Pharmacokinetics and Metabolism of $[^{14}C]$Viramidine in Rats and Cynomolagus Monkeys

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Ribavirin (1-β-d-ribofuranosyl-1,2,4-triazole-3-carboxamide) is a purine nucleoside analog with activity against a variety of DNA and RNA viral infections (10, 11). The clinical efficacies of interferon alfa-2b and pegylated interferon alfa-2b in combination with ribavirin are about 40% (6, 9) and 54% (5), respectively, in terms of a sustained virologic response when they are used to treat chronic hepatitis C virus infection. At present, combination therapy with ribavirin and pegylated interferon alfa-2b is the “gold standard” for the treatment of chronic hepatitis C. However, ribavirin has a dose-limiting side effect, hemolytic anemia. After absorption into the circulation, a significant portion of ribavirin is transported into red blood cells (RBCs) and accumulate over time, leading to hemolytic anemia (1, 3). This side effect is dose limiting and necessitates dose reduction and withdrawal in some patients. A new ribavirin which retains those properties deemed critical in the treatment of chronic hepatitis C, but with less potential for hemolytic anemia, would be highly desirable.

Viramidine is a prodrug of ribavirin. Preliminary studies in our laboratories indicated that viramidine can be retained and converted to ribavirin in the liver (J. Lau et al., Abstr. 52nd Am. Assoc. Study Liver Dis., abstr. 1020, 2001). Following daily oral dosing (30 mg/kg of body weight) and monkeys (10 mg/kg) following intravenous (i.v.) and oral administration. The levels of oral absorption and bioavailabilities were 61.7 and 9.91%, respectively, in rats and 43.9 and 13.6%, respectively, in monkeys. Following i.v. administration, the elimination half-lives were 2.7 h in rats and 28.9 h in monkeys. Total body clearances were 14.0 liters/h/kg in rats and 1.23 liters/h/kg in monkeys; the apparent volumes of distribution were 15.6 liters/kg in rats and 18.6 liters/kg in monkeys. Following oral administration, viramidine was extensively converted to ribavirin, followed by further metabolism of ribavirin in both species, with a faster rate of metabolism in rats than in monkeys. In rats, excretion of total radioactivity in urine accounted for 77.0% of the i.v. dose and 60.8% of the oral dose, while in monkeys it accounted for 44.4% of the i.v. dose and 39.0% of the oral dose. The amount of unchanged viramidine and ribavirin in urine was small in both species after i.v. and oral administration of viramidine.

The pharmacokinetics of $[^{14}C]$viramidine, a prodrug of ribavirin, were studied in rats (30 mg/kg of body weight) and monkeys (10 mg/kg) following intravenous (i.v.) and oral administration. The levels of oral absorption and bioavailabilities were 61.7 and 9.91%, respectively, in rats and 43.9 and 13.6%, respectively, in monkeys. Following i.v. administration, the elimination half-lives were 2.7 h in rats and 28.9 h in monkeys. Total body clearances were 14.0 liters/h/kg in rats and 1.23 liters/h/kg in monkeys; the apparent volumes of distribution were 15.6 liters/kg in rats and 18.6 liters/kg in monkeys. Following oral administration, viramidine was extensively converted to ribavirin, followed by further metabolism of ribavirin in both species, with a faster rate of metabolism in rats than in monkeys. In rats, excretion of total radioactivity in urine accounted for 77.0% of the i.v. dose and 60.8% of the oral dose, while in monkeys it accounted for 44.4% of the i.v. dose and 39.0% of the oral dose. The amount of unchanged viramidine and ribavirin in urine was small in both species after i.v. and oral administration of viramidine.

MATERIALS AND METHODS

Compound. The compound [5-$^{14}$C]viramidine (Fig. 1) was synthesized by using [14C]barium carbonate as a precursor. The labeled nucleoside was extensively purified by column chromatography and repetitive recrystallization. The chemical identity and purity were verified by mass spectrometry and proton magnetic resonance spectrometry. The radiopurity (>98%) of the preparation was confirmed by high-pressure liquid chromatography (HPLC) coupled with radiolabel detection. Ribavirin, triazole carboxamide (TCONH$_2$), and triazole carboxylic acid nucleoside (RTC(OH)$_2$) were obtained from Ribapharm, Inc.

