higher (range, 0.5 to >64 µg/ml) than those for erm(B)- and mef(A)-negative isolates (range, 0.008 to 2 µg/ml). The corresponding values for telithromycin were lower for both groups, with ranges of 0.004 to 1 and 0.002 to 0.06 µg/ml, respectively. The erythromycin MIC was high for a large number of erm(B)-positive isolates, but the telithromycin MIC was low for these isolates. These results indicate the potential usefulness of telithromycin for the treatment of infections caused by erythromycin-susceptible and -resistant S. pneumoniae isolates when macrolides are indicated.

Macrolide resistance among Streptococcus pneumoniae isolates has risen to prominence during the last decade (1–11). This situation is of particular concern as, in most cases, it is coupled with resistance to other first-line antibiotics (11). According to recent surveys, almost 40% of pneumococci in Spain are penicillin resistant. This resistance is frequently associated with resistance to macrolides, tetracyclines, and chloramphenicol (1, 12). Ketolides, a novel class of antibiotics (5), appear to be an alternative to macrolides for the treatment of pneumococcal infections in which multidrug-resistant strains are involved. A high intrinsic in vitro activity coupled with a favorable pharmacokinetic profile and the lack of inducibility properties make these compounds promising alternatives for the treatment of infections caused by respiratory pathogens (3, 4, 9).

In the present study, the in vitro activity of telithromycin was evaluated against clinical isolates of S. pneumoniae prospectively collected in different geographical areas of Spain and was compared with those of erythromycin A, clindamycin, penicillin G, quinupristin-dalfopristin, cefotaxime, and levofloxacin. The resistance mechanisms involved in erythromycin-intermediate and -resistant strains were phenotypically and genotypically characterized. The lack of inductive activity of telithromycin was also assessed.

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MATERIALS AND METHODS

Antibiotics. The following antibiotics were supplied as powders of known potency by the indicated manufacturers: telithromycin (HMR 3647), erythromycin A, penicillin G, cefotaxime, and levofloxacin, Aventis Pharma, Romainville, France; clindamycin, The Upjohn Co., Kalamazoo, Mich.; and quinupristin-dalfopristin, Rhône Poulenc Rorer, Paris, France. Telithromycin (15 µg), erythromycin (15 µg), clindamycin (2 µg), spiramycin (100 µg), chloramphenicol (30 µg), and tetracycline (30 µg) disks were purchased from Oxoid Ltd. (Basingstoke, United Kingdom).

Bacterial strains. A total of 203 clinical isolates of S. pneumoniae prospectively collected during 1999 and 2000 from 14 Spanish hospitals representing 14 different geographical areas were studied. Isolates obtained from clinical samples were distributed as follows: sputum (n = 69) and other samples from the lower respiratory tract (n = 48), blood (n = 43), eye (n = 16), nasopharynx (n = 12), middle ear (n = 7), catheter (n = 5), and organic fluids (n = 3).

Susceptibility testing. The MICs for the S. pneumoniae isolates were determined by an adaptation of the standard agar dilution test recommended for other organisms by the National Committee for Clinical Laboratory Standards (NCCLS) (18). Mueller-Hinton agar (Oxoid Ltd.) supplemented with 5% sheep blood was used, and the plates were incubated overnight in ambient air at 35°C. S. pneumoniae ATCC 49619 was included in each run as a control strain to ensure that the results were within the acceptable quality control limits of the NCCLS microdilution method for pneumococci (18). The MIC breakpoints recommended by NCCLS were considered for all antibiotics (18). In the case of telithromycin, the MIC breakpoint proposed by the manufacturer (Aventis Pharma) was applied (susceptible, ≤0.5 µg/ml; resistant, ≥4 µg/ml). This breakpoint has been validated by two committees in Europe, including MENSURA (Mesa Española de Normalización de las Sensibilidad y Resistencia a los Antibióticos) and CA-SFM (Comité de l’Antibiogramme de la Société Française de Microbiologie) (26).

