Pharmacokinetics of Tigecycline after Single and Multiple Doses in Healthy Subjects

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Tigecycline, a novel glycylcycline antibiotic, exhibits strong activity against gram-positive, gram-negative, aerobic, anaerobic, and atypical bacterial species, including antibiotic-resistant strains. In studies with clinical isolates, tigecycline exhibits activity against tetracycline-resistant bacteria such as methicillin-susceptible Staphylococcus aureus, methicillin-resistant S. aureus, and glycopeptide-intermediate S. aureus. Penicillin-susceptible and -resistant Streptococcus pneumoniae (10) and vancomycin-resistant enterococci are also susceptible to tigecycline. In addition, tigecycline is active against most gram-negative pathogens, including Enterobacteriaceae, Acinetobacter spp., Stenotrophomonas maltophilia (1, 9, 13), Haemophilus influenzae, and Neisseria gonorrhoeae (5). Tigecycline’s expanded broad-spectrum activity is further evidenced by its activity against Legionella pneumophila (6), Chlamydia (20), rapidly growing nontuberculosis mycobacteria (25), and anaerobes (18). A few reports on the pharmacokinetics of tigecycline in animals are documented in the literature. After administration of 14C-labeled tigecycline to rats, tigecycline tissue levels, with the highest concentrations in bone, liver, spleen, and kidney, exceeded those in plasma and persisted longer (23). In the same study, tigecycline exhibited a long terminal-phase disposition half-life (t1/2) of 36 h in plasma, 208 h in bone, 128 h in thyroid, and 77 h in kidney (23). Tigecycline mean serum t1/2 in a rabbit model of enterococcal endocarditis ranged from 3.3 to 3.6 h (11). Pharmacokinetic data obtained by using a murine thigh infection model in neutropenic mice receiving tigecycline doses in the range of 3 to 48 mg/kg demonstrated a dose-dependent t1/2 in the range 1.05 to 2.34 h, a peak concentration in serum (Cmax) in the range 0.42 to 11.1 μg/ml, and a serum protein binding of ca. 60% (24).

The ascending single-dose study in healthy subjects demonstrated that after ascending single doses of tigecycline up to 300 mg, the tigecycline Cmax and area under the serum concentration-time curve (AUC) increased in a dose-related manner. The pharmacokinetics of tigecycline after single ascending doses of 25 to 150 mg in Japanese men were similar to those seen in the study involving a predominantly Caucasian population (21). Nearly 15% of tigecycline was excreted in urine as unchanged drug.

In an open-label, single-dose study, in which tigecycline was administered to men and women belonging to various age groups, mean Cmax and AUC values, which were between 0.9 to 1.1 μg/ml and 4.2 to 5.5 μg h/ml, respectively, were similar across ages and sex (14). As seen in the animal studies, tigecycline was extensively distributed into the tissues (Vss was 5.6 to 6.1 liters/kg in women and 5.5 to 7.1 liters/kg in men).

Full studies on tigecycline clinical safety and pharmacokinetics in humans have not been presented to date. This report describes the safety, tolerability, and pharmacokinetics of tigecycline after administration of single and multiple ascending intravenous (i.v.) doses in healthy subjects. In addition, the safety and tolerability of tigecycline given by various volumes and infusion rates in healthy subjects are also documented.

(The ascending single-dose study was presented previously [G. Muralidharan, J. Getsy, P. Mayer, et al., poster at the 39th Annual Interscience Conference on Antimicrobial Agents and Chemotherapy, 26 to 29 September 1999].)
In the VVIR study, tigecycline concentrations in serum were quantified by a high-performance liquid chromatography method with UV detection (350 nm) with a lower limit of detection of 2 µg/ml. Briefly, the method consisted of protein precipitation of the serum samples by addition of acetonitrile, separation of tigecycline on a Supelco LC-18 DB column (150 mm x 4.6 mm, internal diameter), 3-µm particle size) and subsequent UV detection of tigecycline at 350 nm. The extraction efficiency was higher than 76%, and the lower limit of quantitation was 25 ng/ml. The intrabatch and interbatch coefficients of variation (CV) were <13% at all concentrations.

