Invasive aspergillosis is a life-threatening disease among patients with leukemia and bone marrow and solid-organ transplantation (1). Aspergillus fumigatus is the most common pathogen causing invasive aspergillosis, accounting for >70% of the cases, followed by Aspergillus flavus, Aspergillus terreus, and Aspergillus niger (2). Conventional amphotericin B was for many years the drug of choice and is still used against invasive aspergillosis. Despite its in vitro potent antifungal activity against Aspergillus spp., clinical trials have shown that conventional amphotericin B therapy is associated with <60% survival in patients with invasive aspergillosis (3–6). Although underlying disease, immunosuppression, toxicity, and timing of antifungal therapy affect the mortality of these infections, pathogen susceptibility and drug serum concentrations also represent important contributing factors. In vivo studies have shown that the best efficacy was achieved when the maximum concentration of amphotericin B in serum exceeded 2.4 times the MIC of the A. fumigatus isolate (7). However, in vitro antifungal susceptibility testing of amphotericin B is challenged by the lack of reliable susceptibility breakpoints for each Aspergillus species.

The epidemiological cutoff values 2, 2, and 4 mg/liter for the CLSI method and 1, 4, and 4 mg/liter for the EUCAST method were determined for A. fumigatus, A. flavus, and A. terreus, respectively, in order to detect isolates with extreme MICs (8, 9). The current susceptibility breakpoints for Aspergillus spp. with the CLSI and EUCAST methodologies are ≤ 1 mg/liter (9, 10). However, the supporting clinical and experimental in vivo data are poor, and no pharmacokinetic-pharmacodynamic (PK-PD) studies have validated this breakpoint (9). In a retrospective case-control study of 29 patients, 22/23 patients with isolate MICs of ≥ 2 mg/liter died, whereas 6/6 patients with isolate MICs of ≤ 1 mg/liter survived (11). Of note, 5/6 survivors with isolate MICs of ≤ 1 mg/liter underwent surgery or resolved their neutropenia. In two other large retrospective studies of 160 and 40 patients, no correlation was found between an Aspergillus MIC of ≥ 2 mg/liter and clinical outcome in patients with invasive aspergillosis treated with conventional amphotericin B (12, 13). Notably, more than 95% of A. fumigatus isolates have MICs of ≤ 1 mg/liter (8). Therefore, the majority of the isolates would be considered wild-type susceptible based on the 1-mg/liter cutoff, while conventional amphotericin B therapy has been associated with <60% of survival in all clinical trials (3, 4, 14). Furthermore, treatment failure was observed in patients infected with A. fumigatus and A. flavus isolates with CLSI MICs of 0.25 to 0.5 mg/liter and 1 mg/liter, respectively, when treated with the standard dose of 1 mg/kg of conventional amphotericin B (15, 16). Thus, a clinically relevant and...
useful susceptibility breakpoint for amphotericin B and Aspergillus spp, is warranted.

An in vitro pharmacokinetic-pharmacodynamic model was recently developed for Aspergillus spp, simulating pharmacokinetics of antifungal drugs (17). This model revealed considerable pharmacodynamic differences among Aspergillus isolates reflecting differences in time- and concentration-dependent inhibitory and fungicidal activity that were not captured by the MIC (18). Using the same model and simulating multidose human pharmacokinetics of voriconazole, in vitro data were predictive of experimental animal data and clinical outcome for A. fumigatus, enabling the determination of susceptibility breakpoints and target values for therapeutic drug monitoring (19).

Therefore, in the present study, we simulated multidose pharmacokinetics of amphotericin B and studied its activity against A. fumigatus, A. flavus, and A. terreus. The in vitro PK-PD relationship was described and bridged with human PKs in order to determine susceptibility breakpoints for each species.

**MATERIALS AND METHODS**

**Isolates.** Three clinical strains of A. fumigatus (AFM 4215; ATCC MYA-1163), A. flavus (AFL 113), and A. terreus (AT 137) isolated from patients with invasive aspergillosis were studied. The MICs of amphotericin B as determined thrice according to the CLSI broth microdilution method were 1 mg/liter for all Aspergillus species. The strains were maintained at −70°C in 10% glycerol and cultured twice in Sabouraud dextrose agar at 30°C for 5 to 7 days. A conidial suspension was prepared in normal saline with 1% Tween 20. Conidia were counted with a Neubauer chamber in order to obtain a final suspension of 1 × 10⁹ CFU/ml, and their concentration was confirmed by quantitative cultures on Sabouraud dextrose agar.

