Antimalarial Activity of Simalikalactone E, A New Quassinoid from *Quassia amara* (Simaroubaceae)\(\uparrow\)

N. Cachet,\(\ddagger\)‡ F. Hoakwie,\(\ddagger\)§ S. Bertani,\(\ddagger\)¶ G. Bourdy,\(\ddagger\)\(\ddagger\) E. Deharo,\(\ddagger\)\(\ddagger\) D. Stien,\(\ddagger\) E. Houel,\(\ddagger\) H. Gornitzka,\(\ddagger\) J. Fillaux,\(\ddagger\) S. Chevalley,\(\ddagger\) A. Valentín,\(\ddagger\)¶ and V. Jullian\(\ddagger\)\(\ddagger\)¶

Laboratoire de Pharmacochimie des Substances Naturelles et Pharmacophores Redox, UMR 152, UPS, Université de Toulouse, 118 Route de Narbonne, F-31062 Toulouse Cedex 9, France; Institut de Recherche pour le Développement, UMR 152, 118 Route de Narbonne, F-31062 Toulouse Cedex 9, France; USM 0307, Laboratoire de Parasitologie Comparée et Modèles Expérimentaux, Muséum National d’Histoire Naturelle, Paris, France; Institut de Recherche pour le Développement, UMR 152, Mission IRD Casilla 18-1209 Lima, Peru; CNRS, UMR Ecofog, Université des Antilles et de la Guyane, Cayenne, France; CNRS, LCC, UPR 8241, 205 Route de Narbonne, 31077 Toulouse Cedex 4, France; and Service de Parasitologie-Mycologie, CHU Rangueil, 1 Avenue de Pr. Jean Pouldhès, TSA 50032, 31095 Toulouse Cedex, France

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We report the isolation and identification of a new quassinoid named simalikalactone E (SkE), extracted from a widely used Amazonian antimalarial remedy made out of *Quassia amara* (Simaroubaceae) leaves. This new molecule inhibited the growth of *Plasmodium falciparum* cultured in vitro by 50%, in the concentration range from 24 to 68 nM, independently of the strain sensitivity to chloroquine. We also showed that this compound was able to decrease gametocytemia with a 50% inhibitory concentration sevenfold lower than that of primaquine. SkE was found to be less toxic than simalikalactone D (SkD), another antimalarial quassinoid from *Q. amara*, and its cytotoxicity on mammalian cells was dependent on the cell line, displaying a good selectivity index when tested on nontumorigenic cells. In vivo, SkE inhibited murine malaria growth of *Plasmodium vinckei petteri* by 50% at 1 and 0.5 mg/kg of body weight/day, by the oral or intraperitoneal routes, respectively. The contribution of quassinoids as a source of antimalarial molecules needs therefore to be reconsidered.

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*Corresponding author. Mailing address: Institut de Recherche pour le Développement, UMR 152, Faculté de Pharmacie, Université de Toulouse, 35 Chemin des Maraichers, Toulouse 31062, France. Phone: 33 5 62 25 68 23, E-mail: jullian@ict.fr.

† Supplemental material for this article may be found at http://aac.asm.org.

‡ Present address: LCMBIA, UMR 6001, CNRS-Université Nice Sophia Antipolis, 06108 Nice Cedex 02, France.

§ Present address: LSPCMIB, UMR 5068, CNRS-Université Paul Sabatier, 31062 Toulouse Cedex 09, France.

¶ Present address: Department of Biochemistry, University of California, Riverside, CA 92521.

\(\ddagger\) These two senior investigators made equal contributions to this study.

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Parasites were cultured by the method of Trager and Jensen (32) with modifications (34). The cultures were synchronized by a combination of magnetic enrichment and 5% n-sorbitol lysis (Merkert, Darmstadt, Germany) (22-27). In vitro antimalarial activity testing was performed by the method of Desjardins et al. (12) with modifications (34).