Tigecycline in urine was quantified in the SAD study by a high-pressure liquid chromatography method with UV detection (350 nm) with a lower limit of detection of 2 µg/ml. Briefly, the urine samples were diluted to 1:10, and tigecycline was chromatographed on a C18 column (150 mm x 4.6 mm inner diameter), 3-µm particle size). The intraday CV and bias for the low quality control (QC) samples (6 µg/ml) were 4.6 and −1.5%, respectively. The interday CV and bias values at 6 µg/ml were 7.0 and −1.5%, respectively. The interday (n = 25) precision (CV) values for the middle QC (12 and 36 µg/ml) and the high QC (72 µg/ml) samples were between 5.1 and 9.0%, and the interday bias ranged from −7.9 to −0.6%. The precision (CV) and accuracy (expressed as bias) of tigecycline calibration standards in the lower-tier curve were between 1 and 3.5% and 3.0% and between 2.5 and 3.0%, respectively, to separate cohorts of subjects. Because of the hypothesized relationship between serotonin release and nausea observed after tigecycline administration, urinary excretion of the serotonin metabolite 5-hydroxyindoleacetic acid (5-HIAA) was measured in all subjects (4). Subject diets on days 1 through 5 were modified so that foods containing serotonin precursors were limited.

For all three studies, tigecycline lyophilized powder was supplied in 5-ml open-label vials containing 100 mg of drug. Normal saline (0.9% NaCl in water) was used as the placebo.

**Subjects.** Healthy men aged 18 to 50 years were recruited in all three studies. No clinically abnormal findings on medical history, physical examination, 12-lead electrocardiogram (ECG), or clinical laboratory evaluation were allowed. Subjects who had not taken any over-the-counter, investigational, or prescription drug 14 days before the study or did not have any condition known to interfere with the absorption, distribution, metabolism, or excretion of drugs were included in the study. Subjects who tested positive for human immunodeficiency virus antibodies, hepatitis B surface antigen, and/or hepatitis C antibodies or who had a history or presence of any allergic condition or any major organ or systemic disease were excluded from the study.

All subjects gave written informed consent, and all studies were performed in accordance with the Declaration of Helsinki and its amendments and in accordance with local laws and guidelines. Each of the study protocols was reviewed and approved by an institutional review board or independent ethics committee.

### Bioanalytical methods.

In the SAD and MAD studies, tigecycline was assayed in serum by a high-performance liquid chromatography method with calibrators in the range of 25 to 12,500 ng/ml. Briefly, the method consisted of protein precipitation of the serum samples by addition of acetonitrile, separation of tigecycline on a Supelco LC-18 DB column (150 mm x 4.6 mm [internal diameter], 3-µm particle size) and subsequent UV detection of tigecycline at 350 nm. The extraction efficiency was higher than 76%, and the lower limit of quantitation was 25 ng/ml. The intrabatch and interbatch coefficients of variation (CV) were <13% at all concentrations.

Tigecycline in urine was quantified in the SAD study by a high-pressure liquid chromatography method with UV detection (350 nm) with a lower limit of detection of 2 µg/ml. Briefly, the urine samples were diluted to 1:10, and tigecycline was chromatographed on a C18 column (150 mm x 4.6 mm inner diameter), 3-µm particle size). The intraday CV and bias for the low quality control (QC) samples (6 µg/ml) were 4.6 and −1.5%, respectively. The interday CV and bias values at 6 µg/ml were 7.0 and −1.5%, respectively. The interday (n = 25) precision (CV) values for the middle QC (12 and 36 µg/ml) and the high QC (72 µg/ml) samples were between 5.1 and 9.0%, and the interday bias ranged from −7.9 to −0.6%. The precision (CV) and accuracy (expressed as bias) of tigecycline calibration standards in the lower-tier curve were between 1 and 3.5% and between −2 and 2.3%, respectively, and in the upper-tier curve were between 0.9 and 3.0% and between −5.5 and 3.3%, respectively.