**Antifungal drug and medium.** Pure powder of amphotericin B (Sigma-Aldrich, Bioline, Athens, Greece) was reconstituted at 5,000 mg/liter in dimethyl-sulfoxide and was stored at −70°C. The medium contained 10.4 g/liter RPMI 1640 with glutamine without sodium bicarbonate (Sigma-Aldrich, St. Louis, MO) and 0.165 M buffer morpholinopropanesulfonic acid (MOPS) (Invitrogen, Carlsbad, CA), pH 7.0, with 100 mg/liter chloramphenicol (Sigma-Aldrich, St. Louis, MO).

**In vitro pharmacokinetic-pharmacodynamic model.** The in vitro pharmacokinetic simulation model consisted of (i) a glass beaker containing 700 ml medium (external compartment [EC]) in which was placed (ii) a dialysis tube of a 10-ml volume (internal compartment [IC]) made of a permeable cellulose membrane, which allowed free diffusion of molecules with a molecular mass of <20 kDa, and (iii) a peristaltic pump (Minipuls Evolution, Gilson, France), which removed the content of EC and added fresh medium within it at a rate equivalent to drug removal from human serum (17). The conidial suspension was inoculated in the IC, where it remained trapped together with the galactomannan (molecular mass of 20 kDa to 60 kDa) produced by the growing mycelia, while nutrients and drug diffused freely between the IC and EC. The concentration of the galactomannan increased with fungal growth, as found previously (20).

After inoculation of the IC, the drug was injected into the EC and IC (for rapid equilibration between the two compartments) every 24 h, and its concentration declined over time, with a half-life (t₁/₂) observed in human serum after intravenous administration of amphotericin B. Because amphotericin B degrades in vitro at 37°C, concentrations were adjusted to account for the higher clearance. The EC was covered with aluminum foil in order to minimize light exposure and placed on a heated magnetic stirrer (37°C). At the beginning of, during, and at the end of each experiment, temperature and flow rate were measured to ensure that they were at the expected values. All experiments were repeated twice.

**Bioassay.** The drug levels in the IC were determined by a microbiological agar diffusion method using the strain Paecilomyces variotii ATCC 22319 (18, 21). Specifically, P. variotii conidia at final concentration of 5 × 10⁴ CFU/ml were inoculated into prewarmed (54°C) RPMI 1640 medium and MOPS with 15 g/liter agar and poured onto 10- by 10-cm plastic plates. After solidification of the agar, 1-cm-diameter holes were opened and filled with 100 μl of known drug dilutions (range of 0.1 to 8 mg/liter), as well as 100 μl of IC samples. The plates were incubated at 37°C for 24 h, when diameters of inhibition zones were measured. Unknown drug concentrations in the IC samples were determined using the standard curve constructed from known drug concentrations and corresponding diameters of inhibition zones. The diameters of inhibition zones correlated linearly with amphotericin B concentrations (R² > 0.9). The lowest limit of detection was 0.1 mg/liter, and the inter- and intraday variations were <10% within the detection range.

**Pharmacokinetic analysis.** The time-concentration profile of free amphotericin B observed in human serum after intravenous administration of 0.6 and 1 mg/kg/day (22, 23) was simulated in the in vitro model. Thus, the free peak concentrations (fCmax) of 0.1, 0.3, 0.6, 1.2, and 2.4 mg/liter were introduced in the system once daily for 3 days (for A. flavus and A. terreus, the fCmax of 0.8 and 1 mg/liter were also tested in order to differentiate their pharmacodynamics). Although fCmax of >1 mg/liter are not clinically achievable in human serum because of the drug’s solubility limitation, these concentrations were simulated in order to fully describe the in vitro exposure-effect relationship. The biphasic 24-h time-concentration profiles of amphotericin B with an alpha phase with a short half-life of <1 h observed within 2 h after drug administration followed by a beta phase with a longer half-life of 6 to 10 h observed 2 to 24 h after drug administration were simulated. Amphotericin B concentrations were determined at 0 h, 4 h, 6 h, 8 h, 20 h, 24 h, 44 h, 48 h, and 72 h after the introduction of the drug in the IC using the bioassay. The data were analyzed with nonlinear regression based on a two-compartment model described by the equation C = C₀e⁻kt₀ + C₀e⁻kt₀, where k₀ and kₜ₀ are the rate constants, C₀ and Cₜ₀ are the y intercepts for alpha and beta, respectively, and C is the concentration at a given time t. The half-lives of alpha and beta phases were calculated using the equations t₁/₂α = ln(2)/2, respectively.