The sensitivity of the different asexual erythrocytic stages of \textit{P. falciparum} to SkE was determined on the FcB1 strain, synchronized on a 4-hour period as previously described (19). Cultures in 24-well plates (1% parasitemia, 2% hematocrit) were subjected to 8-hour pulses of SkE at 8.6 and 86.5 nM. After the pulses, the cultures were washed three times with culture medium and returned to normal culture conditions. At the end of the experiment (70 hours, young trophozoite stage of the next erythrocytic cycle), parasitemia was evaluated in each well by visual examination by counting Giemsa-stained smears (5,000 erythrocytes/smear). Results are expressed as the percentage of inhibition of parasitic growth.

In vitro activity was expressed as fold inhibition (F32, FcB1, and W2 strains) and sexual (W2 Indochina strain) stages and cytotoxicity of SkE.

### Table 1. Antiplasmodial activity against asexual (F32, FcB1, and W2 strains) and sexual (W2 Indochina strain) stages and cytotoxicity of SkE

<table>
<thead>
<tr>
<th>Drug (^a)</th>
<th>F32 Tanzania</th>
<th>FcB1 Colombia</th>
<th>W2 Indochina</th>
<th>W2 Indochina (gametocytes)</th>
<th>Vero</th>
<th>MCF7</th>
<th>THP1</th>
</tr>
</thead>
<tbody>
<tr>
<td>SkE</td>
<td>68 ± 12</td>
<td>45 ± 32</td>
<td>24 ± 10</td>
<td>1,120 ± 400</td>
<td>6,574 ± 264</td>
<td>47 ± 2</td>
<td>33 ± 5</td>
</tr>
<tr>
<td>SkD</td>
<td>NT</td>
<td>1</td>
<td>NT</td>
<td>NT</td>
<td>58 ± 11</td>
<td>&lt;20(c)</td>
<td>&lt;2(d)</td>
</tr>
<tr>
<td>CQ</td>
<td>36 ± 3</td>
<td>167 ± 32</td>
<td>196 ± 16</td>
<td>NT</td>
<td>&gt;500(d)</td>
<td>&gt;500(d)</td>
<td>&gt;500(d)</td>
</tr>
<tr>
<td>PMQ</td>
<td>6.17 ± 1.2</td>
<td>8.9 ± 1.1</td>
<td>7.14 ± 0.9</td>
<td>8.9 ± 2.3</td>
<td>340 ± 29</td>
<td>NT</td>
<td>NT</td>
</tr>
</tbody>
</table>

\(\text{IC}_{90}\) values are expressed as nanomolar (except for PMQ, which is shown in micromolar). Each value corresponds to the mean ± standard error of the mean from at least three independent experiments. NT, not tested.

### Results

The chloroform extract of a tea made with defatted dry and mature leaves, which retained biological activity, was depigmented and further fractionated by countercurrent chromatography (see Scheme S1 in the supplemental material). Seven fractions were obtained (fraction 1 [F1] to F7). Previous purification with the same protocol but on a smaller quantity allowed us to identify two active fractions (IC\(_{90}\) < 1 \(\mu\)g/ml) with a similar composition to F1, F2, and F6 (according to their thin-layer chromatography profiles). F1, F2, and F6 were therefore further purified. From F6, we were not able to isolate any pure compounds. SkD was isolated from F2 (0.0002% yield \{i.e., 2 mg from 1 kg \(\text{dry weight}\) of plant\}). More interestingly, a new quassinoid structurally related to SkD (that we called SkE) was isolated from F1 and showed a very good activity in vitro on the different \textit{P. falciparum} strains (Table 1). However, the yield obtained for this active compound was very low (0.00035% \{i.e., 3.5 mg from 1 kg \(\text{dry weight}\) of plant\}) and precluded any further investigation of its antimalarial properties.

We then set up an improved extraction procedure from the methanol extract of the mature dry leaves of \textit{Q. amara} and increased the yield to 0.004% (40 mg from 1 kg of plant). Careful analysis of infrared, optical rotation, and mass and NMR spectra enabled us to establish the structure of SkE. Crystals were also obtained from deuterated methanol, and X-ray diffraction confirmed the established structure and its relative stereochemistry (Fig. 1 and 2).