In the VVIR study, tigecycline concentrations in serum were quantified by using a validated methodology that used the API 3000 LC/MS/MS system. The lower limit of quantitation was 10 ng/ml, and the upper limit of quantitation was 2,000 ng/ml. QC samples of tigecycline prepared in human serum at concentrations of 1,500 ng/ml (high), 200 ng/ml (medium), and 25 ng/ml (low) were analyzed, along with the subject samples. The overall precision and accuracy for
the standards and the QC samples were in the range of 0.9 to 12% and 93 to 110%, respectively.

**Pharmacokinetic methods.** Pharmacokinetic parameters based on serum data for tigecycline were estimated by noncompartmental methods with a validated SAS (versions 6.12 and 8.02) program. The AUC from 0 h to the last quantifiable concentration was estimated by using the linear-trapezoidal rule for increasing concentrations and log-trapezoidal rule for decreasing concentrations. The trough serum concentrations of tigecycline ($C_{\text{min}}$) and $C_{\text{max}}$ were obtained directly from observed data. Systemic tigecycline clearance (CL) was normalized to body weight by using the dose/AUC/body weight. The apparent volume of distribution at steady-state ($V_{\text{ss}}$) was calculated based on the following formula: CL/$\lambda_z$ = [(AUMC/AUC) - $T_e$]/2, where AUMC is the area under the first moment curve and $T_e$ is the duration of the i.v. infusion. The apparent terminal-phase disposition rate constant ($\lambda_z$) was estimated by regression of the terminal log-linear concentration time points. The apparent $t_{1/2}$ was estimated as the ln(2)/$\lambda_z$.

In the SAD study, the percentage of tigecycline excreted unchanged in urine ($f_{\text{e,ex}}$ [%]), the amount of tigecycline excreted in urine over 48 h ($Ae_{0-48}$), and renal clearance (CLR = $Ae_{0-48}$/AUC$_{0-48}$) were also determined.

**Statistical methods.** Arithmetic means, standard deviations, and CVs were determined for all pharmacokinetic parameters by using the SAS software. The use of the word “significant” refers to statistically significant results at the level of 0.05. All tests of hypotheses were two sided.

In the SAD study, potential differences among dose groups in concentrations in plasma and pharmacokinetic parameters of tigecycline were assessed by using a one-factor analysis of variance (ANOVA) after normalization of the dose-dependent pharmacokinetic parameters (e.g., $C_{\text{max}}$ and AUC) to the 100-mg dose. The pairwise comparisons among dose groups were made by using the Tukey multiple comparison test.

In the MAD study, tigecycline concentrations in serum and pharmacokinetic parameters on days 1 and 10 were compared by using a one-factor ANOVA and

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**FIG. 1.** Mean tigecycline concentrations in serum in the single ascending dose study in fasting subjects as 1-h infusions (A), as 1-h infusions in the 200-mg dose group (B), and as 4-h infusions (C).
the Tukey multiple comparison test. All $C_{\text{max}}$ and AUC values were normalized to the 100-mg dose before ANOVA was performed. In addition, the dose-normalized serum trough concentrations of tigecycline were compared across dose groups by using ANOVA.

**Safety.** In all three studies, safety was evaluated on the basis of results from scheduled physical examinations, vital sign measurements, ECG, and clinical laboratory evaluations (hematology, blood chemistry, and urinalysis). Adverse events were recorded throughout the study. In the VVIR study, excretion of 5-HIAA and creatinine in urine was determined.

**RESULTS**

**Subjects.** Healthy subjects aged 18 to 44 years participated in the SAD study. Body weights ranged from 51 to 95 kg. Subjects with similar demographics completed the MAD and VVIR studies. Altogether, 66 subjects in the SAD study were analyzed for pharmacokinetic characteristics of tigecycline. In the SAD study, two subjects were excluded from the pharmacokinetic analysis due to improper dosing (injection site reaction in one subject and infusion pump malfunction in another subject), but these subjects were included in the safety evaluation.