Agar disk diffusion assays. All strains were screened for macrolide-lincosamide-streptogramin B (MLSb) resistance by the disk diffusion method on Mueller-Hinton agar supplemented with 5% sheep blood. The plates were inoculated
controls in the PCRs for the erm and 5 (PCR product of ca. 640 bp); for
9 AAC GGT ACT TAA ATT GTT TAC-3
Streptococcus pyogenes
TAC TAA AAG TGG-3
9 of the
Markers III and V; Roche Diagnostic GmbH, Mannheim, Germany). Detection
of the PCR products were estimated with DNA molecular weight markers (DNA
for 45 s. A final elongation step of 72°C for 10 min was included. Electrophoresis
itations were performed on a PTC-100 (MJ Research Inc., Watertown, Mass.)
Biosystems, Foster City, Calif.), and 10
of each primer,2Uo fAmpliTaq Gold DNA polymerase (Perkin-Elmer Applied
l PCR mixture contained 15 mM Tris-
Laboratories, Hercules, Calif.). The 25-
of the DNA preparation. Amplifica-
m
Tris-HCl, 50 mM KCl (pH 8.0), 2 mM MgCl2, 100 µM (each) deoxyribonucleotide, 2 pmol
each of primer, 2 U of AmpliTaq Gold DNA polymerase (Perkin-Elmer Applied
Biosystems, Foster City, Calif.), and 10 µl of the DNA preparation. Amplifica-
tions were performed on a PTC-100 (MJ Research Inc., Watertown, Mass.)
thermocycler; and the program was as follows: 94°C for 12 min and 35 cycles of
denaturation at 94°C for 1 min, annealing at 58°C for 45 s, and elongation at 72°C
for 45 s. A final elongation step of 72°C for 10 min was included. Electrophoresis
was carried out on 2% agarose gels stained with ethidium bromide, and the sizes
of the PCR products were estimated with DNA molecular weight markers (DNA
Markers III and V; Roche Diagnostic GmbH, Mannheim, Germany). Detection of the
erm(B) and mef(A) (formerly mefE) (23) genes was performed with the
InstaGene Matrix (Bio-Rad Laboratories, Hercules, Calif.). The 25-µl PCR mixture contained 15 mM Tris-
nock for clindamycin and spiramycin without blunting were assigned the M phenotype (cBlox). The inductive activity of telithromycin
was tested by replacing the erythromycin disk by the ketolide disk (3). Suscepti-
tibilities to chloramphenicol and tetracycline were also determined by the standard
agar diffusion test with the commercial disks cited above (19).

Detection of erythromycin resistance genes. Total DNA was obtained from all
erythromycin-intermediate and -resistant streptococcal strains and from five
susceptible isolates (negative controls) by using the InstaGene Matrix (Bio-Rad
Laboratories, Hercules, Calif.). The 25-µl PCR mixture contained 15 mM Tris-
HCl, 50 mM KCl (pH 8.0), 2 mM MgCl2, 100 µM (each) deoxyribonucleotide, 2 pmol
each of primer, 2 U of AmpliTaq Gold DNA polymerase (Perkin-Elmer Applied
Biosystems, Foster City, Calif.), and 10 µl of the DNA preparation. Amplification
was performed on a PTC-100 (MJ Research Inc., Watertown, Mass.)
thermocycler; and the program was as follows: 94°C for 12 min and 35 cycles of
denaturation at 94°C for 1 min, annealing at 58°C for 45 s, and elongation at 72°C
for 45 s. A final elongation step of 72°C for 10 min was included. Electrophoresis
was carried out on 2% agarose gels stained with ethidium bromide, and the sizes
of the PCR products were estimated with DNA molecular weight markers (DNA
Markers III and V; Roche Diagnostic GmbH, Mannheim, Germany). Detection of the
erm(B) and mef(A) (formerly mefE) (23) genes was performed with the
primers reported previously (27) (Amersham Pharmacia Biotech, Uppsala, Swe-
den): for erm(B), 5'-GAA AAG GTA CTC AAC AAT ATA-3' and 5'-AGT
AAC GGT ACT TAA ATT GTT TAC-3' (PCR product of ca. 640 bp); for
mef(A), 5'-AGT ATC ATT AAT CAC TAG TGC-3' and 5'-TTC TTC TCG
TAC TAA AAG TGG-3' (PCR product of ca. 350 bp). Genomic DNAs from
Streptococcus pyogenes AC1 and S. pyogenes 02C1064 were used as positive
controls in the PCRs for the erm(B) and mef(A) genes, respectively. A negative
control in which DNA was omitted was also included in each run.

