Lack of Vancomycin Tolerance in *Streptococcus pneumoniae* Strains Isolated in Barcelona, Spain, from 1999 to 2001

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To evaluate the incidence of vancomycin tolerance among *Streptococcus pneumoniae* isolates, we performed killing curve studies with 633 isolates. The penicillin MIC was ≥0.12 mg/liter for 481 (76%) of the isolates. All strains were susceptible to vancomycin. Killing curve studies were performed with a vancomycin concentration of 2.5 mg/liter. The Tupelo strain was used for quality control. No vancomycin-tolerant strain was detected.

*Streptococcus pneumoniae* is the most common bacterial cause of acute meningitis and pneumonia, for which mortality rates among treated patients are 30 and 5%, respectively (9, 10). Resistance to β-lactams, such as penicillin, represents a major problem in the treatment of pneumococcal infections. Penicillin resistance has been found among up to 50% of clinical isolates, according to reports from many parts of the world (2). In addition, an increasing amount of multidrug resistance makes the treatment of these infections even more difficult. No resistance of the pneumococcus to the glycopeptide vancomycin has yet been reported, so this drug represents the ultimate backup drug for the treatment of infections caused by multidrug-resistant pneumococci (3).

Antibiotic tolerance is the ability of bacteria to survive but not proliferate in the presence of a particular bactericidal antibiotic. This translates to increased bacterial survival and regrowth when the antibiotic is removed. Tolerance to penicillin was first recognized among clinical isolates of pneumococci 15 years ago, when it was demonstrated that five of six multidrug-resistant clinical isolates of pneumococci were also tolerant to β-lactam antibiotics (5). Recently, vancomycin-tolerant *S. pneumoniae* strains have been described in the laboratory (8) and in a patient with recurrent meningitis (6). This problem of tolerance to vancomycin is especially significant in those countries where the incidence of penicillin resistance among pneumococci is high, necessitating inclusion of vancomycin in the initial empirical treatment of meningitis. Tolerance to vancomycin is not detected by routine in vitro susceptibility tests, so specific studies are needed to evaluate the incidence of vancomycin tolerance among pneumococcal isolates in our region.

(This study was presented at the 42nd Interscience Conference on Antimicrobial Agents and Chemotherapy [M. Ortega et al., Abstr. 42nd Intersci. Conf. Antimicrob. Agents Chemother., abstr. E-1648, 2002].)

From January 1999 to December 2001 a total of 1,033 *S. pneumoniae* strains were isolated from different samples submitted to the microbiology laboratory of our institution. The MICs for all strains were determined by a commercial microdilution method (Sensititre; Trek Diagnostic Systems, West Sussex, United Kingdom) with cation-adjusted Mueller-Hinton broth supplemented with 5% lysed sheep blood. The recommendations of NCCLS (7) were followed for classification of the strains as susceptible, intermediate, or resistant to the antimicrobial agents tested, including penicillin, cefotaxime, erythromycin, levofloxacin, and vancomycin. Quality control was assured by use of the following strains: *S. pneumoniae* ATCC 49619, *Staphylococcus aureus* ATCC 29213, and *Enterococcus faecalis* ATCC 29212. All strains were stored at −70°C in double-strength skim milk until needed.

Tolerance to vancomycin was studied by a killing curve method with Todd-Hewitt broth plus 0.5% yeast extract as the culture medium. Vancomycin was considered bactericidal when a ≥3-log_{10} reduction in colony counts was reached at 6 h; otherwise, the strain was considered tolerant (6). The strains tested were inoculated (final concentration, 10^7 to 10^8 CFU/ml) in 10 ml of broth without antibiotic (positive growth) or supplemented with 2.5 mg of vancomycin per liter. The test tubes were incubated at 35°C, and the numbers of CFU per milliliter were determined after 0 and 6 h. All tests were performed in duplicate. Strains that showed ≥10^4 CFU/ml after 6 h of incubation were reanalyzed, and the numbers of CFU per milliliter were determined after 0, 2, 4, and 6 h. The Tupelo strain (a vancomycin-tolerant strain), a gift from J. A. McCullers (6), was used for quality control.