**Safety.** The maximum tolerated single dose for fasting subjects was 100 mg. The 200-mg dose was not tolerated by fasting subjects but was well tolerated by fed subjects. Ondansetron appeared to partially improve the tolerability of tigecycline for fasting subjects. GI adverse events were dose limiting at 300 mg, and dose escalation was halted at this dose. Overall, the most common adverse events were nausea (33 of 68 tigecycline recipients [48.5%]) and vomiting (20 of 68 tigecycline recipients [29.4%]), and both of these adverse events tended to increase in frequency with increasing dose (e.g., for fasting administration of 1-h infusions, zero of six subjects experienced nausea at 12.5 mg compared to three of six subjects at 100 mg and five of six subjects at 200 mg). In addition, there were no clinically relevant changes in clinical laboratory parameters, blood pressure, or ECG intervals.

In the MAD study, the dose administration schedule was well tolerated by subjects in the tigecycline 25- and 50-mg dose groups, although GI intolerance occurred in all six tigecycline-treated subjects in the 100-mg dose group, and treatment in that dose group was discontinued after 9 days (total of 17 doses). After the 100-mg dose group completed, an additional 75-mg dose group was enrolled, but this group was discontinued after only 5 days (total of nine doses) due to GI intolerance. Twelve (12) subjects withdrew from the study: one subject (25-mg dose group) developed a rash after receiving 12 doses of tigecycline; ten subjects (one in the 50-mg dose group, six in the 75-mg dose group, and three in the 100-mg dose group) discontinued because of nausea and/or vomiting; and one patient receiving placebo discontinued because of thrombophlebitis. The incidence of nausea increased with increasing dose (four of six subjects, five of six subjects, six of six subjects, and six of six subjects for 25, 50, 75, and 100 mg q12h, respectively). In addition, there were no clinically relevant changes in clinical laboratory parameters, blood pressure, or ECG intervals.

In the VVIR study, the incidence of adverse events did not vary greatly among the treatment groups and the incidence of local irritations, specifically injection site phlebitis, was similar in placebo-treated and tigecycline-treated subjects. Six subjects withdrew from the study because of adverse events: three subjects in the 100-ml/60-min group with reasons that included the presence of gonorrhea before enrollment in the study, as well as diarrhea or vomiting; one subject each in the 100-ml/30-min and 250-ml/60-min groups because of vomiting; and one subject who received placebo in the 250-ml/60-min group because of pruritus and urticaria. Among subjects treated with tigecy-
cline, the incidence of nausea and vomiting was lowest in the 100-ml/30-min treatment group. Subjects given tigecycline at 250 ml/60 min had fewer infusion-related adverse events. However, the incidence of other adverse events, such as nausea and vomiting, was similar to that of the other treatment groups, suggesting that there is little benefit to infusing the drug more slowly and in a larger volume. No deaths and no serious adverse events occurred in any of the three studies.

Tigecycline is most likely excreted in bile, and it is hypothesized that tigecycline may irritate the GI tract, releasing serotonin, which then causes nausea (4). The serotonin metabolite 5-HIAA is commonly measured in urine as a marker of serotonin release. In the VVIR study, there was no definite relationship between the occurrence of nausea and the excretion of 5-HIAA in the urine of subjects (data not shown), suggesting that the nausea and vomiting associated with tigecycline administration are not associated with increased serotonin production.

**Concomitant medications.** The most commonly administered concomitant medications were given for nausea and vomiting in all three studies. In the SAD study, the most commonly administered concomitant medication was the prokinetic agent, metoclopramide used to treat nausea and vomiting in groups 6 through 11. No adverse events were attributed to ondansetron (32 mg) administration. In the MAD study, concomitant medications were not permitted during the study with the exception of antiemetics such as prochlorperazine, trimethobenzamide, or ondansetron. In the VVIR study, the most frequently used concomitant medications were sucralfate—commonly administered concomitant medication was the prokinetic agent, metoclopramide used to treat nausea and vomiting in all three studies. In the SAD study, the most commonly administered concomitant medication was the prokinetic agent, metoclopramide used to treat nausea and vomiting in groups 6 through 11. No adverse events were attributed to ondansetron (32 mg) administration. In the MAD study, concomitant medications were not permitted during the study with the exception of antiemetics such as prochlorperazine, trimethobenzamide, or ondansetron. In the VVIR study, the most frequently used concomitant medications were sucralfate—both given for nausea and vomiting.