Other already known quassinoids could be also identified: quassin (from fraction F3), picrasin H (from F4), picrasin B (from F5), and picrasin J (from F7). These compounds showed no significant antimalarial activity (D. Stien, S. Bertani, G. Bourdy, E. Deharo, E. Houe¨l, V. Jullian, A. Valentin, and S. Chevalley, presented at the ZingConference on Natural Products Chemistry, Antigua, 10 to 13 January 2008).

The antimalarial activity of SkE was determined on three strains of \textit{P. falciparum} and gave IC\(_{90}\) ranging from 24 to 68 nM (Table 1). When tested on highly synchronized parasite cultures, SkE had a maximal activity beginning at the second half of the erythrocytic cycle (Fig. 3). This peak decreased at the 40- to 48-h pulse for the lowest concentration tested (about 1/10 of the IC\(_{90}\)) (2).

SkE reduced gametocytemia by 50% at a dose sevenfold lower than that of the reference drug.
lower than primaquine, a reference molecule for the elimination of gametocytes (Table 1) (33).

The cytotoxicity of SkE varied, with an IC\textsubscript{50} from 6 \mu M (Vero cells) to 33 nM (THP1 cells) (Table 1).

SkE was then orally and intraperitoneally administered to \textit{P. vinckei petteri}-infected mice. The i.p. route (50\% effective dose [ED\textsubscript{50}], 0.5 mg/kg/day) was almost twice as effective as the p.o. route (ED\textsubscript{50}, 1 mg/kg/day), the control being chloroquine given i.p. (ED\textsubscript{50}, 3 mg/kg/day) (Table 2). The survival of mice was monitored for 3 weeks, and the mean survival time of the mice was evaluated (Table 3). The highest values were obtained with mice treated with SkE i.p. at 1 mg/kg/day (18.6 days; extended mean, 93.89 days) and with CQ at 10 mg/kg/day (19.20 days; extended mean, 43.87 days). Lower survival means were obtained with SkE given i.p. at a higher dose or by oral route. However, all the treated mice had a significantly higher survival time than that of control mice (\textit{P} \textless 0.05 for CQ given i.p. [1 mg/kg/day] and \textit{P} \textless 0.01 for the other treatments). The Kaplan-Meier curves (Fig. 4) showed similar aspects for SkE given 0.5 and 1 mg/kg/day i.p. and for CQ given 10 mg/kg/day i.p.

**DISCUSSION**

Biodiversity is clearly a source of new drugs (26). When biodiversity analysis combines with traditional treatments, there is the hope of finding some promising candidate molecules for pharmaceutical development. Here, we describe a new quassinoid, obtained after bioguided fractionation of a widely used Amazonian traditional remedy for malaria (35).

This quassinoid, named simalikalactone E, exhibited very good antiplasmodial activity against the three \textit{P. falciparum} strains tested, whatever their geographic origin or chloroquine sensitivity. The IC\textsubscript{50}s obtained were in the range of most commercially available antimalarial drugs tested under similar conditions (16)

<table>
<thead>
<tr>
<th>Treatment and route (a)</th>
<th>% Inhibition of parasitemia</th>
</tr>
</thead>
<tbody>
<tr>
<td>DMSO</td>
<td>0</td>
</tr>
<tr>
<td>CMC</td>
<td>0</td>
</tr>
<tr>
<td>CQ i.p.</td>
<td>0</td>
</tr>
<tr>
<td>1</td>
<td>0</td>
</tr>
<tr>
<td>5</td>
<td>100</td>
</tr>
<tr>
<td>10</td>
<td>100</td>
</tr>
<tr>
<td>SkE i.p.</td>
<td>59</td>
</tr>
<tr>
<td>0.5</td>
<td>98</td>
</tr>
<tr>
<td>1</td>
<td>100</td>
</tr>
<tr>
<td>5</td>
<td>100</td>
</tr>
<tr>
<td>p.o.</td>
<td>55</td>
</tr>
<tr>
<td>1</td>
<td>95</td>
</tr>
<tr>
<td>10</td>
<td>100</td>
</tr>
</tbody>
</table>

\(\text{a}\) The control mice treated with dimethyl sulfoxide alone showed 100\% parasitemia.
Against *P. falciparum* gametocytes, a stage which is fundamental for transmission to mosquitoes, SkE was more active (IC<sub>50</sub> 1.2 μM) than the reference compound primaquine (IC<sub>50</sub> 8.9 μM). In a previous study, Benoit-Vical et al. (3) showed that an artemisinin derivative, artesunate, inhibited gametocyte growth at a 10-fold-lower dose (around 0.1 μM).

