Optimized Elution of Daptomycin from Polymethylmethacrylate Beads

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We demonstrate that xylitol can be added to polymethylmethacrylate (PMMA) bone cement to enhance the elution of daptomycin in terms of both the peak and sustained release of antibiotic. We also demonstrate that a PMMA-xylitol formulation optimized for daptomycin can be used to enhance the elution of both vancomycin and gentamicin.

The treatment of chronic musculoskeletal and implant-associated infection requires a multifactorial approach that can include systemic antibiotic therapy, the surgical debridement of the affected site, including the removal of infected devices, and local antibiotic therapy targeted directly to the site of infection (1). The latter often is done by incorporating antibiotics into handmade polymethylmethacrylate (PMMA) beads (2). Elution profiles from antibiotic-laden PMMA beads can be modified by the incorporation of biologically inert fillers that increase both the rate and duration of antibiotic release (2–5). One such filler is glycine; indeed, PMMA beads containing glycine and gentamicin are commercially available in Europe under the trade name Septopal (Biomet Europe, Dordrecht, The Netherlands), and these beads exhibit an enhanced elution profile compared to that of handmade beads lacking glycine (6). However, in previous studies we found that xylitol enhances elution to a greater extent than glycine (3). The experiments described in this report were aimed at optimizing the use of xylitol as a means of enhancing the elution of antibiotics from PMMA.

To evaluate the impact of xylitol on the elution of daptomycin from PMMA, a 40-g packet of Palacos-R PMMA (Zimmer, Inc., Dover, OH) powder was hand mixed with the designated amounts of daptomycin with or without xylitol crystals (Sigma Chemical Co., St. Louis, MO). Polymerization was done using a single vial of monomer irrespective of the presence or absence of xylitol. Each PMMA preparation was mixed until a single vial of monomer irrespective of the presence or absence of xylitol. Each PMMA preparation was mixed until well homogenized, and then formulated to a predetermined PMMA concentration as follows: 1.0 g of daptomycin and 18 or 22 g of xylitol per 40-g packet of PMMA. Based on our earlier experiments demonstrating that the incorporation of 1.0 g of daptomycin and 28.0 g of xylitol failed to yield an elution profile that met either of these objectives (3), we initially used 2.0 g of daptomycin and 0, 7, 14, or 28 g of xylitol per 40-g packet of PMMA. Based on two-way analysis of variance with Tukey post hoc adjustments (SigmaStat, version 2; SPSS Inc., Chicago, IL), we confirmed a significant difference (P ≤ 0.001) not only between PMMA and PMMA containing as little as 7 g of xylitol but also between PMMA formulations containing increasing amounts of xylitol. While all formulations, including the one that lacked xylitol, yielded a peak concentration of ≥100 μg/ml, the only formulation that yielded a sustained concentration at least 5× the MIC was the one containing 28 g of xylitol (Fig. 1).

We chose as our goal an elution profile that included a peak concentration at least 100× and a sustained concentration (defined at 10 days postelution) at least 5× the MIC defined by the CLSI as the breakpoint for a daptomycin-sensitive strain of S. aureus (UAMS-1; daptomycin MIC, ≤0.5 μg/ml). Based on our earlier experiments demonstrating that the incorporation of 1.0 g of daptomycin and 28.0 g of xylitol failed to yield an elution profile that met either of these objectives (3), we initially used 2.0 g of daptomycin and 0, 7, 14, or 28 g of xylitol per 40-g packet of PMMA. Based on two-way analysis of variance with Tukey post hoc adjustments (SigmaStat, version 2; SPSS Inc., Chicago, IL), we confirmed a significant difference (P ≤ 0.001) not only between PMMA and PMMA containing as little as 7 g of xylitol but also between PMMA formulations containing increasing amounts of xylitol. While all formulations, including the one that lacked xylitol, yielded a peak concentration of ≥100 μg/ml, the only formulation that yielded a sustained concentration at least 5× the MIC was the one containing 28 g of xylitol (Fig. 1).

We next carried out an experiment using formulations containing 2.0 g of daptomycin and 18 or 22 g of xylitol per 40-g packet of PMMA. The results demonstrated that the 100× elution objective was met with either formulation, but that 18 g of xylitol was not sufficient with respect to the 5× objective...
The lower dashed line indicates the 5 g/ml threshold (5 µg/ml), while the lower dashed line represents the 5× threshold (100 µg/ml). Statistical analysis confirmed a significant difference (P ≤ 0.05) between PMMA formulations containing 2 g of vancomycin with (●) and without (▲) xylitol. In contrast, there was no significant difference between a PMMA formulation containing 4 g of vancomycin without xylitol (●) and 2 g of vancomycin with xylitol (■).

FIG. 2. Elution profile with the optimized formulation of PMMA containing 2.0 g daptomycin and 22 g of xylitol per 40-g packet of PMMA. Results are reported as the averages ± standard deviations from six replicates. The upper dashed line indicates the 100× threshold (100 µg/ml), while the lower dashed line represents the 5× threshold (5 µg/ml). In contrast, a PMMA formulation containing 3.6 g of gentamicin but without xylitol fell below the 5× objective (20 µg/ml) irrespective of the inclusion of xylitol (Fig. 4). Nevertheless, the addition of xylitol did enhance elution, as evidenced by the fact that a PMMA formulation containing 3.6 g of gentamicin but without xylitol fell below the 5× objective by day 3 and reached undetectable levels by day 7 (Fig. 4). In contrast, a formulation containing 3.6 g of gentamicin and 22 g of xylitol yielded an elution profile that met or exceeded the 5× threshold through day 6 and yielded detectable levels of antibiotic (10 µg/ml) even on day 10 (Fig. 4). Moreover, this was true even when the amount of gentamicin was reduced from 3.6 to 2.0 g per 40 g of PMMA. In fact, PMMA formulations containing xylitol were the only ones in which the amount of antibiotic observed on day 10 remained above the CSLI breakpoint MIC (≤4.0 µg/ml) for a gentamicin-sensitive strain of S. aureus. Subsequent analysis confirmed that the differences observed in the elution profile of PMMA with and without xylitol were statistically significant (P ≤ 0.001).

Taken together, the results we report demonstrate that the addition of 22 g of xylitol to a 40-g packet of PMMA enhances the elution of vancomycin from PMMA, creating a need for future work to determine the role of xylitol in PMMA formulations.
the elution profiles of daptomycin and, to a somewhat lesser extent, vancomycin and gentamicin. This was true in terms of both peak and sustained concentrations of antibiotic. The inclusion of xylitol did not necessitate the alteration of the standard method for preparing PMMA beads, in that the single vial of monomer provided by the manufacturer remained sufficient to achieve polymerization. The verification of the significance of these in vitro observations will require in vivo studies taking into account both the complete resolution of the infection and the time required to achieve this outcome, but taken together we believe the results we report support the hypothesis that PMMA beads containing xylitol offer a therapeutic advantage over the current clinical practice.

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REFERENCES