**Pharmacokinetics. (i) SAD Trial.** The mean concentration in serum versus time profiles in each of the seven-dose levels used in the study are shown in Fig. 1A. No appreciable differences were seen in tigecycline concentrations in serum after a 200-mg dose, regardless of whether tigecycline was given with or without food (see Fig. 1B). In addition, the concentrations of tigecycline (200 mg given as a 1-h infusion) in serum were not affected by pretreatment with ondansetron (Fig. 1B). Except for the anticipated delay in t_max and the reduction in C_max values due to the slower rate of infusion, the mean pharmacokinetic profiles of subjects receiving 200 and 300 mg as a 4-h infusion (Fig. 1C) were not markedly different from those receiving identical doses as a 1-h infusion (Fig. 1A).

Mean tigecycline C_max after a 1-h infusion ranged from ca. 109 ng/ml after a single dose of 12.5 mg to a 2,817 ng/ml after a dose of 300 mg (Table 2). As expected, mean serum C_max values in the 200-mg (680 ng/ml) and 300-mg (960 ng/ml) dose groups, where the length of the infusion was 4 h, were lower than the values seen after a 1-h infusion of similar doses. Mean values for C_max obtained in the fed (1,528 ng/ml) and fasting (1,643 ng/ml) groups receiving 200-mg doses of tigecycline were not markedly different. Although statistically not significant, the mean C_max values observed in the 200-mg dose group, in which subjects were pretreated with ondansetron, was slightly higher (2,189 ng/ml) than in the other 200-mg dose groups. Differences in the dose-normalized values for C_max were significant (P < 0.05) when examined across all dose groups. However, such differences in dose-normalized C_max values did not exist within the dose groups receiving 1-h in-
TABLE 3. Mean pharmacokinetic parameters of tigecycline after various i.v. doses infused over a 1-h period q12h in the MAD study.

<table>
<thead>
<tr>
<th>Tigecycline dose group (mg)</th>
<th>Day 1</th>
<th>Mean Value (%CV) at:</th>
<th>Day 10*</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>$C_{\text{max}}$ (ng/ml)</td>
<td>$\text{AUC}_{0-\text{ss}}$ (ng h/ml)</td>
<td>$C_{\text{max}}$ (ng/ml)</td>
</tr>
<tr>
<td>25</td>
<td>261 (14)</td>
<td>796 (8)</td>
<td>324 (17)</td>
</tr>
<tr>
<td>50</td>
<td>487 (17)</td>
<td>1,440 (14)</td>
<td>621 (15)</td>
</tr>
<tr>
<td>100</td>
<td>816 (15)</td>
<td>2,389 (13)</td>
<td>1,173 (15)</td>
</tr>
</tbody>
</table>

* Day 10 values for the 100-mg tigecycline dose group are actually day 9 values.

sions or between the two dose groups receiving 4-h tigecycline infusions.

Similarly, the mean AUC ranged from 753 ng h/ml after a 12.5-mg dose to 17,856 ng h/ml after a 300-mg dose, when the doses were given as a 1-h infusion. Dose-normalized AUCs were not significantly different among dose groups ($P > 0.05$, Table 2).

