Antimicrobial Drug Resistance of *Salmonella enterica* Serovar Typhi in Asia and Molecular Mechanism of Reduced Susceptibility to the Fluoroquinolones

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This study describes the pattern and extent of drug resistance in 1,774 strains of *Salmonella enterica* serovar Typhi isolated across Asia between 1993 and 2005 and characterizes the molecular mechanisms underlying the reduced susceptibilities to fluoroquinolones of these strains. For 1,393 serovar Typhi strains collected in southern Vietnam, the proportion of multidrug resistance has remained high since 1993 (50% in 2004) and there was a dramatic increase in nalidixic acid resistance between 1993 (4%) and 2005 (97%). In a cross-sectional sample of 381 serovar Typhi strains from 8 Asian countries, Bangladesh, China, India, Indonesia, Laos, Nepal, Pakistan, and central Vietnam, collected in 2002 to 2004, various rates of multidrug resistance (16 to 37%) and nalidixic acid resistance (5 to 51%) were found. The eight Asian countries involved in this study are home to approximately 80% of the world’s typhoid fever cases. These results document the scale of drug resistance across Asia. The Ser83→Phe substitution in GyrA was the predominant alteration in serovar Typhi strains from Vietnam (117/127 isolates; 92.1%). No mutations in gyrB, parC, or parE were detected in 55 of these strains. In vitro time-kill experiments showed a reduction in the efficacy of ofloxacin against strains harboring a single-aminocacid substitution at codon 83 or 87 of GyrA; this effect was more marked against a strain with a double substitution. The 8-methoxy fluoroquinolone gatifloxacin showed rapid killing of serovar Typhi harboring both the single- and double-aminocacid substitutions.

There are approximately 21 million cases of typhoid fever worldwide, with a particularly high incidence in Asia. An estimated 220,000 deaths per year occur as a consequence of the disease (11).

This article describes the extent and pattern of drug resistance of *Salmonella enterica* serovar Typhi across Asia. This information is vital for guiding treatment and is also important for helping policy makers to plan vaccination campaigns. The emergence and spread of drug resistance have limited treatment options for typhoid fever in many countries.

Since the isolation of multidrug-resistant (MDR) serovar Typhi strains which show resistance to all first-line antibiotics (chloramphenicol, ampicillin, and trimethoprim-sulfamethoxazole) in the 1980s, the fluoroquinolone class of antibiotics has become the treatment of choice for enteric fever (4, 38). Unfortunately, outbreaks of serovar Typhi strains that were resis-

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tant to nalidixic acid (the prototype quinolone, which is used for in vitro screening tests) and showed reduced susceptibility to the fluoroquinolones have been reported subsequently in a number of countries (25). Vietnam and particularly the Mekong Delta region of Vietnam faced a series of typhoid fever epidemics over the last decade, imposed on a background of endemic disease, that reflected changes in resistance patterns and pointed to a serious problem of drug resistance (24). MDR is associated with a transferable plasmid (36), while reduced susceptibility to the fluoroquinolones in serovar Typhi is usually associated with point mutations in the bacterial target genes encoding DNA gyrase and/or DNA topoisomerase IV.

This study describes the magnitude and patterns of drug resistance in 1,393 serovar Typhi strains isolated from 1993 to 2005 in Vietnam and from a cross-sectional sample set of 381 serovar Typhi strains isolated in 2002 to 2004 in eight Asian countries (Bangladesh, China, India, Indonesia, Laos, Nepal, Pakistan, and central Vietnam). These countries are home to more than 80% of the world’s typhoid fever cases (11). We defined the molecular mechanism of nalidixic acid resistance and performed in vitro bacterial time-kill experiments with isolates that harbored the common mutations in the gyrA gene.

The time-kill experiments allowed us to model the impact of the gyrA mutations on the time course of the antimicrobial effects of older (ofloxacin) and newer-generation (gatifloxacin) fluoroquinolones.

