Danoprevir Monotherapy Decreases Inflammatory Markers in Patients with Chronic Hepatitis C Virus Infection

Caralee J. Schaefer,1 Karl Kossen,1 Sharlene R. Lim,1 Jiing-Huey Lin,1 Lin Pan,1 Williamson Bradford,1 Patrick F. Smith,2 and Scott D. Seiwert1,*

InterMune, Inc., 3280 Bayshore Blvd., Brisbane, California 94005,1 and Roche-Genentech, 1 DNA Way, South San Francisco, California 940802

Received 31 January 2011/Returned for modification 17 March 2011/Accepted 6 April 2011

Danoprevir is a potent and selective direct-acting antiviral agent that targets the protease activity of hepatitis C virus (HCV) NS3/4A. This agent results in a significant rapid decline in HCV RNA levels when it is used in monotherapy. The present study evaluated whether plasma concentrations of the inflammatory markers gamma interferon-inducible protein 10 (IP-10) and neopterin or the interferon-stimulated gene product 2′-5′-oligoadenylate synthetase (OAS-1) were correlated with the plasma HCV RNA concentration before or during 14-day danoprevir monotherapy. In contrast to pegylated interferon and ribavirin treatment, a higher baseline IP-10 concentration was positively correlated with a greater first-phase HCV RNA decline upon danoprevir administration. Changes in the IP-10 plasma concentration during danoprevir administration were also associated with categorical changes in HCV RNA concentration at days 7 and 14. The neopterin concentration appeared to be moderately decreased during danoprevir administration, although these changes were not statistically significant. However, changes in neopterin concentration showed a statistically significant correlation with changes in IP-10 concentration. Considerable variation in the OAS-1 concentration was observed before and during treatment, including in patients treated with placebo and/or patients with minimal virologic response. Overall, these results suggest that effective treatment with a direct-acting antiviral agent may reduce hepatic inflammation and that first-phase HCV RNA decline during treatment with an NS3/4A protease inhibitor is more robust in patients with high baseline IP-10 concentrations.

Hepatitis C virus (HCV) is a positive-strand RNA virus belonging to the Flaviviridae family. Approximately 170 million people worldwide suffer from chronic HCV infection (2, 47), which is a major cause of chronic liver disease, cirrhosis, and primary hepatocellular carcinoma and is currently among the leading indications for liver transplantation in both the United States and Europe (1, 45). Progression to liver cirrhosis/fibrosis may be asymptomatic and can evolve over many years (4). The current standard of care (SOC) for chronic HCV infection is a 24- to 48-week regimen comprised of weekly subcutaneous injections of pegylated alpha interferon (PEG-IFN) and twice daily (BID) oral ribavirin treatment. This regimen leads to a sustained virologic response (SVR) in approximately half of the patients infected with HCV genotype 1 (5, 13).

Chronic HCV infection is characterized by persistent inflammation of hepatic lobules and the presence of CXCR3-positive T cells (21, 56). The extent of cellular infiltration correlates with stage of disease (54, 55). The majority of these cells are not HCV specific and not competent to clear virus (28). However, they contribute to a persistent inflammatory process which is thought to contribute to cell death, liver damage, and the advent and progression of hepatic fibrosis (28, 56).

Biomarkers that predict antiviral treatment response or correlate with the extent of ongoing inflammation or fibrosis would be useful to monitor the disease state in patients with chronic HCV infection. Several biomarkers have recently been evaluated in the context of HCV infection, including interferon-responsive genes (26, 30, 37, 40, 51, 53), chemokines (28, 29, 31, 54, 56), interleukin-28B (IL-28B) gene polymorphisms (16, 18, 39), fibrinogen-like protein 2 (11), complement component C3a (25), and osteopontin (23).

Several of the biomarkers evaluated in patients with HCV infection are associated with an interferon-mediated inflammatory response. One such biomarker is gamma interferon (IFN-γ)-inducible protein 10 (IP-10 or CXCL10), a chemokine produced by a variety of cells, including hepatocytes in areas of liver inflammation (21). IP-10 is a chemoktractant for CXCR3 receptor-expressing cells, including monocytes, natural killer cells, and T cells, and has been proposed to recruit activated T cells to hepatic lesions in chronic viral hepatitis (34, 41, 49, 55).