Mean CL was not significantly different among dose levels ($P > 0.2$, Table 2) and ranged from 0.2 to 0.3 liter/h/kg across dose groups. Because of the limitations in sensitivity of the assay that was used to quantify tigecycline, the $t_{1/2}$ of tigecycline could be characterized accurately only after single doses of 100 mg or higher. The mean $t_{1/2}$ was ca. 40 to 60 h in these groups. Tigecycline was well distributed into various tissues as shown by the mean $V_{\text{ss}}$, in excess of 8 liter/kg, at all dose levels at which the $t_{1/2}$ could be accurately characterized ($\geq$100 mg, Table 2). Less than 13% of tigecycline was excreted in urine as unchanged drug. In addition, no significant differences in tigecycline pharmacokinetics were seen between subjects who received tigecycline in the fasting state and those who were given tigecycline in the fed state.

(iii) MAD trial. Pharmacokinetics determined in the MAD study are presented in Table 3 and concentration levels in serum on days 1 and 10 are presented in Fig. 2A and 2B, respectively. The mean $\text{AUC}_{0-\text{ss}}$ values on day 1 ranged from 796 ng h/ml in the tigecycline 25-mg dose group to 2,389 ng h/ml in the tigecycline 100-mg dose group. None of the steady-state pharmacokinetic parameter values (obtained on day 10) were significantly different among dose groups ($P > 0.05$). In all dose groups, tigecycline had a mean CL value of 0.2 liter/h/kg. Mean $t_{1/2}$ of tigecycline obtained on day 10 ranged from 36.9 h in the 50-mg dose group to 66.5 h in the 100-mg dose group. High intersubject variability (CV > 30%) was seen in the $t_{1/2}$ of tigecycline in each dose group examined.

Trough concentrations of tigecycline in serum were compared across days 7, 8, 9, and 10 for the 25-, 50-, and 100-mg q12h dose groups (Table 4). The differences among days were not significant ($P > 0.05$) for the 25- and 100-mg dose groups. In the 50-mg dose group, the trough tigecycline concentrations obtained on days 8 and 9 were higher ($P < 0.05$) than those obtained on days 7 and 10, but the day 7 trough concentrations in this group were not significantly different from those obtained on day 10 ($P > 0.05$). In addition, differences in trough tigecycline concentrations in serum (normalized to 100-mg dose) among dose groups on each of the days 7, 8, 9, and 10 were not significant ($P > 0.05$, Table 4).

(iii) VVIR trial. Only limited pharmacokinetic data were collected in the VVIR trial. The mean tigecycline single-dose $C_{\text{max}}$ (%CV) was 642 ng/ml (24%) for the 100-mg/60-min regimen, 668 ng/ml (3%) for the 250-mg/60-min regimen, and 969 ng/ml (15%) for the 100-mg/30-min regimen. Thus, as expected, the tigecycline $C_{\text{max}}$ was higher for the 30-min infusion than for the 60-min infusion regimens, and the infusion volume did not affect the mean $C_{\text{max}}$ value.

Dose proportionality. In the SAD study tigecycline $C_{\text{max}}$ and AUC increased proportionally to the dose in the range of 12.5 to 300 mg. In fact, a linear relationship was seen between $C_{\text{max}}$ and dose ($r^2 = 0.99$, Fig. 3A) and also between AUC and dose ($r^2 = 0.99$, Fig. 3B).

In the MAD study, dose-normalized values for $C_{\text{max}}$ and AUC on day 1 were not significantly different among the dose groups, and $C_{\text{max}}$ ranged from 261 ng/ml in the 25-mg dose group to 816 ng/ml in the 100-mg dose group (Table 3). The mean $C_{\text{max}}$ values obtained at steady state ranged from 324 ng/ml in the 25-mg dose group to 1,173 ng/ml in the 100-mg dose group. The mean $\text{AUC}_{0-\text{ss}}$ values increased proportionately from 1,482 ng h/ml to 4,980 ng h/ml with an increase in dose from 25 to 100 mg.

DISCUSSION

Tigecycline, a first-in-class glycylcycline with broad-spectrum activity against many pathogens, is being developed for treatment of skin and skin structure infections and intra-abdominal infections (15, 19).