Drug administration and sample collection for rats. Following an overnight fast, six male Sprague-Dawley rats received an intravenous (i.v.) dose of 30 mg of $[^{14}C]$viramidine per kg via a tail vein and six rats received an oral dose of 30 mg of $[^{14}C]$viramidine (75 µCi) per kg via oral gavage. Serial blood samples from three rats receiving drug by each dose route were collected directly in heparinized Vacutainer tubes and immediately centrifuged to harvest the plasma. Urine and fecal samples from three rats receiving drug by each dose route were collected for analysis.

Drug administration and sample collection for cynomolgus monkeys. Following an overnight fast, four male cynomolgus monkeys received an i.v. bolus dose of 10 mg of $[^{14}C]$viramidine per kg via a saphenous vein or an oral dose of 10 mg of $[^{14}C]$viramidine (0.5 mCi) per kg via oral gavage. Serial blood samples from each monkey were collected directly in heparinized Vacutainer tubes and immediately centrifuged to harvest the plasma. Urine and fecal samples were collected from each monkey for analysis.

Measurement of radioactivity. The levels of radioactivity in plasma (0.5 ml) and urine (0.2 ml) were measured by using Ultima Gold XR scintillation cocktail and a liquid scintillation counter (model 1900TR; Packard Instrument Company, Meriden, Conn.). Fecal samples were combusted in a sample oxidizer (model 306; Packard Instrument Company), and the resulting $^{14}$CO$_2$ was trapped in a mixture of Perma Fluor E and Carbo-Sorb E, followed by liquid scintillation counting. Scintillation counting data were automatically corrected for counting efficiency, based on an external standard, and an instrument-stored quench curve generated from a series of sealed quenched standards.

LC-MS-MS method for determination of viramidine and ribavirin in plasma. The analytical liquid chromatography (LC)-mass spectrometry (MS)-MS method involved the addition of an internal standard (acyclovir), protein precipitation with acetonitrile, solvent evaporation, reconstitution of residue, and separation...
on an Inertsil Silica column, followed by MS-MS detection. A Perkin-Elmer Sciex API 3000 instrument in the multiple-reaction monitoring mode with positive electrospray ionization was used to monitor the transitions from $m/z$ 245 to 113 and 259 to 128 for ribavirin and the internal standard.

For viramidine, the limit of quantitation was 10 ng/ml, with a coefficient of variation (CV) of 9% and a bias of 1.3%. Linear regression of the concentration data (range, 10 to 5,000 ng/ml) yielded a correlation coefficient of $R^2 = 0.999$. The LC-MS-MS method was accurate (bias, $<6\%$) and reproducible (CV, $<8.2\%$).

For ribavirin, the limit of quantitation was 10 ng/ml, with a CV of 8.5% and a bias of 1.8%. Linear regression of the concentration data (range, 10 to 5,000 ng/ml) yielded a correlation coefficient of $R^2 = 0.999$. The LC-MS-MS method was accurate (bias, $<5\%$) and reproducible (CV, $<8.0\%$).

**HPLC procedure for studying the metabolic profiles of the drug in plasma and urine.** Plasma and urine were mixed with an equal volume of acetonitrile. The mixtures were centrifuged, and the supernatant was injected into the HPLC apparatus coupled with a radioactivity detector (β-Ram model 2; INU Systems, Inc., Tampa, Fla.). The high-pressure liquid chromatograph (model SCL 10VP; Shimadzu) was equipped with an Amide-80 column (4.6 mm by 110 cm; Tosohaas). The column was eluted with a solvent mixture consisting of 95% of an organic mobile phase (acetonitrile) and 10% of an aqueous phase (25 mM ammonium acetate) at a flow rate of 1.2 ml/min. Immediately after the injection, the solvent mixture was switched to 70% organic phase and 30% aqueous phase. Identification of radioactive peaks in rat and monkey plasma and urine as ribavirin, TCONH$_2$, and RTCOOH was based on the retention times and LC-MS-MS of authentic standards.