Patients with chronic HCV infection display elevated plasma concentrations of IP-10 (5, 8, 27, 33) which correlate with the degree of liver inflammation and fibrosis (28, 42, 54, 55). Elevated IP-10 mRNA expression in HCV-infected liver tissue also correlates with the accumulation of CXCR3-expressing T cells in the liver as well as histological fibrosis scores (21). Notably, a low baseline IP-10 concentration predicts a robust first-phase decline in HCV RNA in response to SOC (3), and the plasma IP-10 concentration is inversely correlated with rates of SVR (3, 5, 8, 9, 27).

Neopterin is associated with activation of the cellular im-
nune system and is used as a marker of inflammation. Neopterin is released by monocytes/macrophages after activation by IFN-γ, and its concentration is elevated in several disease states, including bacterial and viral infections and autoimmune disorders (20, 24, 36, 52). Neopterin plasma concentrations are increased during HCV infection (10, 17, 19, 36, 52).

2′-5′-Oligoadenylate synthetase (OAS-1) is an interferon-stimulated gene product that is responsible for activation of the RNase L pathway in response to viral RNA. OAS-1 binds to double-stranded RNA and catalyzes the formation of 2′-5′-linked oligoadenylate, which in turn activates RNase L, an endogenous protein that degrades viral RNA, thereby inhibiting viral protein synthesis and replication (26, 32, 48).

Several studies have reported increased OAS-1 expression through chronic HCV infection (26, 38, 48). Similar to the case with IP-10 (3, 5, 8, 9, 27), the baseline OAS-1 concentration is higher in nonresponders (NRs) to SOC treatment than in those that achieve SVR (48).

Danoprevir (ITMN191/RG7227) is a highly potent and selective macrocyclic, peptidomimetic inhibitor of the HCV NS3/4A serine protease (46). Interim analysis of a phase 2b program in combination with SOC demonstrated rapid virologic response (RVR) and early virologic response (EVR) rates of up to 86% and 92%, respectively (50). In a first-of-its-kind study combining two orally active direct-acting antiviral agents in patients with chronic HCV infection, treatment with the combination of danoprevir and the NSSP polymerase inhibitor mericitabine (RG7128) for 13 days resulted in up to 62% (5/8) of patients having HCV loads below the limit of detection (15). Danoprevir is currently in clinical development as a ritonavir-boosted agent in both types of regimens.

In early-stage clinical trials, danoprevir monotherapy resulted in rapid, dose-dependent reductions in HCV RNA levels. When danoprevir was administered for 14 days as monotherapy, the best-performing treatment-naive (TN) cohort experienced median maximal and median end-of-treatment (EOT) viral load reductions of −3.9 and −3.8 log10 IU/ml, respectively. A single cohort of NRs to prior SOC treatment experienced more muted changes on these endpoints of −2.9 and −2.5 log10 IU/ml, respectively. The overall rate of viral rebound was low (10/37) and similar in TN (8/29) and NR (2/8) cohorts. Median viral response profiles for continuous decline and rebound patients showed nearly coincident median viral response profiles through day 7 that diverged thereafter, suggesting relatively late emergence of viral resistance (44).

The current study examined plasma concentrations of IP-10, neopterin, and OAS-1 in samples collected during the above-mentioned 14-day multiple-ascending-dose study of danoprevir monotherapy in patients chronically infected with HCV genotype 1. IP-10 and neopterin were examined, as they are associated with HCV-induced liver inflammation. As a prototypical interferon-stimulated gene, OAS-1 was used to examine the hypothesis that NS3/4A can dampen viral sensing and interferon production (14, 43).

**MATERIALS AND METHODS**

**Danoprevir monotherapy phase 1 clinical trial.** Details of a 14-day, randomized, double-blind, placebo-controlled study evaluating danoprevir monotherapy in patients chronically infected with HCV genotype 1 have recently been published (12). The first part of this study evaluated multiple ascending doses of danoprevir in treatment-naive patients. Patients in this portion of the study received oral danoprevir at doses of 100 mg BID (n = 8), 100 mg three times day (TID) (n = 8), 200 mg BID (n = 5), or 200 mg TID (n = 8) or placebo equivalent (n = 8). Fewer patients were in the 200-mg BID cohort, as three patients were incorrectly dosed and excluded from all data analyses (including those of viral kinetics and biomarkers). The second part of the study examined a single dose level (300 mg BID, n = 8) or placebo (n = 2) in a cohort of nonresponders to prior PEG-IFN–ribavirin therapy. Of the patients described above, 4/17 were included in the biomarker analysis. One patient (receiving 100 mg BID) was excluded from biomarker analysis due to the lack of baseline biomarker data. A second excluded patient (a TN patient receiving placebo) was omitted due to the lack of viral load determination at baseline.