Regarding cytotoxicity, the SkE IC<sub>50</sub>s were similar to those obtained against *P. falciparum* when using cancer-derived cell lines (MCF7 and THP1). However, when tested on Vero cells (which are of primate origin and are not cancer-derived cells) SkE displayed an IC50 around 100 times higher than that obtained against *P. falcipa* (which are of primate origin and are not cancer-derived cells).

In vivo, at day 5, the lowest tested doses (0.5 mg/kg/day i.p. and 1 mg/kg/day p.o.) were close to the ED<sub>50</sub>. Higher doses led to a complete cure of the mice at day 5 (Table 2). However, when the survival of mice was evaluated over a longer period, clear differences appeared between the i.p. and p.o. routes. The analysis of survival times showed that the i.p. route, at the same dose, seemed to be more efficient than the p.o. route as illustrated by the Kaplan-Meier curves (Fig. 4). It is to be noted that SkE administered by the i.p. route was considerably more active than CQ (about 10-fold more efficient when looking at SkE [1 mg/kg/day i.p.] and CQ [10 mg/kg/day i.p.], Table 3 and Fig. 4). On the other hand, at the higher i.p. dose (5 mg/kg/day), the survival time was shorter than for the other drugs, which could be explained by toxicity. The statistical analysis of the survival times (Table 3) showed better activity of SkE compared with the control, and at the doses tested, a better activity than that of CQ at 1 mg/kg/day, except for the highest dose (5 mg/kg/day, i.p. route) where a toxic effect was probably emerging. It is also to be noted that there were no significant differences between the SkE-treated mice given 1 and 0.5 mg/kg/day (i.p.) and the CQ-treated mice given 10 mg/kg/day (i.p.). Taken together, these data demonstrated that SkE showed good antimalarial activity in mice, with the best dose for a complete cure being around 1 mg/kg/day.

This activity was in the same range as in vivo activities previously reported for other quassinoids. When administered by the i.p. route, sergeolide (13), glucarubicinone (23), cedronine (24), and bruceolide and its carbonate derivatives (25) had ED<sub>50</sub>s between 0.2 mg/kg/day and 1.8 mg/kg/day. We also showed that SkD inhibits 50% of *Plasmodium yoelii yoelii* rodent malaria parasite growth at 3.7 mg/kg/day in vivo by the oral route (6). Toxicity was also monitored, and at the higher dose of SkE (5 mg/kg/day given i.p., 10 times the IC<sub>50</sub>), an immediate toxicity close to the LD<sub>50</sub> was observed (three deaths out of five mice in 3 days), while at lower doses, the antimalarial activity was clear.

The study of the activity of SkE on the different stages of the *P. falciparum* life cycle showed that SkE had a better inhibitory effect on stages where DNA synthesis occurred, but our results do not enable a distinction to be made between an SkE-DNA interaction and an inhibition of proteins implicated in DNA synthesis. DNA and protein synthesis inhibition in *P. falcipa* has been reported for several quassinoids. In general, the inhibition of DNA synthesis was less pronounced and seemed to be a consequence of protein synthesis inhibition (15, 21, 28). Recent reinvestigation of the antineoplastic activity of various quassinoids showed that NF-κB activation (9) and downregulation of c-myc (10) were implicated in the cell differentiation and apoptosis induced by quassinoids. Mitochondrial membrane depolarization and caspase 3 activation also played a role in this process (29).