RESULTS

Overall susceptibility and resistance rates. The results ob-
tained for S. pneumoniae ATCC 49619 were within acceptable
quality control limits of the NCCLS microdilution method for
pneumococci (18). The MICs, ranges of MICs, and percent
susceptibilities to each antibiotic for all S. pneumoniae isolates
tested are shown in Table 1.

Telithromycin and levofloxacin were the most active agents
among the antibiotics tested, with 98.9 and 99.6% of the strains
being susceptible to these two antibiotics, respectively. The
overall rate of penicillin susceptibility among the 203 S. pneu-
moniae isolates was 51.7%; 34.5% of the strains showed inter-
mediate resistance and 13.8% were fully resistant to this anti-

biotic. The corresponding values for cefotaxime were 94, 6.0,
and 0%, respectively. The overall rate of resistance (interme-
diate plus resistant isolates) to erythromycin was 39.0%. As has
been noted in worldwide studies of the susceptibilities of S.
pneumoniae isolates (11), erythromycin resistance was more
prevalent among penicillin-resistant (75.0%) and penicillin-
intermediate (62.8%) isolates than among penicillin-suscepti-
ble isolates (13.3%). The rates of clindamycin and quinupristin-
dalfopristin resistance were 32 and 30.1%, respectively.

According to the results of the disk diffusion tests, the rate of
resistance to tetracycline was 38.4% (n = 78), and in the case of
chloramphenicol, the rate of resistance was 21.7% (n = 44).
Tetracycline and chloramphenicol resistance was more fre-

TABLE 1. Comparative in vitro activities of telithromycin against 203 S. pneumoniae isolates

<table>
<thead>
<tr>
<th>Antimicrobial agent</th>
<th>MIC (µg/ml)a</th>
<th>% of fully susceptible isolates b</th>
</tr>
</thead>
<tbody>
<tr>
<td>Penicillin</td>
<td>≤0.008–4</td>
<td>94.0</td>
</tr>
<tr>
<td>Cefotaxime</td>
<td>≤0.004–1</td>
<td>94.0</td>
</tr>
<tr>
<td>Telithromycin</td>
<td>≤0.002–1</td>
<td>94.0</td>
</tr>
<tr>
<td>Erythromycin</td>
<td>≤0.008–&gt;64</td>
<td>94.0</td>
</tr>
<tr>
<td>Clindamycin</td>
<td>≤0.008–&gt;64</td>
<td>61.0</td>
</tr>
<tr>
<td>Quinupristin-dalfopristin</td>
<td>0.06–&gt;64</td>
<td>68.0</td>
</tr>
<tr>
<td>Levofloxacin</td>
<td>0.12–8</td>
<td>99.9</td>
</tr>
</tbody>
</table>

a 50% and 90%, MICs at which 50 and 90% of isolates are inhibited, respec-
tively. b NCCLS breakpoints (18).
results are consistent with the fact that the therapeutic activity of telithromycin is slightly affected by either the \textit{erm} (B)- or the \textit{mef} (A)-related erythromycin resistance mechanisms.

The classical inducible MLS\textsubscript{B} phenotype (erythromycin resistance and clindamycin susceptibility) and constitutive MLS\textsubscript{B} phenotype (high-level cross-resistance), first distinguished in \textit{Staphylococcus aureus}, are difficult to apply to \textit{S. pneumoniae}, as many strains with the constitutive phenotype are even inducible (24). Indeed, we found a wide range of erythromycin concentrations inhibitory for \textit{erm} (B)-positive \textit{S. pneumoniae} isolates (0.5 to 64 \(\mu\)g/ml), particularly among those isolates for which erythromycin MICs are lower (0.5 to 16 \(\mu\)g/ml), which may correspond to strains with different levels of inducibility. Because telithromycin is a weak inducer, most of these strains retain their susceptibilities to this drug. Moreover, the erythromycin, clindamycin, and spiramycin triple-disk induction test was not always positive for \textit{S. pneumoniae} isolates with the classical inducible MLS\textsubscript{B} phenotype according to the MICs for the isolates. In fact, only 11 of 74 \textit{erm} (B)-positive isolates (14.8\%) displayed an inducible phenotype by the disk induction test. Nevertheless, disk diffusion tests also confirmed that telithromycin has no inducing activity against erythromycin, clindamycin, and spiramycin for those strains for which, on the contrary, this profile was clearly detected for erythromycin. As expected, no inducing activity of erythromycin or telithromycin was observed in \textit{mef} (A)-positive isolates. Quinupristin-dalfopristin, which combines the activities of streptogramins A and B, which are synergistic, was active against 70\% of the strains tested; and significant levels of susceptibility to quinupristin-dalfopristin were retained even in the presence of the \textit{erm} (B) determinant. For 67.5\% of the \textit{erm} (B)-positive isolates, the quinupristin-dalfopristin MIC was \(\leq 1 \mu\)g/ml. The last group included isolates with the inducible phenotype (11 isolates) as well as isolates with the constitutive MLS\textsubscript{B} phenotype by the disk induction test (39 isolates).