The clinical study was conducted in full accordance with the 1996 Declaration of Helsinki. The study protocol was reviewed and approved by the independent ethics committee at each participating clinical research facility, and written informed consent was obtained from each patient or legal guardian prior to screening for study eligibility.

**Viral kinetics and virologic fate.** The antiviral activity of danoprevir was assessed by measuring plasma HCV RNA at scheduled intervals from baseline to day 14 using a Cobas Ampliprep/Cobas TaqMan HCV assay (Roche Molecular Diagnostics, Pleasanton, CA). Viral kinetics were calculated and represented as the median log change from baseline (day 0). Virologic response patterns (rebound, plateau, continuous decline) were determined in danoprevir-treated patients by comparison of the EOT viral load relative to the nadir viral load using the following definitions. Rebound patients (n = 10) were defined as those with an increase from the nadir of ≥1.0 log10 IU/ml. Plateau patients (n = 13) were defined as those with an increase from the nadir of <1.0 log10 IU/ml. All other patients were classified as having a continuous decline (n = 14).

**Biomarker analysis.** Biomarkers were assessed in plasma samples collected prior to treatment on day 0 (baseline sample) and on day 7 and day 14.

**Plasma IP-10 concentration.** Plasma IP-10 concentration was measured in duplicate at Southern Research (Birmingham, AL) using a Quantikine human CCL10/IP-10 immunoassay kit (R&D Systems, Minneapolis, MN), according to the manufacturer’s instructions. Average values were reported as picograms per milliliter. On the basis of the manufacturer’s specifications, the mean minimal detection limit of this kit is 1.7 pg/ml (range, 0.4 to 4.5 pg/ml).

**Plasma neopterin concentration.** Plasma neopterin concentration was measured in duplicate at Southern Research (Birmingham, AL) using human neopterin enzyme immunoassay kits purchased from Alpco Diagnostics (Salem, NH), according to the manufacturer’s instructions. Average values were reported as nanomoles per liter (nM). On the basis of the manufacturer’s specifications, the assay has a range of 2 to 250 nM.

**Plasma OAS-1 concentration.** Plasma OAS-1 levels were measured in triplicate at Southern Research (Birmingham AL) using human 2′-5′-oligoadenyl-5′-triphosphate (2-5A) radioimmunoassay (RIA) kits (Eiken Chemical Company, Tokyo, Japan), according to the manufacturer’s instructions. Briefly, OAS-1 in the samples was bound to a poly(I)·poly(C) agarose gel, and the bound complex was then separated from other serum components by centrifugation. The bound, partially purified OAS-1 was allowed to degrade the production of 2-5A. The amount of 2-5A produced, which is proportional to the OAS-1 enzyme activity, was quantified in a competitive RIA using 121I-labeled 2-5A. In this assay, newly formed 2-5A competed with 121I-labeled 2-5A for the binding sites on the anti-2-5A antibody. The antibody-bound 121I-labeled 2-5A was separated from unbound label by centrifugation, and bound radioactivity was quantified using a Packard Cobra 905 gamma counter (GMI, Ramsey, MN). Average values were reported as picomoles per liter (pM). On the basis of the manufacturer’s specifications, the kit has a range of 100 to 8,100 pM.

**Statistical analysis.** Statistical comparisons between groups were performed by 1-way analysis of variance (ANOVA), followed by Dunnett’s multiple comparison test, using either Prism software (GraphPad Software, La Jolla, CA) or JMP7 software (SAS Institute, Inc., Cary, NC). Correlations between continuous variables were assessed by linear regression using GraphPad Prism software.

**RESULTS**

**IP-10 concentrations.** At baseline, the median IP-10 plasma concentration for all patients was 421 pg/ml (see Fig. S1A in the supplemental material). Significant interpatient variability was noted, with a range of 113 to 4,004 pg/ml (35-fold). TN and NR patients showed median IP-10 values of 344 and 526 pg/ml, respectively. Baseline IP-10 concentrations of 3.9 and 2.5 log10 IU/ml.
787 pg/ml were observed in patients that experienced virologic continuous decline, plateau, and rebound, respectively (see Fig. S1B in the supplemental material). The range of IP-10 values in TN patients and NRs and in the continuous-decline, plateau, and rebound groups overlapped significantly (see Fig. S1A and B, respectively, in the supplemental material). Baseline values of IP-10 were not correlated with baseline HCV RNA concentrations across all patients ($r^2 = 0.01$, $P = 0.50$) or during the second phase of HCV RNA kinetics (defined as an HCV RNA concentration change between day 3 and EOT; $r^2 = 0.05$, $P = 0.21$).