No serious adverse events were reported in any of the three studies. The most common adverse events reported in all studies that appeared to be dose-related were nausea and vomiting, which are common to the tetracycline class of antibiotics. At the higher doses in the SAD study (200 and 300 mg), prolonging the infusion duration to 4 h did not improve the incidence or severity of nausea in this small study ($n = 6$ per group), indicating that the nausea is not directly related to the drug’s $C_{\text{max}}$ in serum. Also, the nausea and vomiting diminished when the 200-mg dose was administered to subjects who had been fed compared to those who had fasted. In phase 2 trials with tigecycline in patients with skin and skin structure infections (tigecycline at 25 or 50 mg q12h) (19) or in patients with intra-abdominal infections (tigecycline at 50 mg q12h) (15), the incidence of nausea and vomiting was lower than was observed in these healthy subjects.

Tigecycline is likely excreted in bile in humans. It has been hypothesized that tigecycline might irritate the GI tract, releasing serotonin, which then causes nausea. In the VVIR study, the urinary recovery of 5-HIAA on study day 1 was $5.57 \pm 1.23$ mg in the nine tigecycline-treated subjects who experienced...
nausea or vomiting on day 1, 5.86 ± 2.94 mg in the nine
tigecycline-treated subjects who did not experience nausea or
vomiting on day 1, and 5.56 ± 0.83 mg in the placebo-treated
subjects, none of whom experienced nausea on day 1. There-
fore, the urinary recovery of 5-HIAA did not correlate with the
occurrence of nausea or vomiting, suggesting that the nausea
and vomiting associated with tigecycline administration may
not be caused by increased serotonin release in the GI tract.

The data obtained from the VVIR study showed that there
were no clinically important differences in safety among the
three groups with various infusion rates. The incidence of
adverse events such as nausea and vomiting was similar among
all three groups, with slightly less nausea and vomiting with the

<table>
<thead>
<tr>
<th>Day</th>
<th>Mean (% cv) concn (ng/ml)</th>
<th>25 mg</th>
<th>50 mg</th>
<th>100 mg</th>
<th>P</th>
</tr>
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<tr>
<td>7</td>
<td>86 (28)</td>
<td>153 (13)</td>
<td>214 (13)</td>
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</tr>
<tr>
<td>8</td>
<td>81 (34)</td>
<td>195 (15)</td>
<td>255 (16)</td>
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<tr>
<td>9</td>
<td>83 (28)</td>
<td>204 (17)</td>
<td>257 (17)</td>
<td>0.06</td>
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</tr>
<tr>
<td>10</td>
<td>87 (26)</td>
<td>145 (16)</td>
<td>NA</td>
<td>0.23</td>
<td></td>
</tr>
</tbody>
</table>

*For comparison among dose groups, concentrations were dose normalized to
the 100-mg dose before statistical analysis. For the 25-, 50-, and 100-mg dose
groups, the P values were 0.98, 0.01, and 0.37, respectively. NA, not available.
shorter 30-min infusion compared to the longer infusions. Adjusting the infusion of tigecycline more rapidly (30 min) and in a smaller volume (100 ml) can be accomplished without significant safety issues. It is important to note that the 30-min infusion was also not associated with an increase in the incidence of local injection site reactions compared to the 60-min infusion. These results indicate that tigecycline could be safely administered as a 30-min infusion.

The serum concentration-time profile of tigecycline after ascending single or multiple doses shows that the decline in drug concentrations after the end of tigecycline infusion follows a polyphasic pattern (Fig. 1 and 2). The steep decrease in tigecycline concentrations in serum at the end of infusion demonstrates the rapid distribution of the drug from the central compartment into the various tissues. This is followed by a long terminal-phase $t_{1/2}$ of ca. 40 h for tigecycline. The long $t_{1/2}$ of tigecycline and the extended postantibiotic effect demonstrated against certain microorganisms (16, 18) are among the reasons for the anticipated efficacy of tigecycline in patients.