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### MATERIALS AND METHODS

#### Bacterial isolates.

(i) Serovar Typhi isolates collected in southern Vietnam from 1993 to 2005. One thousand three hundred ninety-three serovar Typhi isolates were collected consecutively from patients with uncomplicated typhoid fever during prospective hospital-based clinical studies between 1993 and 2005 conducted at Dong Thap Provincial Hospital, Dong Nai Pediatric Hospital, An Giang Provincial Hospital, and the Hospital for Tropical Diseases, Ho Chi Minh City, all located in southern Vietnam. These studies have been described previously (7, 8, 23, 26, 32).

(ii) Serovar Typhi isolates from eight Asian countries in 2002 to 2004.

One hundred eighty-two serovar Typhi isolates were collected consecutively from patients with uncomplicated typhoid fever during the indicated periods and came from geographically contiguous areas. The isolates were unselected and were representative of the population they came from.

#### Identification and antimicrobial susceptibilities.

Isolates were identified using the API20E biochemical identification system (bioMerieux, Paris, France). Serology was carried out using specific antisera (polyvalent O, O9, H4, and Vi) (Murex, Dartford, United Kingdom).

Antimicrobial susceptibility testing with ampicillin, chloramphenicol, trimethoprim-sulfamethoxazole, nalidixic acid, ofloxacin, ciprofloxacin, gatifloxacin, and ceftriaxone was performed by disc diffusion according to Clinical and Laboratory Standards Institute (CLSI) methods (10) and interpreted following CLSI guidelines (9). The control strains used for all susceptibility tests were Escherichia coli ATCC 25922, Pseudomonas aeruginosa ATCC 27853, and Staphylococcus aureus ATCC 29213. MICs were determined by using the E-test (AB Biodisk, Solna, Sweden). MDR of isolates was defined as resistance to chloramphenicol (MIC > 32 μg/ml), ampicillin (MIC > 32 μg/ml), and trimethoprim-sulfamethoxazole (MIC > 8/152 μg/ml). Nalidixic acid resistance was defined as a MIC of ≥32 μg/ml. The breakpoints for oxolinic and gatifloxacin were ≥2 μg/ml (susceptible) and ≥8 μg/ml (resistant), and for ciprofloxacin, ≤1 μg/ml (susceptible) and ≥4 mg/ml (resistant) (9). All tests were performed at the Hospital for Tropical Diseases (HTD), Ho Chi Minh City, Vietnam, except for the isolates from Nepal, which were tested at Patan Hospital, Kathmandu, Nepal, using identical methods.

#### DNA isolation.

A single colony was inoculated in 6 ml of LB broth (Sigma) and incubated overnight at 37°C. DNA was extracted using the Qian Genomic-tip 100 G and Genomic DNA buffer set (Qiagen, Ltd., Hilden, Germany) or the isolates from Nepal, which were tested at Patan Hospital, Kathmandu, Nepal, using identical methods.

#### PCR and sequencing.

Oligonucleotide primer pairs are shown in Table 1. PCR amplifications of gyrA (347 bp), gyrB (345 bp), parC (270 bp), and parE (240 bp) were performed with 30 cycles of denaturation at 92°C for 1 min, annealing at 62°C for 1 min, and extension at 74°C for 2 min, followed by a final extension step at 74°C for 1 min.

PCR products were purified using the QIAquick PCR purification kit (Qiagen GmbH, Hilden, Germany) and used directly as templates for sequencing, which was performed with the CEQ DTCs-Quick Start kit and analyzed using an automated sequencer, the CEQ8000 genetic analysis system (Beckman Coulter, Inc., Fullerton, CA).

Selected strains were screened for the presence of the qnrA and qnrS genes by PCR. The PCR conditions for the amplification of qnrS were as follows: 94°C for 2 min; 34 cycles of 94°C for 45 s, 48°C for 45 s, and 72°C for 45 s; final extension at 74°C for 5 min. PCR conditions for qnrS were identical except for the annealing temperature, which was 53°C. The positive control used was a Citrobacter sp. isolate (identified by API20E) harboring both the qnrS and qnrS genes, as confirmed by sequencing of PCR products.