**Determination of fungal growth.** Fungal growth in the IC was assessed in samples of 100 μl at regular time intervals by determining galactomannan production using an enzyme-linked immunosorbent assay (ELISA) (Platelia; Bio-Rad, Athens, Greece). Samples were diluted with 200 μl saline in order to reach the final volume of 300 μl before processing. Results were expressed as a galactomannan index (GI) according to the manufacturer’s instructions. Galactomannan levels were also determined in the EC in order to ensure that no galactomannan escaped from the IC.

**Pharmacodynamic analysis.** In vitro pharmacodynamics of each amphotericin B dose and Aspergillus species were determined based on the GI-time relationship analyzed with the ECmax model: E = E₀min + E₀max × T/(T₀ + Tₜ₀), where E is the GI (dependent variable), E₀max and E₀min are the maximum and minimum GI, respectively, T is the incubation time (independent variable), T₀ and Tₜ₀ are the time corresponding to 50% of the E₀max, and γ is the slope of the curve. As shown previously, the parameters E₀max, γ, and Tₜ₀ describe the extent, rate, and time of galactomannan production, respectively, whereas the area under the galactomannan index-time curve (AUCGI) is a surrogate marker of fungal growth capturing any differences in extent, rate, and time of antifungal activity. The higher the AUCGI, the greater is the fungal growth over time. The area under the galactomannan index-time curve (AUCGI) was calculated for each amphotericin B dose and drug-free control. The percentage of fungal growth at each dose was calculated based on the AUCGI of each dose divided by the AUCGI of the drug-free growth control. The percent growth-tCmax/MIC relationship was analyzed with the ECmax model: E = E₀min + E₀max × P/(P₀ + Pₜ₀), where E is the percent growth (dependent variable), E₀max and E₀min are the maximum and minimum percent growth, respectively, P is the PK-PD index tCmax/MIC (independent variable), P₀ and Pₜ₀ are the tCmax/MIC corresponding to 50% of E₀max, and γ is the slope of the curve. In addition, the tCmax/MIC corresponding to near-maximum an-
Monte Carlo analysis was also performed for a lower dose of conventional human PKs, the steady-state patients and for each Aspergillus species. Coefficient of variations was ±15% between two replicates.

Monte Carlo simulation. In order to bridge the in vitro data with human PKs, the steady-state \( f_{C_{\text{max}}} \) of 1,000 patients receiving the standard dose of amphotericin B, 1 mg/kg intravenously, were simulated with Monte Carlo analysis. This dosage resulted in steady-state total maximum concentrations in human serum (\( t_{C_{\text{max}}} \)) of 2.83 ± 1.17 mg/liter, respectively (23), which corresponds to free maximum concentrations (\( f_{C_{\text{max}}} \)) of 0.14 ± 0.06 mg/liter based on the 95% protein binding rate of amphotericin B previously found at these concentrations (24). The percentage of patients with \( f_{C_{\text{max}}}/\text{MIC} \) exceeding the median, upper, and lower 95% CI limits of the PK-PD targets \( P_{f\text{so}} \) and \( P_{f\text{so}} \) were calculated for different MICs and for each Aspergillus species.