**TABLE 3. Mean survival time of the treated mice and statistical significance**

<table>
<thead>
<tr>
<th>Treatment and route&lt;sup&gt;a&lt;/sup&gt;</th>
<th>No. of mice</th>
<th>Mean survival time (days) [95% CI]&lt;sup&gt;a&lt;/sup&gt;</th>
<th>Extended mean survival time (days)</th>
<th>Statistical significance (&lt;i&gt;P&lt;/i&gt; value) compared to the following group:</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>15</td>
<td>4.33 [3.76–4.91]</td>
<td></td>
<td>Control (i.p., 1 mg/kg/day)</td>
</tr>
<tr>
<td>CQ</td>
<td>5</td>
<td>7.40 [6.70–8.10]</td>
<td></td>
<td></td>
</tr>
<tr>
<td>i.p. 1</td>
<td>5</td>
<td>12.80 [11.51–14.09]</td>
<td>43.87</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>10</td>
<td>5</td>
<td>19.20 [16.82–21.58]&lt;sup&gt;d&lt;/sup&gt;</td>
<td></td>
<td>0.001</td>
</tr>
<tr>
<td>SkE i.p.</td>
<td>0.5</td>
<td>16.40 [11.45–21.35]&lt;sup&gt;c&lt;/sup&gt;</td>
<td>41.07</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>1</td>
<td>5</td>
<td>18.60 [14.39–22.81]&lt;sup&gt;c&lt;/sup&gt;</td>
<td>93.89</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>5</td>
<td>5</td>
<td>11.40 [4.51–18.29]&lt;sup&gt;c&lt;/sup&gt;</td>
<td>20.57</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>p.o. 1</td>
<td>5</td>
<td>11.00 [6.46–15.54]&lt;sup&gt;c&lt;/sup&gt;</td>
<td>13.61</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>10</td>
<td>5</td>
<td>12.40 [8.42–16.38]&lt;sup&gt;c&lt;/sup&gt;</td>
<td>15.01</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>20</td>
<td>5</td>
<td>13.80 [9.32–18.28]&lt;sup&gt;c&lt;/sup&gt;</td>
<td>16.41</td>
<td>&lt;0.001</td>
</tr>
</tbody>
</table>

<sup>a</sup> Treatment and routes were as follows: chloroquine (CQ) (1, 5, or 10 mg/kg/day for 4 days); simalikalactone E (SkE) (0.5, 1, 5, 10 or 20 mg/kg/day); p.o., oral route; i.p., intraperitoneal route.

<sup>b</sup> 95% CI, 95% confidence interval.

<sup>c</sup> As some mice were still alive at the end of the study, the largest observed analysis time was not available, and therefore, the mean is underestimated.

<sup>d</sup> NS, not significant.

and varied from 24 to 68 nM. These data clearly indicate that SkE does not interfere with CQ resistance pathways.
nylchaparrin were also identified as inhibitors of the transcription factor AP-1, but this function did not correlate with cytotoxicity or protein synthesis inhibition (7). Those new findings suggest that the antiplasmodial mechanism of action of quassinoids merits further detailed investigation.

The structural requirements for the antimalarial activity of quassinoids are well-documented, and both SkD and SkE meet them: they have an α,β-unsaturated lactone on ring A and an oxymethylene bridge between C-8 and C-11 or between C-8 and C-13. The isolation of SkE allowed us to compare the antiplasmodial potencies of SkE and SkD and thus evaluate the effect of the carboxylate group in the C-6 position. C-6 substitution occurs sometimes in quassinoids. In a review describing 230 quassinoids, 10% were shown to be substituted on C-6, and hydroxy or carboxy groups were the only substituents reported (11). For one of these quassinoids, 15-desacylun-dulatone, isolated from Quassia undulata (1) and Hannoa chlorantha (14), a good antiplasmodial activity was reported. However, there is no clear evidence in the literature of the influence of a substituent on the C-6 position on antiplasmodial activity.

The present report demonstrated that despite their reputation as toxic molecules, quassinoids remain potentially interesting as antimalarials, and further research should be done on rationalizing the effect of the C-6 substitution to improve their efficacy as drugs and lower their toxicity. Because quassinoids are the active ingredients of many traditional antimalarial preparations all over the world, this type of research would be of great interest for people living in places where malaria is endemic and relying on these preparations.

REFERENCES