#### In vitro time-kill analysis.

All time-kill experiments were performed in duplicate. Ofl oxacin powder was purchased from Sigma, Steinheim, Germany, and gatifloxacin powder was provided from Bristol-Myers Squibb, New Brunswick, NJ. Three serovar Typhi colonies were taken and inoculated in 10 ml Mueller-Hinton broth (Oxoid, Basingsfield, United Kingdom) at 37°C for 15 to 18 h. Two drops of this broth were inoculated into 10 ml of Mueller-Hinton broth and incubated at 37°C for 1 h to give 2 × 10⁶ CFU/ml. Ten milliliters of Mueller-Hinton broth containing ofloxacin or gatifloxacin at 32X MIC was added at time zero to give a final concentration of 16X MIC; serial twofold dilutions were used.
to obtain $8 \times 4 \times 2 \times 1 \times 1$ MIC. The growth control contained no antibiotic. The cultures were incubated at 35 to 37°C for 24 h. Viable counts were measured immediately prior to the addition of the antibiotic and at 30 min and 1, 2, 4, 6, 8, and 24 h after the addition of the antibiotic. Viable counts were performed by using the Miles and Misra technique on nutrient agar plates following serial dilution in maximum-recovery diluents (Oxoid, United Kingdom). The lower limit of detection was $10^2$ CFU/ml.

**Nucleotide sequence accession numbers.** The partial DNA sequences of the gyrA gene of serovar Typhi AG 152 and DT 18 have been registered in the GenBank nucleotide sequence database under the accession numbers EF680460 and EF680461, respectively.

**RESULTS**

**Antimicrobial susceptibility testing. (i) Serovar Typhi isolated in southern Vietnam from 1993 to 2005.** Between 1993 and 2005, 1,393 isolates of serovar Typhi were collected (Fig. 1). The proportion of MDR serovar Typhi was 63.2% (36/57 strains) in 1993 and increased to more than 80% in the late 1990s and early 2000. During the same period, there was a dramatic increase in nalidixic acid resistance. In 1993, 2 out of 57 (3.5%) serovar Typhi isolated from patients in southern Vietnam were nalidixic acid resistant (respective MICs of ofloxacin, 0.250 and 0.125 $\mu$g/ml) (37). Nalidixic acid resistance surged to 88.6% (109/123) in 1998. It has remained at high levels since then, with 97% (196/202) of isolates in 2004. Since 1998, a high proportion of strains show the combination of MDR and nalidixic acid resistance (Fig. 1).

The antimicrobial susceptibility data of 202 serovar Typhi isolated in 2004 in southern Vietnam are shown in more detail in Table 2.

**Antimicrobial susceptibility testing. (ii) Serovar Typhi strains isolated in eight Asian countries in 2002 to 2004.** The antimicrobial susceptibilities of 381 serovar Typhi isolates collected in 2002 to 2004 from eight Asian countries were analyzed (Table 2). There were various rates of MDR across the sites, ranging from 16% (8/50) of isolates from Laos to 37.5% (15/40) from Bangladesh. China and Indonesia were exceptions, with no MDR serovar Typhi identified.

The percentages of nalidixic acid-resistant serovar Typhi isolates ranged from 0% in Indonesia and Laos and 4.8% (1/21) in China to 51% (76/149) in Nepal (Table 2). The com-

![FIG. 1. Antimicrobial drug resistance of serovar Typhi strains isolated during clinical studies in southern Vietnam from 1993 to 2005. Percentages of MDR and nalidixic acid-resistant serovar Typhi isolates. The number of isolates from each year is shown on top of the bars.](http://aac.asm.org/)
<table>
<thead>
<tr>
<th>Isolate*</th>
<th>Yr of isolation</th>
<th>Country or province*</th>
<th>Amino acid substitution(s) in gyrA</th>
<th>Nucleotide change(s) in gyrB</th>
<th>ORDR profile/ ( \text{MIC of drug (µg/ml)} )</th>
<th>Presence of MDR</th>
<th>MIC of drug (µg/ml)</th>
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<td>0.125 1 0.094</td>
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<td>TCC→TTC</td>
<td>Yes</td>
<td>256</td>
<td>0.25 1 0.19</td>
</tr>
</tbody>
</table>

* Isolate names consist of an abbreviation for the study followed by the isolate number. *, strain screened for presence of gyrA and gyrB genes by PCR.

b) An Giang Province, Dong Thap Province, Can Tho Province, Tien Giang Province, Long An Province, and Ho Chi Minh City (HCMC) are located in southern Vietnam.

c) wt, wild type.
bination of MDR and nalidixic acid resistance was found in 4.3% (2/47) of serovar Typhi isolates from central Vietnam, 8.7% (2/23) of isolates from India, 23.5% (8/140) of isolates from Pakistan, and 30% (12/40) of isolates from Bangladesh. In Nepal, 18.1% (27/149) of serovar Typhi isolates were resistant to chloramphenicol and nalidixic acid.

