Lysophosphatidylcholine (LPC) is a major component of phospholipids that is involved in the recruitment and stimulation of immune cells (1–3) and therefore may eliminate dead eukaryotic and prokaryotic cells during infection (4, 5). Recent attention has been drawn to the role of LPC as a chemotactic factor during bacterial infection (1, 5). In eukaryotic cells, LPC was released after activation of calcium-independent phospholipase A2 (iPLA2) and cytosolic phospholipase A2 (cPLA2) (4, 6, 7). The latter plays an important role in the cell damage induced by Pseudomonas aeruginosa, Escherichia coli, and Staphylococcus aureus (8–10).

Acinetobacter baumannii is a Gram-negative bacillus that is very important in the clinic due to an increase in the number of nosocomial infections and because of its resistance to most clinically available antimicrobials, which makes it difficult to manage A. baumannii infections. Infections caused by A. baumannii include the following, in order of prevalence and associated mortality: pneumonia, bacteremia, urinary tract infections, surgical wound infections, and meningitis (11). A. baumannii pneumonia and bacteremia are typically acquired in the hospital setting and are associated with significant mortality (12).

A. baumannii infection requires two steps. First, A. baumannii must adhere to the host cells and then penetrate these cells to join the surrounding tissue (13, 14). It has been shown that A. baumannii uses a common strategy to traverse the host cell: this microorganism induces the death of host cells in a caspase-3- and calpain calcium-dependent manner (14–16). The induction of death in host cells results in the release of LPC, which is an important factor for the stimulation of immune cells (1–3). Therefore, we hypothesized that LPC regulates the inflammatory cascade caused by A. baumannii and that, in this sense, it may be useful as an adjuvant in combination with antimicrobial agents for the treatment of infections by this pathogen. We showed that LPC pretreatment in murine models of peritoneal sepsis and pneumonia caused by A. baumannii reduced the bacterial loads and bacteremia and increased the mouse survival rates.

### MATERIALS AND METHODS

**Bacterial strain and growth conditions.** A. baumannii ATCC 17978 was used in this study. The bacterial cells were grown in Luria-Bertani (LB) broth at 37°C with shaking. The growth of the bacterial cells was monitored for 4 or 24 h by determining the culture’s optical density at 600 nm (OD$_{4\text{h}}$ = 0.5 and OD$_{24\text{h}}$ = 1.2).

**In vitro susceptibility testing.** The MIC of LPC for ATCC 17978 was determined by a microdilution assay, using a standard inoculum of $1 \times 10^5$ to $5 \times 10^6$ CFU/ml, as previously described (16).

**Animals.** Immunocompetent C57BL/6 female mice (16 to 18 g) were obtained from the University of Seville facility; they had a sanitary status of murine pathogen free and were assessed for genetic authenticity. Animals were housed in regulation cages, with food and water provided ad libitum. This study was carried out in strict accordance with the recommendations in the Guide for the Care and Use of Laboratory Animals (17).

The protocol was approved by the Committee on the Ethics of Animal Experiments of the University Hospital of Virgen del Rocío, Seville, Spain (03/2010). All surgery was performed under sodium thiopental anesthesia, and all efforts were made to minimize suffering.
**A. baumannii peritoneal sepsis model.** A murine peritoneal sepsis model with *A. baumannii* ATCC 17978 was established by intraperitoneal (i.p.) inoculation of bacteria (18). Briefly, female C57BL/6 mice were inoculated with 0.5 ml of a bacterial suspension which had been incubated for 20 to 24 h in LB broth at 37°C and mixed at a 1:1 ratio with a saline solution containing 10% (wt/vol) porcine mucin (Sigma, Spain). The minimal lethal dose (MLD_{50}), 50% lethal dose (LD_{50}), and maximum tolerated dose (LD_{x}) were determined by inoculating various groups of mice (6 mice per group) with decreasing amounts of *A. baumannii* ATCC 17978 inoculum, from 8.5 to 2.3 log CFU/ml, and monitoring the survival of the mice for 7 days. The LD_{50} and the LD_{50} were defined as the concentrations causing 0% and 50% morality, respectively.

