The coagulase-negative staphylococci (CoNS) are a heterogeneous group of at least 15 different species of Gram-positive bacteria that have emerged in recent decades as important nosocomial pathogens (10, 31). A particularly problematic species is *Staphylococcus epidermidis*, which is responsible for a growing number of infections among hospital patients with compromised immune systems and is especially notorious for forming biofilms that adhere to surgical equipment and other hospital surfaces and indwelling devices (3, 18). Methicillin was traditionally the first-line antibiotic against CoNS, but its widespread use has resulted in resistance in 50% to 80% of CoNS infections and 75 to 90% of nosocomial *S. epidermidis* infections (18). As a result, vancomycin is now the first-line agent for treating CoNS infections; however, isolates with reduced susceptibility to vancomycin have also been observed (11, 26), and the emergence of enterococci harboring mobile elements that confer vancomycin resistance has raised concerns that resistance was generally correlated with the absence of the previously identified Pro residue, several cases were identified where additional factors also appear to contribute.

The aryomycins are a class of natural-product antibiotics that act via the inhibition of type I signal peptidase (SPase), and we have found in diverse bacteria that their activity is limited by the presence of a resistance-conferring Pro residue in SPase that reduces inhibitor binding. We have also demonstrated that *Staphylococcus epidermidis*, which lacks this Pro residue, is extremely susceptible to the aryomycins. Here, to further explore the potential utility of the aryomycins, we report an analysis of the activity of a synthetic aryomycin derivative, aryomycin C16, against clinical isolates of *S. epidermidis* and other coagulase-negative staphylococci (CoNS) from distinct geographical locations. Against many important species of CoNS, including *S. epidermidis*, *S. haemolyticus*, *S. lugdunensis*, and *S. hominis*, we find that aryomycin C16 exhibits activity equal to or greater than that of vancomycin, the antibiotic most commonly used to treat CoNS infections. While the susceptibility was generally correlated with the absence of the previously identified Pro residue, several cases were identified where additional factors also appear to contribute.

In *In Vitro* Activities of Aryomycin Natural-Product Antibiotics against *Staphylococcus epidermidis* and Other Coagulase-Negative Staphylococci

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The aryomycins are a class of natural-product antibiotics that act via the inhibition of type I signal peptidase (SPase), and we have found in diverse bacteria that their activity is limited by the presence of a resistance-conferring Pro residue in SPase that reduces inhibitor binding. We have also demonstrated that *Staphylococcus epidermidis*, which lacks this Pro residue, is extremely susceptible to the aryomycins. Here, to further explore the potential utility of the aryomycins, we report an analysis of the activity of a synthetic aryomycin derivative, aryomycin C16, against clinical isolates of *S. epidermidis* and other coagulase-negative staphylococci (CoNS) from distinct geographical locations. Against many important species of CoNS, including *S. epidermidis*, *S. haemolyticus*, *S. lugdunensis*, and *S. hominis*, we find that aryomycin C16 exhibits activity equal to or greater than that of vancomycin, the antibiotic most commonly used to treat CoNS infections. While the susceptibility was generally correlated with the absence of the previously identified Pro residue, several cases were identified where additional factors also appear to contribute.

The aryomycins (Fig. 1) are a novel class of natural-product antibiotics that act by inhibiting bacterial type I signal peptidase (SPase) (19, 25). SPase is a Ser-Lys dyad protease that removes N-terminal signal sequences from preproteins following their translocation across the cytoplasmic membrane (5, 20). SPase is an attractive target for antibiotic therapy because it is conserved, essential, and located in the relatively accessible outer leaflet of the cytoplasmic membrane. Furthermore, because bacterial SPase acts via a catalytic mechanism that is distinct from that of its eukaryotic homologues, the aryomycins are unlikely to exhibit mechanistic toxicity in humans (5, 20). Despite the apparent accessibility, essentiality, and conservation of SPase, initial reports suggested that the aryomycins were active against only a few Gram-positive bacteria, including *Streptococcus pneumoniae*, *Rhodococcus opacus*, and *Brevibacillus brevis* (15, 25), and not against other important Gram-positive pathogens or against any Gram-negative bacteria. However, after reporting the first synthesis of an aryomycin, aryomycin A4, as well as the synthetic derivative aryomycin C16 (Fig. 1), we found that each potently inhibits the growth of *S. epidermidis* (24) and that *S. epidermidis* evolves resistance to the aryomycins by mutating residue 29 of one of its two SPases, SpSIB, from Ser (Ser29) to Pro (Pro29) (29). Moreover, a Pro residue is naturally present at the analogous position in *S. epi*dermidis, *S. haemolyticus*, *S. lugdunensis*, and *S. hominis*. However, while resistance in *S. epidermidis* is associated with the absence of the previously identified Pro residue, some cases were identified where additional factors also appear to contribute.