However, using current CLSI breakpoints, all isolates remained susceptible in vitro to ciprofloxacin and ofloxacin, with the exception of one isolate from southern Vietnam, AG 152, with intermediate susceptibility (MIC, 3.0 μg/ml) to ofloxacin (Table 3) and six isolates (4%) from Nepal that were ciprofloxacin resistant. The highest MICs of gatifloxacin at which 50% and 90% of serovar Typhi isolates were inhibited were 0.125 μg/ml and 0.25 μg/ml, respectively (Table 2). All isolates were susceptible to ceftriaxone.

**DNA sequence analysis of QRDR of DNA gyrase and DNA topoisomerase IV and effect of mutations on fluoroquinolone susceptibility.** One hundred twenty-seven nalidixic acid-resistant serovar Typhi isolates (118 from southern Vietnam, 5 from India, and 4 from Pakistan) with reduced susceptibilities to the fluoroquinolones (MIC of ofloxacin ranging from 0.5 μg/ml to 3 μg/ml) were selected for molecular analysis of the quinolone resistance determining region (QRDR) of gyrA. Six different types of mutations were detected. The most prevalent amino acid substitution was Ser83→Phe (TCC→TTC) in 117/127 (92.1%) strains. Four isolates (3.1%) had an alteration at codon 83 changing Ser to Tyr (TCC→TAC). Two isolates showed the Asp87→Gly (GAC→GCC) substitution and two isolates the Asp87→Ala (GAC→GCC) substitution. Two serovar Typhi isolates had double-amino-acid substitutions in GyrA: isolates DT 18 (Ser83→Phe and Asp87→Gly) and AG 152 (Ser83→Phe and Asp87→Asn), as shown in Table 3.

Fifty-five of these strains were analyzed for mutations in the QRDR of gyrB, parC, and parE (13, 20); no mutations were detected (Table 3). Twenty-five isolates (indicated with an asterisk in Table 3) were screened for the presence of the plasmid-mediated quinolone resistance genes qnrA and qnrS (15); none were detected in these isolates.

**In vitro time-kill analysis.** One isolate representing each mutation group was selected for in vitro time-kill experiments: CT 76, wild-type strain (MICs, 0.064 μg/ml for ofloxacin and 0.008 μg/ml for gatifloxacin); HTD 798 (Ser83→Phe; MICs, 1.0 μg/ml for ofloxacin and 0.13 μg/ml for gatifloxacin); CT 144 (Asp87→Gly; MICs, 1 μg/ml for ofloxacin and 0.094 μg/ml for gatifloxacin); and DT 18 (Ser83→Phe and Asp87→Gly; MICs, 2.0 μg/ml for ofloxacin and 0.25 μg/ml for gatifloxacin). The mean changes in log10 CFU/ml are presented in Fig. 2. Ofloxacin showed rapid killing of wild-type strain CT 76 (Fig. 2a); viable counts of serovar Typhi HTD 798 and CT 144 decreased after 4 h at ×4 MIC, but complete killing could not be achieved (Fig. 2b and c). No bactericidal activity was achieved against serovar Typhi DT 18 (Fig. 2d). Gatifloxacin at ×4 MIC decreased the bacterial population of CT 76, HTD 798, and CT 144 (Fig. 2e, f, and g) in the first 30 min and showed complete killing after 6 h. Viable counts of serovar Typhi DT 18 decreased after 4 h, followed by regrowth; higher concentrations (×8 or 16× MIC) showed a more pronounced bactericidal effect against this double mutant (Fig. 2h).