Among the 1,033 *S. pneumoniae* strains isolated, 633 were selected for use in the killing curve studies. All strains (n = 233) recovered from sterile fluids (blood, cerebrospinal fluid) were included. Among the other isolates, only those for which the penicillin MIC was ≥0.12 mg/liter (n = 400) were analyzed. The strains were classified according to the penicillin MICs as follows: 152 strains were susceptible (MICs, ≤0.06 mg/liter), 270 showed intermediate susceptibilities (MICs, 0.12 to 1 mg/liter), and 211 were resistant (MICs, ≥2 mg/liter). Table 1 shows the overall rates of resistance to penicillin, cefotaxime, erythromycin, and levofloxacin over the 3-year study period. All strains were susceptible to vancomycin. For analysis pur-
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TABLE 1. Incidence of strains with reduced susceptibilities to different antimicrobial agents during the 3 years of the study

<table>
<thead>
<tr>
<th>Yr/total no. of isolates</th>
<th>Penicillin MIC, ≤0.12 μg/ml</th>
<th>Cefotaxime MIC, =1 μg/ml</th>
<th>Erythromycin MIC, ≤0.5 μg/ml</th>
<th>Levofloxacin MIC, ≤4 μg/ml</th>
</tr>
</thead>
<tbody>
<tr>
<td>1999/361</td>
<td>170 (47)</td>
<td>62 (17)</td>
<td>130 (36)</td>
<td>7 (2)</td>
</tr>
<tr>
<td>2000/380</td>
<td>178 (46)</td>
<td>61 (16)</td>
<td>155 (41)</td>
<td>16 (4)</td>
</tr>
<tr>
<td>2001/292</td>
<td>133 (45)</td>
<td>53 (18)</td>
<td>115 (39)</td>
<td>6 (2)</td>
</tr>
</tbody>
</table>

The tolerance of strains to lytic antibiotics is caused by a defective death response that is due either to a defective autolysin or to a defect in the signal transduction cascade that activates the autolysin. Vancomycin is an important drug in the management of multidrug-resistant pneumococci. No resistance of pneumococci to this drug has yet been reported. However, other groups have reported that the incidence of vancomycin tolerance among laboratory isolates is about 3 to 5% (C. A. Rodriguez, C. G. Whitney, and E. Tuomanen, Abstr. 40th Intersci. Conf. Antimicrob. Agents Chemother., abstr. 1777, 2000), and recently a case of clinical failure following vancomycin treatment of meningitis caused by a vancomycin-tolerant isolate (the Tupelo strain) was reported (6).

Henriques et al. (4) determined the prevalence of vancomycin and penicillin tolerance among 116 clinical isolates of pneumococci by monitoring lysis and viability after exposure to each antibiotic for 4 h. Eight percent of the strains were tolerant to penicillin, and 3% were tolerant to vancomycin. The three vancomycin-tolerant strains had reduced susceptibilities to penicillin, and only one was also tolerant to penicillin.

A group from Madrid studied 120 strains of S. pneumoniae isolated in 1999 (1). Although they did not indicate the susceptibilities of the pneumococci in the paper, the final result was that any strain studied was vancomycin tolerant. In our study we included a larger number of strains (n = 633) and strains with a high level of resistance (penicillin MIC, ≥0.12 mg/liter for 481 strains [76%]) because vancomycin tolerance had been detected among penicillin-resistant strains and empirical treatment with vancomycin is needed for meningitis. However, vancomycin tolerance was not detected in any of the strains studied.

Although large numbers of pneumococcal isolates in Spain have reduced susceptibilities to penicillin (intermediate or resistant), strains tolerant to vancomycin, which may compromise the ability to use this antibiotic in the treatment of invasive pneumococcal infections, have not yet been found. Nevertheless, it would be advisable to carry out this type of epidemiological study in order to detect the possible appearance of this sort of strain.

M.O. and F.M. were responsible for the microbiological study. A.S. and E.G. aided with the performance of the statistical analysis. M.O. and F.M. wrote the manuscript. J.A.M. and J.M. carried out the critical review of the paper.

REFERENCES