### DISCUSSION

Multidrug resistance in \textit{S. pneumoniae} is well documented all over the world. As a consequence, current oral antibiotics are losing their efficacies for the treatment of infections caused by this organism (2, 15). Although the frequency of penicillin-intermediate and -resistant \textit{S. pneumoniae} isolates varies among countries, it appears to be increasing worldwide. Moreover, macrolide resistance is increasing among both penicillin-resistant and -susceptible isolates (1, 11, 12). In this scenario,

<table>
<thead>
<tr>
<th>MLS resistance genotype (no. of isolates) and antibiotic</th>
<th>MIC ((\mu)g/ml)\textsuperscript{a}</th>
<th>Range 50% 90%</th>
<th>Geometric mean</th>
</tr>
</thead>
<tbody>
<tr>
<td>\textit{erm} (B) and \textit{mef} (A) negative (125)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Penicillin</td>
<td>0.008–4</td>
<td>0.03</td>
<td>1</td>
</tr>
<tr>
<td>Cefotaxime</td>
<td>0.004–1</td>
<td>0.01</td>
<td>0.25</td>
</tr>
<tr>
<td>Telithromycin</td>
<td>0.002–0.06</td>
<td>0.01</td>
<td>0.06</td>
</tr>
<tr>
<td>Erythromycin</td>
<td>0.008–2</td>
<td>0.03</td>
<td>0.06</td>
</tr>
<tr>
<td>Clindamycin</td>
<td>0.008–0.12</td>
<td>0.03</td>
<td>0.06</td>
</tr>
<tr>
<td>Quinupristin-dalfopristin</td>
<td>0.06–4</td>
<td>1</td>
<td>2</td>
</tr>
<tr>
<td>Levofloxacin</td>
<td>0.12–1</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>\textit{erm} (B) positive and \textit{mef} (A) negative (73)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Penicillin</td>
<td>0.04–1</td>
<td>0.5</td>
<td>2</td>
</tr>
<tr>
<td>Cefotaxime</td>
<td>0.008–1</td>
<td>0.25</td>
<td>1</td>
</tr>
<tr>
<td>Telithromycin</td>
<td>0.004–1</td>
<td>0.03</td>
<td>0.12</td>
</tr>
<tr>
<td>Erythromycin</td>
<td>0.5–&gt;64</td>
<td>&gt;64</td>
<td>&gt;64</td>
</tr>
<tr>
<td>Clindamycin</td>
<td>0.06–&gt;64</td>
<td>64</td>
<td>&gt;64</td>
</tr>
<tr>
<td>Quinupristin-dalfopristin</td>
<td>0.25–8</td>
<td>1</td>
<td>2</td>
</tr>
<tr>
<td>Levofloxacin</td>
<td>0.25–8</td>
<td>0.5</td>
<td>1</td>
</tr>
<tr>
<td>\textit{erm} (B) negative and \textit{mef} (A) positive (4)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Penicillin</td>
<td>0.03–2</td>
<td>1</td>
<td>0.17</td>
</tr>
<tr>
<td>Cefotaxime</td>
<td>0.008–0.25</td>
<td>0.06</td>
<td>0.09</td>
</tr>
<tr>
<td>Telithromycin</td>
<td>0.01–0.5</td>
<td></td>
<td>2.82</td>
</tr>
<tr>
<td>Erythromycin</td>
<td>0.5–8</td>
<td></td>
<td>0.04</td>
</tr>
<tr>
<td>Clindamycin</td>
<td>0.03–0.06</td>
<td></td>
<td>1.68</td>
</tr>
<tr>
<td>Quinupristin-dalfopristin</td>
<td>1–2</td>
<td></td>
<td>0.84</td>
</tr>
<tr>
<td>Levofloxacin</td>
<td>0.25–2</td>
<td></td>
<td></td>
</tr>
<tr>
<td>\textit{erm} (B) and \textit{mef} (A) positive (1)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Penicillin</td>
<td>0.5</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cefotaxime</td>
<td>0.12</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Telithromycin</td>
<td>0.06</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Erythromycin</td>
<td>&gt;64</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Clindamycin</td>
<td>2</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Quinupristin-dalfopristin</td>
<td>1</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Levofloxacin</td>
<td>1</td>
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</table>