Because the dose of 1 mg/kg is associated with significant toxicity, Monte Carlo analysis was also performed for a lower dose of conventional amphotericin B, 0.6 mg/kg, used in clinical practice. This dose resulted in a serum \( t_{C_{\text{max}}} \) of 1.43 ± 0.2 mg/liter, corresponding to an \( f_{C_{\text{max}}} \) of 0.0715 ± 0.01 mg/liter (22, 24). The median, upper, and lower 95% CI limits of percentage of \( P_{f\text{so}} \) target attainment were calculated for each species and for different MICs. Finally, because trough levels (\( C_{\text{min}} \)) are clinically easier obtained than \( C_{\text{max}} \) levels, the trough levels were assessed using linear regression analysis with \( C_{\text{max}} \) concentrations in the in vitro PK-PD model.

Statistical analysis. All analysis was performed with the Prism 5.01 software (GraphPad Inc., La Jolla, CA). All \( E_{\text{max}} \) models were globally fitted to the data, with \( E_{\text{max}} \) and \( E_{\text{so}} \) shared among data sets. Comparisons between \( E_{\text{max}} \) model parameters among Aspergillus species were assessed using the extra sum-of-squares F test. Monte Carlo analysis was performed with the random function of an Excel spreadsheet (MS Office 97). A P value of <0.05 was considered statistically significant.

**RESULTS**

Pharmacokinetic analysis. The time-concentration profiles of amphotericin B doses are shown in Fig. 1. The calculated \( f_{C_{\text{max}}} \) in the IC were within ±20% of the target \( f_{C_{\text{max}}} \) of 0.1, 0.3, 0.6, 0.8, 1, 1.2, and 2.4 mg/liter. The half-lives of alpha and beta phases were 0.2 to 2 h and 10 to 17 h, respectively.

Pharmacodynamic analysis. Fungal growth was progressively inhibited at higher concentrations of amphotericin B. The GI-time curves followed a sigmoid pattern described well by the \( E_{\text{max}} \) model (\( R^2 > 0.82 \)), and they were characterized by the same \( E_{\text{max}} \) but different slopes and \( T_{50} \)s for the different doses of amphotericin B and Aspergillus species tested (Fig. 2). Complete inhibition of fungal growth was found for \( A. \) fumigatus with an amphotericin B \( f_{C_{\text{max}}} \) of ≥0.3 mg/liter, while for \( A. \) flavus and \( A. \) terreus, complete inhibition was obtained with an amphotericin B \( f_{C_{\text{max}}} \) of ≥0.8 mg/liter. In particular, the percentages of fungal growth at \( f_{C_{\text{max}}} \) of 0.1, 0.3, and 0.6 mg/liter were 97%, 78%, and 74% for \( A. \) flavus and 96%, 82%, and 67% for \( A. \) terreus, respectively.

The \( f_{C_{\text{max}}}/\text{MIC} \) relationship followed a sigmoid pattern (global \( R^2 = 0.96 \)) for all three species (Fig. 3). The \( f_{C_{\text{max}}}/\text{MIC} \) (95% CI)

**FIG 1** Multidose pharmacokinetics of amphotericin B in the in vitro model with different \( f_{C_{\text{max}}} \) and \( t_{C_{\text{max}}} \) of 0.2 to 2 h and 10 to 17 h for alpha and beta phases, respectively. Coefficient of variations was ±15% between two replicates.

**FIG 2** Multidose pharmacodynamic analysis of simulated amphotericin B doses with \( f_{C_{\text{max}}} \) of 0.1, 0.3, 0.6, 0.8, 1, 1.2, and 2.4 mg/liter against \( A. \) fumigatus, \( A. \) flavus, and \( A. \) terreus isolates with a modal CLSI MIC of 1 mg/liter as determined by the galactomannan index in the in vitro PK-PD model.
associated with 50% growth for *A. fumigatus* was 0.145 (0.133 to 0.158), which was statistically significantly lower than the corresponding \( f_{C_{\text{max}}}/\text{MIC} \) of *A. flavus* (0.371; 0.283 to 0.486) and *A. terreus* (0.41; 0.292 to 0.522) \((P < 0.001)\). The \( f_{C_{\text{max}}}/\text{MICs} \) corresponding to near maximal activity (20% growth) were 0.188 (0.166 to 0.213), 0.474 (0.362 to 0.655), and 0.545 (0.376 to 0.714) for *A. fumigatus*, *A. flavus*, and *A. terreus*, respectively.