The aryomycins (Fig. 1) are a novel class of natural-product antibiotics that act by inhibiting bacterial type I signal peptidase (SPase) (19, 25). SPase is a Ser-Lys dyad protease that removes N-terminal signal sequences from preproteins following their translocation across the cytoplasmic membrane (5, 20). SPase is an attractive target for antibiotic therapy because it is conserved, essential, and located in the relatively accessible outer leaflet of the cytoplasmic membrane. Furthermore, because bacterial SPase acts via a catalytic mechanism that is distinct from that of its eukaryotic homologues, the aryomycins are unlikely to exhibit mechanistic toxicity in humans (5, 20). Despite the apparent accessibility, essentiality, and conservation of SPase, initial reports suggested that the aryomycins were active against only a few Gram-positive bacteria, including *Streptococcus pneumoniae*, *Rhodococcus opacus*, and *Brevibacillus brevis* (15, 25), and not against other important Gram-positive pathogens or against any Gram-negative bacteria. However, after reporting the first synthesis of an aryomycin, aryomycin A4, as well as the synthetic derivative aryomycin C16 (Fig. 1), we found that each potently inhibits the growth of *S. epidermidis* (24) and that *S. epidermidis* evolves resistance to the aryomycins by mutating residue 29 of one of its two SPases, SpSIB, from Ser (Ser29) to Pro (Pro29) (29). Moreover, a Pro residue is naturally present at the analogous position in the homologous SPases of the pathogens *Staphylococcus aureus*, *Escherichia coli*, and *Pseudomonas aeruginosa*, and we showed that it imparts resistance by reducing the affinity with which the aryomycins bind. Furthermore, we found that a remarkably diverse range of both Gram-positive and Gram-negative bacteria whose SPases lack a Pro at the analogous position are susceptible to the aryomycins, including *Staphylococcus epidermidis*, *Streptococcus pyogenes*, *Helicobacter pylori*, *Chlamydia trachomatis*, and some strains of *Francisella tularensis* (29). In total, the results suggest that the identified SPase polymorphism is a major contributor to naturally occurring aryomycin resistance. However, we also showed that *Yersinia pestis* and some strains of *S. aureus* are susceptible to the aryomycins despite the presence of an analogous Pro, while others, such as many of the *Lactobacillales*, *Clostridia*, and *Bacteroidetes*, are resistant despite its absence, implying that in some cases, susceptibility must depend on additional factors, such as variable levels of toxicity associated with the inhibition of protein secretion.

The potent activity of the aryomycins against a strain of *S.
epidermidis (RP62A) suggests that they might be useful in the treatment of this and perhaps other CoNS. Here, to examine the spectrum of activity of the aryloymcin against clinical isolates of S. epidermidis and other CoNS, we report the activity of aryloymcin C16 against two panels of isolates from hospitals in geographically diverse locations and compare the activity to that of vancomycin. The results reveal that the aryloymcins have potent antibacterial activity against a range of important CoNS species whose SpsIB orthologs lack the previously identified resistance-conferring Pro, while less activity is observed against species where Pro is present. While we generally observed similar susceptibilities for different isolates within a species, significant differences were observed in several cases, with one atypical instance of susceptibility resulting from the presence of a Ser in place of the resistance-conferring Pro. Significant differences in susceptibility between isolates of the same species are usually observed with clinically deployed antibiotics where selection for resistance has occurred during therapy (2, 9, 14, 21), and therefore, these results may be relevant to understanding the natural evolution of aryloymycin resistance in nature.