**Pharmacokinetic analysis.** The concentrations of radioactivity, viramidine, and ribavirin in plasma were used to determine the values of the pharmacokinetic parameters by noncompartmental methods (WinNonlin-2; Pharsight Corp., Mountain View, Calif.). The maximum concentration in plasma ($C_{\text{max}}$) and the time to $C_{\text{max}}$ ($T_{\text{max}}$) were observed values. The area under the concentration-time curve (AUC) to the last quantifiable sampling time ($t_f$), $AUC_{\text{t_f}}$, was computed by using the linear trapezoidal rule. The AUC to infinity, $AUC_{\infty}$, was calculated as the sum of $AUC_{t_f}$ and the quotient of the last measurable concentration ($C_{t_f}$) and the elimination rate constant ($K$). $K$ was estimated as the negative slope of the regression of the log concentration versus time. The half-life ($t_{1/2}$) was calculated by dividing 0.693 by $K$. The apparent total body clearance (CL) was calculated as the ratio of the dose to $AUC_{\infty}$. The volume of distribution (V) was calculated as the ratio of CL to $K$.

**RESULTS**

**Concentrations of viramidine in rat plasma.** Following i.v. administration of viramidine, plasma viramidine levels decreased with time (Fig. 2), with an elimination $t_{1/2}$ of 2.7 h. The mean $V$ was 15.6 liters/kg, and the mean CL was 14.0 liters/h/kg. Following oral dosing of viramidine, viramidine was rapidly absorbed, with a $T_{\text{max}}$ of 0.50 h and a $C_{\text{max}}$ of 0.10 μg/ml. By comparison of the $AUC_{\infty}$ of viramidine obtained after oral dosing to that obtained after i.v. dosing, the absolute bioavailability of viramidine was calculated to be 9.91% (Table 1).

**Concentrations of ribavirin in rat plasma.** After i.v. administration of viramidine, plasma ribavirin levels decreased with time (Fig. 2), with an elimination $t_{1/2}$ of 6.6 h. After oral dosing of viramidine, plasma ribavirin levels reached a maximum at 4 h, with a $C_{\text{max}}$ of 0.362 μg/ml. Thereafter, plasma ribavirin levels decreased with time, with an elimination $t_{1/2}$ of 5.0 h, which is longer than that of viramidine after either i.v. dosing (2.7 h) or oral dosing (1.4 h) of viramidine. The ribavirin...
AUC$_{Rs}$ were 5.33 μg·h/ml after i.v. dosing of viramidine and 3.39 μg·h/ml after oral dosing of viramidine (Table 1).

Radioactivity levels in rat plasma. Following i.v. administration of [14C]viramidine, the plasma radioactivity level declined, with a $t_{1/2}$ of approximately 10.4 h. The AUC$_{Rs}$ for radioactivity in plasma were 43.5 μg equivalents·h/ml after oral dosing and 32.6 μg equivalents·h/ml after i.v. dosing (Table 1).

Concentrations of viramidine in monkey plasma. Following i.v. administration of viramidine, plasma viramidine levels decreased with time (Fig. 2), with an elimination $t_{1/2}$ of 28.9 h. The mean $V$ was 18.6 liters/kg, and the mean CL was 1.23 liters/h/kg. After oral administration of viramidine, viramidine was rapidly absorbed, with a $T_{max}$ of 1 h and a $C_{max}$ of 0.156 μg/ml. By comparison of the viramidine AUC$_f$ obtained after oral dosing to that obtained after i.v. dosing, the absolute bioavailability of viramidine was calculated to be 13.6% (Table 1).

Concentrations of ribavirin in monkey plasma. Following i.v. administration of viramidine, plasma ribavirin levels decreased with time (Fig. 2), with an elimination $t_{1/2}$ of 80.9 h. Following oral dosing of viramidine, plasma ribavirin levels reached a maximum at 3.0 h, with a $C_{max}$ of 0.089 μg/ml. Thereafter, plasma ribavirin levels declined with time, with an elimination $t_{1/2}$ of 62.2 h. The ribavirin AUC$_{Rs}$ were 9.68 μg·h/ml after i.v. administration of viramidine and 4.62 μg·h/ml after oral dosing of viramidine.

Radioactivity levels in monkey plasma. Following i.v. administration of [14C]viramidine, the plasma radioactivity levels declined, with a $t_{1/2}$ of 88.7 h. The AUC$_{Rs}$ for radioactivity in plasma were 25.0 μg equivalents·h/ml after oral administration and 28.7 μg equivalents·h/ml after i.v. administration.