**Monte Carlo simulation analysis.** Monte Carlo simulation analysis of 1,000 patients receiving the standard amphotericin B dose of 1 mg/kg resulted in mean \( IC_{\text{max}} \) ± standard deviation (SD) values of 2.8 ± 1.2 mg/liter (\( f_{C_{\text{max}}} \) ± SD of 0.14 ± 0.06 mg/liter). These values are close to the target values previously found in clinical studies (23). The percentage of patients attaining the PK-PD target (\( f_{C_{\text{max}}}/\text{MIC} \)) associated with half-maximal (\( P_{50} \)) activity is shown in Fig. 4. Higher-than-75% and lower-than-15% \( P_{50} \) target attainment was found for *A. fumigatus* isolates with MICs of ≥0.5 and ≤2 mg/liter and for *A. flavus* and *A. terreus* isolates with MICs of ≤0.25 and ≥1 mg/liter. In particular, the median (range) percentages of target attainment for *A. fumigatus*, *A. flavus*, and *A. terreus* isolates with a MIC of 1 mg/liter were 47% (39 to 57%), 0% (0 to 0%), and 0% (0 to 0%) and with a MIC of 0.5 mg/liter were 88% (87 to 90%), 24% (5 to 50%), and 15% (2 to 46%), respectively. The percentages of \( P_{50} \) target attainment for *A. fumigatus*, *A. flavus*, and *A. terreus* isolates with a MIC of 1 mg/liter were 22%, 0%, and 0%, respectively (data not shown).

For the lower amphotericin B dose of 0.6 mg/kg, a >98% \( P_{50} \) target attainment was found for MICs of ≤0.25, ≤0.125, and ≤0.125 mg/liter for *A. fumigatus*, *A. flavus*, and *A. terreus*, respectively (Fig. 4). A linear correlation \((r^2 = 0.98)\) was found between the \( C_{\text{max}} \) and the \( C_{\text{max}} \) levels in the *in vitro* model with a slope of 0.20 ± 0.03, indicating that \( C_{\text{max}} \) levels correspond to 1/5 of the \( C_{\text{max}} \) concentrations, as previously found in humans (22, 23).

**DISCUSSION**

The multidose pharmacodynamic study of amphotericin B using an *in vitro* pharmacokinetic-pharmacodynamic model that simulated free serum amphotericin B concentrations revealed differences among the three *Aspergillus* species. Complete growth inhibition was observed at an \( f_{C_{\text{max}}}/\text{MIC} \) of 0.03 mg/liter for *A. fumigatus* and ≥0.8 mg/liter for *A. flavus* and *A. terreus* despite their identical MIC values. The \( f_{C_{\text{max}}}/\text{MICs} \) (95% CI) corresponding to near-maximal activity were 0.188 (0.166 to 0.213), 0.474 (0.362 to 0.655), and 0.545 (0.376 to 0.714) for *A. fumigatus*, *A. flavus*, and *A. terreus*, respectively. These differences are in agreement with our previous study where single-dose pharmacokinetics and a higher initial inoculum were used, providing better discrimination of the *in vitro* activity of amphotericin B against the three species (18). The observed differences among the three species were attributed to the fast inhibitory action and increased killing rate against *A. fumigatus*, a slower inhibitory action and reduced killing efficiency against *A. flavus*, and the slowest inhibitory action and no killing against *A. terreus* (18).

The PK-PD target of near maximal antifungal activity found in the present study against *A. fumigatus* is in agreement with that previously obtained in a murine neutropenic model of experimental pulmonary aspergillosis demonstrating that the \( IC_{\text{max}}/\text{MIC} \) of 2.4 was associated with near-maximal survival (7). Considering that the free-drug percentage of conventional amphotericin B in mouse serum corresponded to 7.4% (25), the *in vivo* \( f_{C_{\text{max}}}/\text{MIC} \) corresponds to 0.178, which is very close to the *in vitro* \( f_{C_{\text{max}}}/\text{MIC} \) of 0.188 found in the present study. In addition, in neutropenic murine models of experimental aspergillosis, the dose of 5 mg/kg of amphotericin B was associated with >80% survival against *A. fumigatus* isolates with MICs of 0.5 and 1 mg/liter and <40% against *A. terreus* and *A. flavus* isolates with MICs...
of 1 to 2 mg/liter (26 to 29). The dose of 5 mg/kg, which corresponds to a $C_{\text{max}}$ around 2.5 mg/liter (i.e., $C_{\text{max}}$ of 0.185 mg/liter) in mice serum (7, 30), would attain the PK-PD targets determined in the present study for A. fumigatus but not for A. terreus and A. flavus isolates. However, there are many confounding factors, like immunosuppression, type and severity of infection, dosing regimens, time of initiation of antifungal therapy, pharmaco-kinetic variation, and in vitro susceptibility testing, that prohibited direct in vitro-in vivo correlation (31 to 33). These factors can be simulated in the present model in order to study the impact of host defense cells, delayed drug administration, and disease progression on the magnitude of the PK-PD parameter.