**DISCUSSION**


In 1993, during the initial outbreak of MDR serovar Typhi in Kien Giang province in the south of Vietnam, the fluoroquinolone antibiotics were introduced for the treatment of typhoid fever (22). Since 1993, the proportion of MDR serovar Typhi has remained at high levels and there has been a dramatic increase in nalidixic acid resistance. In 1998, 5 years after ofloxacin and ciprofloxacin became widely available in an uncontrolled market, 87% of the isolates were resistant to nalidixic acid; this increased to 97% by 2004. The combination of MDR and nalidixic acid resistance is a particular problem in Vietnam, because it severely restricts the therapeutic options for patients with typhoid fever.

Patients infected with nalidixic acid-resistant serovar Typhi show poor clinical response, high failure rates (up to 36%), and prolonged fecal carriage when treated with an older-generation fluoroquinolone, such as ofloxacin (8, 26). The antimicrobial resistance data from southern Vietnam are complemented by the results of a cross-sectional study from eight Asian countries: Bangladesh, China, India, Indonesia, Laos, Nepal, Pakistan, and Vietnam. These countries are home to approximately 80% of the world’s typhoid fever cases (11).

While in southern Vietnam the MDR phenotype of serovar Typhi has remained at high levels over the last 13 years, there have been reports of a return to chloramphenicol sensitivity in some regions (12, 21). However, in our study the prevalence of chloramphenicol resistance remained high in many Asian countries (18% in Laos, 19% in Nepal, 26% in India and Pakistan, and 40% in Bangladesh), with the exception of China and Indonesia.

In 2002 to 2004, all countries in the region, with the exception of China and Laos, faced a problem of nalidixic acid resistance, with southern Vietnam as a particular hot spot. Roumagnac et al. recently suggested that fluoroquinolone use has driven the clonal expansion of a nalidixic acid-resistant serovar Typhi haplotype, H58, in Southeast Asia (29). The emergence of resistance of serovar Typhi to ciprofloxacin (6/149 isolates; 4%) in Nepal, together with reports of high-level ciprofloxacin resistance in India and Bangladesh (14, 28, 30), might be the prelude to a worsening drug resistance problem in Asia.

In this study carried out across Asia, mutations associated with nalidixic acid resistance and reduced susceptibility to fluoroquinolones for serovar Typhi were defined only in gyrA, as single-amino-acid substitutions at either codon 83 or 87 (6, 18, 31, 37), with the exception of two isolates from Vietnam, which had double-amino-acid substitutions. There have been two recent reports of serovar Typhi with the Ser83Phe and Asp87Gly double alteration in high-level-ciprofloxacin-resistant serovar Typhi (28, 30). In our study, the isolates with double mutations in gyrA were less susceptible to the fluoroquinolones, and this phenotype may become more widespread in the future if continued drug pressure is applied. This is a particular problem in many parts of Asia, where antibiotics are readily available in an unregulated marketplace and inadequate doses and durations of antibiotics are often used.
FIG. 2. In vitro time-kill experiments of wild-type serovar Typhi and serovar Typhi harboring single and double amino acid substitutions in GyrA. Figure 2a to d shows exposure to ofloxacin, and Fig. 2e to h shows exposure to gatifloxacin at concentrations of 1× to 16× MIC over 24 h. Results represent means of duplicate values; the standard deviation is indicated by error bars.
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FIG. 2—Continued.

Serovar Typhi CT 76 (wild type) vs. Gatifloxacin

Serovar Typhi HTD798 (Ser83Phe) vs. Gatifloxacin

Serovar Typhi CT 144 (Asp87Gly) vs. Gatifloxacin

Serovar Typhi DT 18 (Ser83Phe and Asp87Gly) vs. Gatifloxacin

FIG. 2—Continued.
Our time-kill experiments suggest that the choice of the fluoroquinolone and the dose used for the treatment of serovar Typhi may be critical and underline that clearly not all the fluoroquinolones are as susceptible to these common mutations. Continued use of the older-generation fluoroquinolones (ofloxacin and ciprofloxacin) may encourage the persistence of fluoroquinolone resistance. Knowledge of the extent of drug resistance should lead to the development of new muta-
tions which might compromise the efficacy of the newer generation. With lower MICs and better responses in the time-kill experiments, which might compromise the efficacy of the newer generation, the choice of the fluoroquinolone and the dose used for the treatment of serovar Typhi and Paratyphi A caus-


3. REFERENCES