The CLR of tigecycline (0.03 liters/h/kg) accounts for ca. 10 to 15% of tigecycline's total systemic CL (ranging from 0.2 to 0.3 liter/h/kg in all SAD and MAD dose groups), and only 10 to 15% of the administered dose is recovered unchanged in urine. Therefore, the renal elimination of tigecycline is a secondary pathway of tigecycline's elimination. Unlike minocycline, which undergoes extensive metabolism (12), no major metabolites of tigecycline have been identified to date, indicating that metabolic elimination is likely to be a minor elimi-
mination pathway. In addition, tigecycline exhibits a high degree of biliary excretion in rats (23). Therefore, the major component of tigecycline systemic CL in humans might be biliary secretion, GI secretion across the gut walls, or both.

In separate experiments, the in vitro protein binding of tigecycline was measured by ultrafiltration and by ultracentrifugation at 37°C (data on file at Wyeth Research). For ultrafiltration, the in vitro protein binding ranged from 71% at 0.1 µg/ml to 87% at 1.0 µg/ml, and for ultracentrifugation the in vitro protein binding ranged from 73% at 0.1 µg/ml to 79% at 1.0 µg/ml. The mechanism for the atypical pattern of increased protein binding at higher concentrations is unknown, but it may be related to the ability of tigecycline to form metal ion complexes. The formation of such complexes by tetracycline has been shown to result in complex interactions with serum proteins that affect diffusion rates across semipermeable membranes and binding to cellular proteins (2, 7, 8).

Tigecycline is extensively distributed into the tissues, as shown by its high volume of distribution in both SAD and MAD studies. In fact, radiolabeled 14C studies in rats have confirmed that tigecycline distributes extensively to various tissues, including lung, skin, liver, heart, and bone (23). The ratio of tigecycline in skin and lungs in rats were nearly three- to fourfold higher than that in plasma. This suggests that for drugs such as tigecycline, concentrations in serum may significantly underestimate the concentration of the drug in various tissues.

After i.v. administration, tigecycline exhibited linear pharmacokinetics after single doses in the range 12.5 to 300 mg and multiple doses of 25 to 100 mg q12h. In fact, the linearity of tigecycline pharmacokinetics is evident by the absence of significant differences in systemic CL among the various dose groups after single and multiple doses. In addition, for dose groups with sufficiently high concentrations in serum to provide a reliable estimate of t1/2 (i.e., doses of >75-mg [single dose]), the Vss did not differ significantly among the dose groups. Trough tigecycline concentrations in serum obtained on days 7, 8, 9, and 10 in all dose groups indicate that tigecycline attained steady-state levels in serum by day 7. As predicted by the t1/2 of 40 to 60 h obtained in these studies, some accumulation of tigecycline is seen after multiple doses, and the observed accumulations (R = C12 h,multiple-dose/C12 h,single-dose) were 2.5, 3.4, and 3.2 for the 25-, 50-, and 100-mg q12h regimens, respectively. Interestingly, the observed accumulation is lower than the theoretical accumulations of 5.3 to 7.7 for a t1/2 of 40 to 60 h [R = 1/(1 - e^{-k/2})], assuming the trough concentration is measured in the terminal disposition phase of the curve. Another method to evaluate the observed and theoretical accumulation is to compare the single-dose AUC0-12 h in serum from the MAD study and the multiple-dose AUC0-12 h in serum from the SAD study showed that the single-dose AUC0-12 h underestimated the multiple-dose AUC0-12 h by ca. 20% for the 50-mg dose and overpredicted the multiple-dose AUC0-12 h by ca. 20% for the 100-mg dose. Therefore, the accumulation of tigecycline after multiple-dose administration is approximately linear, but these comparisons should be interpreted cautiously because the AUCs were measured in different populations and the comparisons are based on a small number of subjects (n = 6 per group).

This article establishes the pharmacokinetic profile and dose proportionality of tigecycline when given as single or multiple twice-daily doses to healthy subjects; it also shows that the infusion duration can be varied without significant safety concerns. These characteristics, along with the in vitro activity and extended postantibiotic effect of tigecycline against several strains of microorganisms, make tigecycline a promising antibiotic agent.

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