Urinary and fecal excretion of radioactivity in rats and monkeys. In rats, 77.0% of the i.v. dose and 60.8% of the oral dose were excreted in urine (0 to 96 h). A total of 1.89% of the i.v. dose and 33.5% of the oral dose were excreted in rat feces (Table 1). These data demonstrate that biliary excretion does not play a significant role in the elimination of viramidine in rats (Table 1). On the basis of urinary excretion and the total recovery of radioactivity after oral dosing in rats, the level of absorption was estimated to be 61.7%.

In monkeys, 44.4% of the i.v. dose and 39.0% of the oral dose were excreted in urine (0 to 168 h). A total of 0.82% of the i.v. dose and 42.1% of the oral dose were excreted in monkey feces. The data indicate that biliary excretion does not play a significant role in the elimination of viramidine in monkeys. On the basis of urinary excretion and the total recovery of radioactivity after oral dosing in monkeys, the level of absorption was estimated to be 43.9%.

**DISCUSSION**

In rats following oral administration of viramidine, the AUC$_f$ of ribavirin (3.39 μg·h/ml) was 15 times the AUC$_f$ of viramidine (0.23 μg·h/ml). Similarly, in monkeys following oral administration of viramidine, the AUC$_f$ of ribavirin (4.62 μg·h/ml) was four times the AUC$_f$ of viramidine (1.14 μg·h/ml). Furthermore, following oral administration of viramidine, the ratios of the viramidine AUC$_f$ to the radioactivity AUC$_f$ were 0.005 for rats and 0.040 for monkeys, whereas the ratios of the ribavirin AUC$_f$ to the radioactivity AUC$_f$ were 0.075 for rats and 0.136 for monkeys. These data suggest that in both rats and monkeys viramidine is orally absorbed and rapidly converted to ribavirin, followed by further metabolism to RTOOH and TCONH$_2$. Ribavirin has been reported to
undergo hydrolytic deamination in vivo and is converted to RTCOOH, a metabolite that does not possess any antiviral activity (4, 7). Recently, Wu et al. (12) reported that the conversion of viramidine to ribavirin was catalyzed by adenosine deaminase. No deamination was observed in the absence of the deaminase or in the presence of 1 nM deoxycoformycin (Calbiochem, La Jolla, Calif.).

It is noteworthy that both the rate of conversion of viramidine to ribavirin and the rate of conversion of ribavirin to other metabolites were much higher in rats than in monkeys. These observations are in good agreement with the findings that the $t_{1/2}$ of viramidine after i.v. administration in rats (2.7 h) was shorter than that in monkeys (28.9 h) and that the CL of viramidine in rats (14.0 liters/min/kg) was much higher than that in monkeys (1.23 liters/min/kg). These findings are also in good agreement with the observation that following oral dosing of $^{14}$C]viramidine, viramidine was the only major radioactive peak in monkey urine collected over 0 to 24 h, whereas

### TABLE 1—Continued

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<td>$C_{\text{max}}$ (mg/liter)</td>
<td>$T_{\text{max}}$ (h)</td>
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**FIG. 3.** Metabolic profile of $^{14}$C]viramidine in rat and monkey plasma and urine following i.v. (IV) and oral (PO) dosing of viramidine.
virmidine, TCONH₂, ribavirin, and RTCOOH were all major radioactive peaks in rat urine collected over 0 to 24 h. The differences in conversion may be related to the presence of higher levels of deaminase and hydroxylase activity in rats compared to those in monkeys.

On the basis of the CL of viramidine for rats and monkeys, allometric scaling (log CL versus log body weight) was used to estimate the CL of viramidine in humans, which was estimated to be about 0.15 liter/h/kg. Given the phylogenetic proximity of monkeys and humans, it may be anticipated that the pharmacokinetic profile and metabolism of viramidine in humans are likely to be more similar to those in monkeys than to those in rats. If this were the case, viramidine would be converted to ribavirin at a rate that will allow the conversion of ribavirin to its phosphorylated metabolites, which are then retained in the liver. A too rapid conversion of viramidine to ribavirin could saturate the phosphorylation processes and favor the redistribution of the converted ribavirin to RBCs.