MATERIALS AND METHODS

A total of 282 nonduplicate, clinical isolates of CoNS, identified to species level, were obtained from the London Health Services Centre, London, Ontario, Canada (13). Of these, 143 isolates were S. epidermidis, while the remaining 139 consisted of 12 groups comprised of various numbers of isolates of Staphylococcus capitis, Staphylococcus caprae, Staphylococcus cohnii, Staphylococcus haemolyticus, Staphylococcus lugdunensis, Staphylococcus saprophyticus, Staphylococcus simulans, Staphylococcus warneri, and Staphylococcus hominis. This panel of CoNS is a subset of a larger panel whose susceptibility to quinupristin-dalfopristin, linezolid, telithromycin, and vancomycin has been reported (13). A second set of nonduplicate clinical isolates of CoNS not identified to species level was obtained from the Department of Microbiology, University of California San Diego Medical Center Hillcrest, San Diego, CA. These strains were collected from patients with infections of various coagulase-negative bacteria between April and June of 2008.

Aryloymycin C16 was synthesized as described previously (24). The MICs of aryloymycin C16 were determined for isolates from both sets of CoNS using a modified CLSI agar dilution method. Bacterial inocula were prepared from log-phase cultures or from suspensions of colonies grown on solid medium, diluted appropriately, and spotted onto tryptic soy agar at a concentration of 1 × 10^4 CFU/spot. Spots were then incubated for 24 h at 37°C, and the MIC was defined as the concentration at which there was no visible growth. MICs of vancomycin were also determined, in an identical manner, to provide a reference for the aryloymycin C16 susceptibilities ranging from highly susceptible to vancomycin, both of which we found to be 2 μg/ml (24) and compare favorably to the MIC50 and MIC90 of vancomycin, both of which we found to be 2 μg/ml, in agreement with values reported previously (13). Additionally, the range of MICs observed among the 143 isolates was narrow, with the most resistant isolate having an aryloymycin C16 MIC of 4 μg/ml.

The remaining 9 species of CoNS examined displayed aryloymycin C16 susceptibilities ranging from highly susceptible to resistant (Table 1). Specifically, we found that S. haemolyticus, S. lugdunensis, and S. hominis are highly susceptible to aryloymycin C16, with MIC50 and MIC90 values of 0.25 to 2 μg/ml. In contrast, we found that S. capitis, S. caprae, and S. cohnii are only moderately susceptible, with MIC50 values between 8 and 16 μg/ml and MIC90 values between 16 and 32 μg/ml, and that S. warneri and S. saprophyticus are resistant, with MIC50 values equal to or greater than 64 μg/ml and MIC90 values of more than 64 μg/ml. Finally, while many S. simulans isolates were sensitive to the aryloymycins (MIC50 of 2 μg/ml), a significant number of isolates were highly resistant, resulting in a MIC90 of ≥64 μg/ml.

Under laboratory conditions, S. epidermidis RP62A evolves aryloymycin resistance by mutation of Ser29 to Pro29 in SpsIB, one of two active SPases in this organism; in the related organism S. aureus, as well as in the more distantly related E. coli and P. aeruginosa, an analogous Pro29 is naturally present and contributes significantly to aryloymycin resistance (29) (here and throughout the remainder of the manuscript, unless otherwise indicated, Ser29 and Pro29 refer to the residue in SpsIB orthologs at the position corresponding to 29 in S. epidermidis).
In contrast, we did not observe mutations in the second \textit{S. epidermidis} SPase gene, \textit{spsI}, in any of the resistant isolates. To determine whether similar mutations might be responsible for the differences in arylomycin susceptibility observed among the different CoNS, we examined the sequences of their \textit{spsI} and \textit{spsIB} genes (Fig. 2; also see Table S3 in the supplemental material). To identify SPase sequences, we used the NCBI BLAST interface to search the sequenced genomes of CoNS (see the supplemental material). Overall, the data support the model that Pro29 contributes significantly to arylomycin resistance within CoNS. \textit{S. lugdunensis}, \textit{S. hominis}, and a significant percentage of \textit{S. simulans} isolates are extremely susceptible, and these species lack \textit{Pro29}. In contrast, the CoNS species most resistant to the arylomycins, \textit{S. warneri} and \textit{S. saprophyticus}, each harbor an SPase with Pro29. The correlation between Pro29 and arylomycin susceptibility is less clear for species that display intermediate susceptibilities; for example, while \textit{S. caprae} and \textit{S. capitis} have Pro29, \textit{S. cohnii} has Ser29, all three display MIC50 values between 4 and 16 \textmu{g}/ml.