\textsuperscript{a} 50\% and 90\%, MICs at which 50 and 90\% of isolates are inhibited, respectively.
new antimicrobials, such as ketolides, have emerged. These drugs have specifically been designed to overcome MLS resistance mechanisms (5). Different studies have previously assayed the activities of telithromycin, a novel ketolide, against S. pneumoniae isolates (8, 14, 16, 17, 21, 22). However, the available information on the in vitro activities of telithromycin against a collection of S. pneumoniae isolates with well-characterized erythromycin resistance mechanisms remains scarce.

In our study, telithromycin displayed significant in vitro activity, with 98.9% of S. pneumoniae isolates tested being susceptible, regardless of the presence of macrolide resistance determinants. This result confirmed previous findings about the good in vitro activity of this ketolide against pneumococci (14, 16), even among resistant isolates. In our pneumococcal population, erm(B)-mediated methylation of the ribosomal target was the most prevalent resistance mechanism in erythromycin-intermediate and -resistant isolates (94.9%), with 36.4% isolates in the collection studied exhibiting the erm(B) determinant. In contrast, mef(A)-positive isolates accounted for only 2.4% of strains, with only one isolate carrying both mechanisms simultaneously (Table 2). Although the erm(B) determinant is always found among erythromycin-resistant isolates from different countries (16, 17, 20), our study revealed a higher incidence of this determinant in Spain than in other areas of the world, including Mediterranean countries and North America (25). This situation could be related to antibiotic consumption and differential selective pressures or to the clonal spread of both penicillin- and erythromycin-resistant strains (6, 10). Notably, erm(B) was detected more among penicillin-resistant (71.4%) and -intermediate (61.4%) isolates than among isolates that were part of the susceptible population (10.5%).

Descheemaeker et al. (8) recently found that 3 of 33 S. pneumoniae isolates displayed an M phenotype, with a range of telithromycin MICs of 0.125 to 0.5 μg/ml. A similar range was found for our four strains with a confirmed mef(A) resistance determinant, for which the telithromycin MICs placed them in the susceptible population (MIC range, 0.01 to 0.5 μg/ml). These results confirm that telithromycin is less affected than erythromycin by efflux-based mechanisms in S. pneumoniae, as erythromycin MICs were 0.5 to 8 μg/ml for isolates with the M phenotype or the mef(A) determinant.

As stated earlier, one isolate for which the erythromycin MIC was 2 μg/ml was negative for the erm(B) and mef(A) genes. Although it has been established that macrolide-resistant pneumococci that are isolated from clinical specimens and that do not carry either the erm or the mef gene occur frequently, ribosome and/or ribosomal protein mutations that are the same as or similar to those observed in recent in vitro mutants may be present in this isolate (28, 29; A. Canu, B.

**FIG. 1.** Correlation of MICs of erythromycin, telithromycin, clindamycin, and quinupristin-dalfopristin with presence of macrolide resistance determinants [□, no determinants; ■, erm(B) alone; □, mef(A) alone; ▲, erm(B) and mef(A)] for 203 S. pneumoniae isolates.
Telithromycin had a higher level of intrinsic activity than erythromycin against susceptible strains and was slightly affected by the erm(B)-mediated erythromycin resistance mechanism. Despite the small number of strains with the mef(A) resistance mechanism, telithromycin displayed higher levels of in vitro activity than erythromycin. This excellent in vitro activity, together with a lesser capacity to select resistant mutants compared to the capacities of other MLS agents (28) and the lack of inductive activity (3), merits further clinical studies on the efficacy of telithromycin against infections caused by *S. pneumoniae* when macrolides are indicated.

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**REFERENCES**