Changes in IP-10 plasma concentration were also associated with categorical changes in HCV RNA concentration upon danoprevir administration (Fig. 2). At day 7 and day 14, patients that displayed a 1- to 2-log$_{10}$ IU/ml reduction in HCV RNA had median reductions in IP-10 of 46% and 43%, respectively (Fig. 2A and B, respectively). Patients with greater changes in HCV RNA concentration experienced marginally increased changes in IP-10 concentration; a 4- to 5-log$_{10}$ IU/ml reduction in HCV RNA was associated with median reductions in IP-10 of 65% at both day 7 and day 14 (Fig. 2A and B, respectively). In contrast, patients that experienced a $<1$-log$_{10}$ IU/ml reduction in HCV RNA (mostly placebo-treated patients) showed a more stable median IP-10 concentration at both time points (Fig. 2A and B). The association of changes in the concentrations of HCV RNA and IP-10 was similar in TN patients and NRs (compare circles and triangles in Fig. 2A and B).

The dynamics of IP-10 plasma concentration changes were distinct in patients experiencing a virologic continuous decline, plateau, or rebound during danoprevir administration (Fig. 3A to C, respectively). Continuous-decline patients showed strong first- and second-phase median HCV RNA declines, with large declines in median IP-10 concentration (Fig. 3A). Continuous-decline patients experienced median reductions of baseline IP-10 concentration of 57% and 60% at day 7 and day 14, respectively. All but one continuous-decline patient had a lower plasma IP-10 concentration at day 14 relative to that at baseline; the remaining patient experienced a 6% increase...
from a low baseline concentration (160 pg/ml). Plateau patients experienced a lesser first-phase HCV RNA decline without evidence of second-phase HCV RNA decline through day 14. The corresponding median IP-10 concentrations in this group were similarly reduced at day 7 and day 14 by 43% and 39%, respectively (Fig. 3B). All but one plateau patient had a lower plasma IP-10 concentration at day 14 relative to that at baseline; the remaining patient experienced a 10% increase from a relatively low baseline concentration (190 pg/ml). Rebound patients displayed a robust first-phase decline in HCV RNA concentration and a median decrease in plasma IP-10 concentration of 55% at day 7. The median IP-10 concentration at day 14 was reduced 71% relative to that at baseline. However, 3/9 rebound patients displayed considerably higher IP-10 concentrations at day 14 relative to the corresponding values at day 7. These three patients had 3 of the 4 lowest baseline IP-10 concentrations among patients experiencing a virologic rebound and the highest ratios of HCV RNA concentration at day 14 relative to that at baseline, and two showed an earlier inflection in HCV RNA concentration during danoprevir administration (data not shown).

Neopterin concentration. At baseline, the median neopterin plasma concentration for all patients was 7.6 nM, with a range of 4.5 to 21.3 nM (5-fold range; see Fig. S2A in the supplemental material). TN and NR patients showed median neopterin values of 7.6 and 7.7 nM, respectively. Median baseline neopterin concentrations of 8.7, 7.5, and 7.6 nM were observed in patients that experienced virologic continuous decline, plateau, and rebound, respectively (see Fig. S2B in the supplemental material). Baseline neopterin concentration was not correlated with baseline HCV RNA concentration across all patients ($r^2 = 0.04, P = 0.20$), in TN patients ($r^2 = 0.02, P = 0.44$), or in NRs ($r^2 = 0.18, P = 0.23$) (see Fig. S2C in the supplemental material).