The present in vitro model indicated that the standard dose of 1 mg/kg of amphotericin B is associated with 47% target attainment using a half-maximum activity PK-PD target ($P_{50}$) and 22% using a near-maximum activity PK-PD target ($P_{80}$) for Aspergillus fumigatus isolates with an amphotericin B MIC of 1 mg/liter. This is in agreement with a previous retrospective study where the MICs of 274 clinical Aspergillus isolates from transplant recipients with proven or probable invasive aspergillosis were analyzed together with survival after 6 and 12 weeks of treatment (34). In that study, isolates with amphotericin B MICs of 1 mg/liter belonging mainly to Aspergillus fumigatus species were associated with 45% 6-week and 25% 12-week survival, in agreement with the percentages of $P_{50}$ and $P_{80}$ target attainment found in the present study, respectively.

The choice of the in vitro PK-PD target ($P_{90}$ or $P_{80}$) for correlating in vitro data with clinical outcome remains controversial. The percentage of $P_{90}$ target attainment was comparable to 12-week survival, whereas the percentage of $P_{80}$ target attainment was comparable to 6-week survival. High serum drug exposure ($P_{80}$) may result in high concentrations at the site of infection and/or fungicidal actions that will likely produce a long-lasting effect (12-week survival) compared to the lower drug exposure ($P_{90}$) that will produce a short-lasting effect (6-week survival). However, since most deaths during the second 6-week period of 12-week therapy were attributed to causes other than invasive aspergillosis, albeit with some uncertainty, the 6-week survival endpoint was suggested to be the preferable endpoint to judge the effectiveness of antifungal therapy (35). Because the percentage of $P_{90}$ target attainment found by Monte Carlo analysis of the present study for A. fumigatus isolates with an amphotericin B MIC of 1 mg/liter was correlated with the 6-week survival found previously in a retrospective study of the MIC-clinical outcome correlation for the same isolates (34), we used the $P_{90}$ endpoint to bridge the in vitro PK-PD data with human PK in Monte Carlo analysis and to determine susceptibility breakpoints.

An important assumption when bridging in vitro concentrations to serum PK is that the serum concentrations correspond to the tissue concentrations at the site of infection, which is the lung in the case of invasive pulmonary aspergillosis. In vivo pharmacokinetic data of conventional amphotericin B in mice showed that although the $C_{\text{max}}$ in the lung was lower than the $C_{\text{max}}$ in serum (3.33 and 4.58 mg/liter, respectively), the $C_{\text{max}}$ values were similar (0.33 and 0.34 mg/liter, respectively) based on the different degree of protein binding in the lung (10%) compared to that in serum (7.4%) (30). In two clinical studies, a wide range of amphotericin B lung concentrations (0.1 to 23.3 μg/g) were detected with high-performance liquid chromatography (HPLC) in patients with mean concentrations of 11.29 and 5.29 μg/g (36, 37). However, only a small (1.5% to 30%) proportion of HPLC-detected amphotericin B was bioactive, and an even smaller proportion of the latter (<10%) corresponded to free amphotericin B (36, 37). Thus, the free concentration of amphotericin B in the lung may be similar to the free drug concentrations in human serum. For other sites of infections where amphotericin B penetrates poorly (e.g., brain), the tissue-to-serum ratio should be taken into account. Microdialysis experiments may provide better estimates of free amphotericin B in tissues (38).