The $t_{1/2}$ of ribavirin were previously found to be 9.9 h in rats and 130 h in monkeys following i.v. dosing of ribavirin (4). In this study, the $t_{1/2}$ of ribavirin after i.v. dosing of viramidine were 6.6 h in rats and 80.9 h in monkeys. These $t_{1/2}$ were slightly shorter than the viramidine $t_{1/2}$ after i.v. dosing of viramidine in rats (2.7 h) and monkeys (28.9 h), since viramidine can be eliminated from the systemic circulation by urinary excretion as unchanged drug, in addition to its conversion to ribavirin. These data indicate that rats had higher rates of conversion of viramidine to ribavirin than monkeys and higher rates of metabolism of ribavirin to other metabolites.

Percent absorption can be estimated from the percentage of radioactivity excreted in urine after oral dosing, corrected for total recovery. On the basis of this approach, the levels of absorption of viramidine were estimated to be 61.7% in rats and 43.9% in monkeys, which were slightly lower than the levels of absorption of ribavirin in rats (80.9%) and monkeys (79.1%) determined previously (4). By comparing the plasma viramidine $AUC_t$ obtained after oral dosing of viramidine to that obtained after i.v. administration of viramidine, the bioavailability of viramidine were estimated to be 9.91% in rats and 13.6% in monkeys. This low bioavailability is expected, since viramidine is a ribavirin prodrug and is rapidly converted to ribavirin after oral dosing.

Despite the lower level of absorption of viramidine compared to that of ribavirin, the plasma ribavirin $AUC_t$ after oral dosing of viramidine in rats (3.39 µg·h/ml) was similar to or slightly higher than the plasma ribavirin $AUC_t$ after oral dosing of ribavirin (3.04 µg·h/ml) detected previously (4). Again, this is in good agreement with the rapid conversion of viramidine to ribavirin in rats. In monkeys, however, the plasma ribavirin $AUC_t$ following oral viramidine dosing (4.62 µg·h/ml) was lower than the plasma ribavirin $AUC_t$ following oral ribavirin dosing (13.1 µg·h/ml) detected previously (4). This is probably related to either the lower level of absorption of viramidine compared to that of ribavirin in monkeys and/or the slower rate of conversion of viramidine to ribavirin.

Quantitative whole-body autoradiography has been used to evaluate the tissue drug distribution in rats following oral dosing of viramidine and ribavirin (C. Lin et al., Abstr. 52nd AASLD, abstr. 1021, 2001). The highest tissue drug level was found in the liver after either viramidine or ribavirin dosing. However, viramidine dosing yielded liver drug concentrations 44% higher than those obtained after ribavirin dosing. The distribution in tissue has also been evaluated in monkeys at 24 h after administration of the 10th oral daily dose (10 mg/kg) of [14C]viramidine or [14C]ribavirin (Lin et al., Abstr. 52nd AASLD, abstr. 1123, 2001). The highest level of radioactivity was found in the liver after oral dosing of viramidine. Ribavirin dosing gave a distribution in tissue similar to that after viramidine dosing, except that viramidine was distributed to the liver at higher levels than ribavirin and to RBCs, the eyes, and fat at lower levels than ribavirin. Although viramidine dosing gave higher drug levels in the monkey liver than ribavirin dosing, viramidine showed a much better safety profile than ribavirin, probably due to the lower plasma and RBC drug levels after viramidine dosing. A 28-day toxicity study was conducted with rats and oral doses of 0, 30, 60, and 120 mg/kg and with monkeys and oral doses of 0, 100, 300, and 600 mg/kg. Ribavirin at 300 mg/kg/day induced significant hematological changes in monkeys, whereas viramidine at 600 mg/kg/day had only a slight effect on hematological parameters (C. Lin et al., Abstr. 53rd AASLD, abstr. 508, 2002).

We have previously demonstrated (Lin et al., Abstr. 52nd AASLD, abstr. 1123, 2001) that viramidine dosing in monkeys gave liver drug levels about three times higher than those achieved after ribavirin dosing but 50% lower RBC and plasma drug levels. Recently, Aroa et al. (S. Aroa et al., Abstr. 53rd AASLD, abstr. 773, 2002) reported that with an oral dose of 600 mg in humans, viramidine dosing gave about 50% lower plasma and RBC ribavirin levels than ribavirin dosing. Although liver drug levels after either viramidine or ribavirin dosing were not yet available, it appears that the monkey, not the rat, is a good model for prediction of the pharmacokinetics and tissue distribution of viramidine in humans.
REFERENCES