In most patients, the plasma concentration of neopterin decreased as the HCV RNA concentration decreased. However, these changes were smaller and more variable than those observed with IP-10. At day 7, patients experiencing a 1- to 2-log$_{10}$-IU/ml reduction in HCV RNA concentration displayed a median reduction in neopterin concentration of 12% relative to that at baseline (Fig. 4A). Progressively larger median decreases of 16% and 29% were observed in patients experiencing 2- to 3- and 3- to 4-log$_{10}$-IU/ml reductions in HCV RNA concentration, respectively. Neopterin concentrations in the two patients experiencing the greatest reduction in HCV RNA concentration (4 to 5 log$_{10}$ IU/ml) were either stable (5% reduction) or slightly higher (22% increase) than their corresponding baseline values. Similarly modest and variable reductions in median neopterin concentration were observed at day 14 (Fig. 4B).

As with IP-10, the dynamics of neopterin plasma concentrations were examined separately in patients experiencing virologic continuous decline, plateau, or rebound during danoprevir administration (Fig. 5A to C, respectively). Similar and modest reductions in median neopterin concentrations were observed in patients comprising the continuous-decline, plateau, and rebound groups at both day 7 (12.3%, 4.3%, and 25%, respectively; Fig. 5A to C) and day 14 (8.0%, 5.6%, and 7.9%, respectively; Fig. 5A to C).

Correlation of IP-10 and neopterin concentrations. The majority of danoprevir-treated patients experienced a reduction in IP-10 plasma concentration and a lesser reduction in neopterin plasma concentration. Therefore, the data were examined to determine whether changes in IP-10 and neopterin concentrations were correlated at the patient level. A positive correlation was observed between neopterin concentration changes and IP-10 concentration changes at both day 7 ($r^2 = 0.35, P <$
The slopes of the associated trend lines are 0.54 and 0.31, respectively, suggesting that changes in IP-10 concentration are 2- to 3-fold greater than the corresponding change in neopterin concentration.

At baseline, the median OAS-1 plasma concentration for all patients was 580 pM (see Fig. S3A in the supplemental material). Significant interpatient variability was noted, with a range 214 to 2,777 pM (13-fold). TN and NR patients showed median OAS-1 values of 580 and 561 pM, respectively. Median baseline OAS-1 concentrations of 572, 513, and 1,075 pM were observed in patients that experienced virologic continuous decline, plateau, and rebound, respectively (see Fig. S3B in the supplemental material). The range of OAS-1 values in TN and NR patients and in the continuous-decline, plateau, and rebound groups overlapped significantly, and differences between the groups were not statistically significant (see Fig. S3A and B in the supplemental material).

Baseline values of plasma IP-10 varied over a 35-fold range, which is consistent with previous reports (5, 15, 22). A previous study suggested a correlation between baseline concentration of IP-10 and baseline HCV RNA using a categorical analysis in a significantly larger patient population (42). No such correlation was evidenced in this study, although this apparent dis-
crepancy may reflect the smaller number of patients available for analysis in the current study.

In the case of interferon-based therapy, a low baseline IP-10 plasma concentration has been shown to be predictive of a strong first-phase decline in HCV RNA (3) and an improved rate of sustained virologic response (5, 8, 27, 42). Interestingly, in this study the magnitude of first-phase decline of HCV RNA during danoprevir monotherapy was directly correlated with baseline IP-10 concentration. These contrasting results may reflect the different antiviral mechanisms of interferon and a direct-acting antiviral agent or simply the small sample size in the current study.

Upon initiation of danoprevir therapy, the IP-10 plasma concentration was significantly reduced in patients that experienced a reduction in HCV RNA. These results are consistent with those of a recently published study demonstrating that patients treated for 13 days with a combination of danoprevir and the nucleoside polymerase inhibitor mericitabine showed significant reductions in IP-10 concentration (15).

In the present monotherapy study, the dynamics of IP-10 concentration during the first-phase decline of HCV RNA are shown in Fig. 5. The change in plasma neopterin concentration and median change in HCV RNA load for patients experiencing a virologic continuous decline (A), plateau (B), or rebound (C). The change in neopterin concentration at days 7 and 14 is shown for individual patients. Solid lines indicate the median change from baseline. The median change in HCV RNA load for the group is shown as closed gray circles connected by a solid gray line.

**FIG. 5.** Change in plasma neopterin concentration and median change in HCV RNA load for patients experiencing a virologic continuous decline (A), plateau (B), or rebound (C). The change in neopterin concentration at days 7 and 14 is shown for individual patients. Solid lines indicate the median change from baseline. The median change in HCV RNA load for the group is shown as closed gray circles connected by a solid gray line.