The results obtained by the Monte Carlo analysis point to the following breakpoints for susceptibility, intermediate susceptibility, and resistance: ≤0.5, 1, and ≥2 for A. fumigatus and ≤0.25, 0.5, and ≥1 for A. flavus and A. terreus. Based on these breakpoints and previous epidemiological susceptibility data, 40% of A. fumigatus, 90% of A. flavus, and 98% of A. terreus isolates would be considered nonsusceptible (8). In a randomized prospective clinical trial of invasive aspergillosis mainly by A. fumigatus, conventional amphotericin B was associated with 42% 6-week mortality, which is in agreement with the 40% of A. fumigatus isolates estimated in the present study being nonsusceptible (3). Furthermore, in a review paper of 60 clinical cases of invasive A. terreus infections published from 1966 to 2003, 28 cases were treated with conventional amphotericin B, and the mortality rate was 96% (27/28), which is also in agreement with the 98% of A. terreus isolates estimated in the present study being nonsusceptible (39). Finally, in a cohort study of 11 cases of invasive A. flavus infections treated only with conventional amphotericin B, the mortality rate was 81% (9/11), in agreement with the 90% of A. flavus isolates estimated in the present study being nonsusceptible (16). Interestingly, in the latter study, all patients with A. flavus isolate MICs of ≥1 mg/liter died. These clinical data provide further support of the breakpoints determined in the present PK-PD model.

In order to maximize the efficacy and minimize the toxicity of conventional amphotericin B treatment, one could increase or decrease the dose based on the MIC of isolated pathogens or epidemiological data of each center. Thus, for a small number of A. fumigatus, A. flavus, and A. terreus strains with MICs of ≤0.25, ≤0.125, and ≤0.125 mg/liter, respectively, the lower dose of 0.6 mg/kg would retain efficacy of the 1-mg/kg dosage but reduce its toxicity. The 0.6-mg/kg dose was associated with 30% renal toxicity compared to >50% renal toxicity associated with the 1-mg/kg dose of amphotericin B (5, 40). For isolates with intermediate susceptibility, therapeutic drug monitoring (TDM) with the goal to attain the PK-PD target could increase the efficacy of the standard dose of 1 mg/kg of amphotericin B as previously suggested (41). Since TDM is easier performed using trough ($C_{\text{min}}$) levels than $C_{\text{max}}$ levels, the $C_{\text{min}}$/MIC ($C_{\text{min}}$/MIC) ratios based on the in vitro model were 0.029 (0.58), 0.074 (1.48), and 0.082 (1.64) for A. fumigatus, A. flavus, and A. terreus, respectively. Since in vivo amphotericin B is bound by tissue and slowly released, accounting for the prolonged (serum) terminal elimination half-life of the drug, the correlation between $C_{\text{min}}$ and $C_{\text{max}}$ may be clinically relevant only for the first few days of therapy. However, the unavoidable toxicity of conventional amphotericin B, which often results in drug discontinuation, will be the limiting factor even for susceptible isolates, limiting the use of conventional amphotericin B, in agreement with the ECIL guidelines for the management of invasive aspergillosis (42). Therefore, lipid formulations of amphotericin B could be used efficiently even for isolates with reduced susceptibility to conventional amphotericin
B, as previously shown in an animal model infected by an A. *fumigatus* isolate with a MIC of 1 mg/liter (43). Lipid formulations can safely deliver high concentrations of amphotericin B in the fungal target and at the site of infection (44).

In conclusion, a PK-PD model was used to simulate free amphotericin B serum concentrations and to study the *in vitro* activity against three *Aspergillus* species. The PK-PD targets were 0.145, 0.371, and 0.41 \( f_{C_{\text{max}}}/\text{MIC} \) for *A. fumigatus*, *A. flavus*, and *A. terreus*, respectively, and the susceptibility breakpoints derived were \( \leq 0.5, 1, \) and \( \geq 2 \) mg/liter for *A. fumigatus* and \( \leq 0.25, 0.5, \) and \( \geq 1 \) mg/liter for *A. flavus* and *A. terreus*. The suggested breakpoints are substantiated with results obtained from previously published clinical studies of invasive aspergillosis.

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