Most of the species examined have the narrow range of susceptibilities to arylomycin \textit{C16} that is expected for an antibiotic that has not been used clinically (2, 9, 14, 21) (Table 1). However, for several species, we found a greater range in susceptibilities or we found outliers with susceptibilities that differed significantly from those of the other isolates (Table 1; also see Fig. S3 in the supplemental material). For example, one isolate of \textit{S. lugdunensis} is significantly more susceptible than the species average (its MIC is 8-fold below the MIC50), while a second isolate is unusually resistant (its MIC is 128-fold above the MIC50). Sequencing of the \textit{spsIB} genes revealed identical sequences at the amino acid level (including at residue 29) (Fig. S4 in the supplemental material). In addition, a bimodal distribution of MICs was observed for isolates of \textit{S. warneri} and \textit{S. simulans}, with peaks at 16 \textmu{g}/ml and >64 \textmu{g}/ml and at 2 \textmu{g}/ml and >64 \textmu{g}/ml, respectively (see Fig. S3 in the supplemental material). Sequencing the \textit{spsIB} genes from representative isolates of each group revealed nearly identical SPase sequences and no variation at residue 29 (see Fig. S4 in the supplemental material).
the supplemental material). Finally, we identified a particularly susceptible isolate of S. warneri, with a MIC that is 32-fold below the species MIC<sub>50</sub>. Interestingly, the particularly susceptible strain of S. warneri had Ser<sup>29</sup> as opposed to Pro<sup>29</sup>, which is typically present in S. warneri isolates, further supporting the genotype/phenotype correlation.

The observed differences in MICs that cannot be accounted for by differences in SpSIP sequence, both within and between species, suggest that additional factors contribute to arylomycin susceptibility. To determine whether these additional factors are shared among related organisms, we examined MICs as a function of phylogeny (Fig. 2). The phylogenetic analysis, based on four highly conserved genes (see Materials and Methods), suggests that S. simulans was the earliest species to diverge from the common ancestor of the examined CoNS, with the other species forming two groups, one containing S. xylosus, S. saprophyticus, and S. cohnii and the second containing the remaining species. Notably, SpSIP SPases have only been shown to be present in the second group, although their presence cannot be ruled out in the unsequenced species.

S. saprophyticus, which has Pro<sup>29</sup> and is extremely resistant to the arylomycins (MIC<sub>50</sub> $>$ 64 μg/ml), is most closely related to S. cohnii, which lacks Pro<sup>29</sup>, and yet is only moderately susceptible to the arylomycins (MIC<sub>50</sub> = 8 μg/ml). To test whether other species that are closely related to S. saprophyticus and S. cohnii share this intrinsically lower susceptibility, we determined the MIC of a typed strain of S. xylosus, which is closely related to S. saprophyticus but, like S. cohnii, lacks Pro<sup>29</sup>. As expected, the strain of S. xylosus is moderately susceptible to the arylomycin (MIC = 4 μg/ml). Thus, this group of related bacteria appears to have a lower basal level of arylomycin susceptibility. In contrast, S. epidermidis, S. hominis, S. lugdunensis, and S. haemolyticus, which lack Pro<sup>29</sup> and display extreme arylomycin susceptibility (MIC<sub>50</sub> values of 0.25 to 2 μg/ml), as well as S. capitis and S. caprae, which have Pro<sup>29</sup> but remain moderately susceptible (MIC<sub>50</sub> values of 8 to 16 μg/ml), are more related to one another than to the other CoNS species examined. Despite the fact that some S. warneri isolates are extremely resistant, the remaining isolates have MICs of ~16 μg/ml, which is consistent with the level of resistance observed for the other CoNS with Pro<sup>29</sup> within the more susceptible phylogenetic group.

Finally, we evaluated the activity of arylomycin C<sub>16</sub> against strains from a CoNS panel that has not been identified to the species level (obtained from the University of California San Diego Medical Center) and observed MIC<sub>50</sub> and MIC<sub>90</sub> values of 2 μg/ml and $>$32 μg/ml, respectively (Fig. 3). In addition to revealing a portion of resistant isolates, the distribution of MICs shows significant variations in the susceptibility of the remaining strains. Although interpretation of these results is complicated by the lack of species identification, the MIC distribution is consistent with a model in which both the presence of Pro<sup>29</sup> and a second, yet-to-be-determined factor combine to yield tiers of arylomycin susceptibility.