**FIG. 6.** Correlation of changes in plasma concentrations of neopterin and IP-10. Changes in neopterin and IP-10 are shown for individual patients treated with danoprevir or placebo at day 7 (A) or day 14 (B). The solid line in each panel shows a linear fit of the data, and dashed lines show the 95% confidence limits. Changes in neopterin and IP-10 concentrations are correlated, with an \( r^2 \) value of 0.35 and a \( P \) value of <0.0001 for day 7 and an \( r^2 \) value of 0.21 and a \( P \) value of 0.0017 for day 14.
plasma concentration were further associated with changes in HCV RNA concentration in patients experiencing rebound, plateau, and continuous decline. At day 7, similar median reductions were observed for both IP-10 and HCV RNA concentrations in the rebound and continuous-decline groups. Despite a considerable divergence of changes in HCV RNA concentration between these groups at day 14, median IP-10 concentrations were similar. Notably, however, three rebound patients with an early inflection in HCV RNA concentration and/or a high HCV RNA concentration at day 14 relative to that at baseline displayed a significant rebound in IP-10 concentration at day 14. The lack of observed rebound in IP-10 concentration in the other patients experiencing a rebound in HCV RNA may suggest that increases in inflammatory responses upon viral rebound are delayed relative to increases in HCV RNA concentrations. Larger studies with extended post-treatment follow-up would be required to test this hypothesis.

In the current study, the median baseline neopterin plasma concentration of 7.6 nM is similar to concentrations previously reported for patients with chronic HCV infection (17, 19). After initiation of danoprevir treatment, the median plasma concentration of neopterin appeared to be moderately decreased, but not in a statistically significant fashion. Patients experiencing 2- to 3- and 3- to 4-log_{10} IU/ml reductions in HCV RNA concentrations experienced 15% and 12% median reductions in neopterin concentrations, respectively, at day 14. A previous study examined changes in neopterin concentration associated with 15-day treatment of chronic HCV patients with telaprevir (17). In that study, two identically treated patient cohorts experienced mean HCV RNA reductions of 3.8 and 4.2 log_{10} IU/ml. These cohorts experienced mean reductions in plasma neopterin of 14% and 30%, respectively, with the latter being statistically significant relative to that at baseline. A third cohort receiving a lower dose of telaprevir experienced a 2.5-log_{10} IU/ml reduction in mean HCV RNA concentration and a 13% reduction in neopterin concentration that was not statistically significant. Taken together, the results of these two studies suggest that treatment with an NS3/4A protease inhibitor promotes reduction of the plasma concentration of neopterin. However, both this study and the previous study (17) suggest that the magnitude of this effect is modest.

Changes in the plasma concentrations of IP-10 and neopterin were statistically significant but weakly correlated. The correlation between changes in IP-10 concentration and neopterin concentration is consistent with each being an inflammatory marker and their persistent elevation in patients with chronic HCV infection (5, 8, 10, 17, 19, 27, 33, 36, 52). The correlation of changes in neopterin and IP-10 suggests that these markers may have similar or overlapping roles in the assessment of HCV patients during treatment with direct-acting antiviral agents.

The median baseline OAS-1 plasma concentration of 580 pM observed in the present study is similar to that in a previous literature report (48). While the significant variability observed in placebo-treated patients over a 2-week period makes it difficult to gauge any danoprevir-related effects on OAS-1 concentration, the variability and lack of discernible correlation with antiviral response may have significant implications for the potential use of OAS-1 as a marker of response to treatment with interferon-free direct-acting antiviral regimens.

The lack of association between OAS-1 and the antiviral effect resulting from danoprevir treatment is interesting, given the hypothesis that NS3/4A can dampen viral sensing and interferon production (14, 45). With interferon-based regimens or other activators of immune responses, a significant induction of OAS-1 is observed (48). Therefore, these results suggest that direct inhibition of HCV replication in the liver does not result in a systemically evidenced interferon response.

In summary, the current study demonstrates a significant reduction in IP-10 plasma concentration in patients with an antiviral response to the NS3/4A protease inhibitor danoprevir. Reductions in IP-10 concentration during therapy correlate with reductions in neopterin concentration that are of a smaller magnitude and more variable. These results suggest that treatment with a direct-acting antiviral agent suppresses hepatic inflammation in patients with chronic HCV infection. Similar correlations were not evidenced with plasma concentrations of the interferon-stimulated gene product OAS-1.

**ACKNOWLEDGMENT**

This work was supported by InterMune, Inc.

**REFERENCES**