**DISCUSSION**

Members of the arylomycin class of natural-product antibiotics act via a novel mechanism of action, the inhibition of SPase and, thus, the inhibition of the essential process of protein secretion. We found that, in addition to excellent activity against S. epidermidis (24), arylomycin C<sub>16</sub> has potent activity against S. haemolyticus, S. lugdunensis, and S. hominis. Each of these members of the CoNS has an SPase with Ser<sup>29</sup>, consistent with the central role of this residue in determining arylomycin susceptibility that we observed previously with other bacteria (29).

The remaining species examined range from only moderately susceptible to extremely resistant, and Pro<sup>29</sup> clearly makes a significant contribution to resistance in many of these cases. Interestingly, phylogenetic analysis suggests that Ser<sup>29</sup> was prevalent during speciation but that Pro<sup>29</sup> evolved in at least two independent instances, once in S. saprophyticus and once in the common ancestor of S. caprae and S. capitis (Fig. 2). Pro<sup>29</sup> in S. warneri may represent a third instance, although it might instead result from common ancestry with S. caprae and S. capitis; this distinction is beyond the resolution of the current analysis. Interestingly, many synonymous and nonsynonymous mutations differentiate the spsIB genes of the different staphylococcal species, suggesting that ample mutational diversity has been sampled, but only Ser and Pro appear to be tolerated at position 29. Importantly, the multiple independent instances where Pro<sup>29</sup> was introduced during staphylococcal speciation suggest that there may be a natural selective pressure for Pro<sup>29</sup>, and the variation within isolates of an extant species suggests that this pressure may have occurred recently and may even still be present. While the nature of the selection pressure is currently unknown, it is tempting to consider that it may be related to the presence of arylomycins in nature (29).

The observed differences in susceptibilities between some of the staphylococcal species reveal that, in addition to SpSIP residue 29, there must be other factors that also contribute to overall susceptibility, and these other factors generally appear to be shared among closely related species, resulting in groups of species having higher or lower basal susceptibilities. One possibility is that the presence of a second SPase gene, orthologs of S. epidermidis spsI, in some species of CoNS could contribute to their different arylomycin susceptibilities. In principle, expressing two arylomycin-sensitive SPase proteins could result in hypersusceptibility if each SPase recognized a different subset of essential preprotein substrates. Alternatively, expressing one sensitive and one resistant SPase could result in either a susceptible or resistant phenotype, depending on the exact mechanism of arylomycin-induced cell death, which is yet...
to be determined. Interestingly, all of the species with higher basal susceptibilities have two SPases, while *S. xylosus* (S. Le-roy, personal communication) and *S. saprophyticus* have only one SPase and display lower basal susceptibilities. Consistent with a possible role for a second SPase contributing to arylomycin susceptibility, *S. aureus* strain 8325 is highly resistant to arylomycin (MIC > 128 μg/ml) (24) despite being closely related to the more susceptible group of CoNS, and this species has only a single SPase, apparently due to a recent deletion of its *spaI* homologue. Further experiments will be required to elucidate the potential role of multiple SPases in CoNS sus-ceptibilities and/or to identify other factors contributing to susceptibility, such as differences in the composition of secreted proteins or the presence of modifying enzymes. Comparison of resistant and susceptible isolates of the same species may provide a valuable approach to identifying additional factors that contribute to arylomycin susceptibility.

Regardless of the origins of arylomycin susceptibility and resistance, it is clear that a broad and clinically important (16) range of CoNS strains are susceptible to the arylomycins. In fact, in these cases, the level of susceptibility compares favorably with the level of susceptibility to vancomycin, the antibiotic currently recommended for treatment of CoNS infections but for which resistance is a major concern. The availability of a novel antibiotic, with a novel mechanism of action, would significantly improve CoNS therapy. Along with previously re-ported data (29), the data also suggest that if the arylomycin class of natural products could be optimized to overcome the loss in affinity mediated by the presence of the resistance-confering Proβ, then they would have broad activity against CoNS and, possibly, other bacteria as well.

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