**Cytokines and LPC assays.** Blood samples were collected from the peribulbar plexuses of 40 anesthetized mice infected with ATCC 17978 at the MLD_{100} as previously described (19). The serum levels of tumor necrosis factor alpha (TNF-α), interleukin-6 (IL-6), IL-1β, IL-10, and LPC were determined in mice at 0, 0.5, 1, 2, 4, 8, 12, and 24 h postinfection by using enzyme-linked immunosorbent assays (ELISAs) (eBioscience, San Diego, CA) and an LPC assay kit (Azwell, Japan).

**In vivo LPC toxicity.** The Reed and Muench method (20) was used to determine toxicity. Mice were inoculated i.p. with a single dose of LPC (obtained from human brain; Sigma), starting at 10 mg/kg of body weight, in 0.5 ml 0.9% NaCl, and the solution was serially diluted until 50% mortality was reached; 6 mice were included in each group.

**LPC pharmacokinetics.** Serum LPC levels were determined in healthy mice after a single i.p. administration of 25 mg/kg LPC. After 5, 15, 30, 60, and 120 min, blood was extracted from the peribulbar plexuses of the anesthetized mice; three mice were used for each time point. The LPC levels were determined as described above. The maximum concentration in serum (C_{max}, reported in micromolar units), the area under the concentration-time curve from time zero to infinity (AUC_{0-\infty}, reported in micromole-hours per liter), and the terminal half-life (t_{1/2}; reported in hours) were calculated using a computer-assisted method (21).

**LPC protective dose.** The protective dose for 50% of the population (PD_{50}) was determined for LPC as described previously (20), with some modifications. Mice were infected i.p. with *A. baumannii* ATCC 17978 at the MLD_{100}. Two hours later, the animals were treated i.p. with single increasing doses of LPC, starting from 10 mg/kg in 0.5 ml 0.9% NaCl and continuing until the LD_{50}, and were observed for 2 days to measure the cumulative survival rates; 6 mice were used for each treatment group.

**Therapeutic LPC effects in experimental animal models.** (i) Peritoneal sepsis model. Mice were inoculated i.p. with *A. baumannii* ATCC 17978 at the MLD_{100}. One hour prior to inoculation, the animals were infected i.p. with a single dose of 25 mg/kg LPC diluted in 0.5 ml 0.9% NaCl. This dose corresponds to half the LD_{50} of LPC. Thirty mice were randomly assigned to two different therapeutic groups: a control group (without treatment) and a group receiving 25 mg/kg LPC. The mice were treated and monitored for mortality (%) for 48 h. After death or sacrifice of the mice at the end of the experimental period, aseptic thoracotomies were performed, and blood samples for qualitative blood culture were obtained by cardiac puncture (data are reported as numbers [%] of positive cultures). The lungs were aseptically removed and homogenized as described above for quantitative culture (data are reported in log_{10} CFU/ml of lung).

(ii) Pneumonia model. A previously described experimental murine pneumonia model (22) was used to evaluate the efficacy of LPC against *A. baumannii* ATCC 17978. Briefly, the mice were anesthetized by an i.p. injection of 5% (wt/vol) sodium thiopental (Braun Medical, Barcelona, Spain). They were suspended vertically, and the trachea of each was then canulated with a blunt-tipped metal needle. The feel of the needle tip against the tracheal cartilage confirmed the intratracheal location. A microtiter syringe (Hamilton Co., Reno, NV) was used for inoculation of 50 µl of a bacterial suspension (8 log CFU/ml) which had been grown for 24 h in LB broth at 37°C and mixed at a 1:1 ratio with a 0.9% NaCl solution containing 10% (wt/vol) porcine mucin. The mice remained in a vertical position for 3 min and then in a 30° position until awake. Preemptive LPC therapy was performed by using a single i.p. dose of 25 mg/kg LPC diluted in 0.5 ml 0.9% NaCl. For the analysis of LPC's therapeutic efficacy against infection, the animals were randomly assigned to two different therapeutic groups: a control group (without treatment) and a group receiving 25 mg/kg LPC. The mice were treated and monitored for mortality (%) for 48 h. After death or sacrifice of the mice at the end of the experimental period, aseptic thoracotomies were performed, and blood samples for qualitative blood culture were obtained by cardiac puncture (data are reported as numbers [%] of positive cultures). The lungs were aseptically removed and homogenized as described above for quantitative culture (data are reported in log_{10} CFU/ml of lung).

**RESULTS**

**MLD_{100}, LD_{50}, and LD_{0} of *A. baumannii***. To determine the MLD_{100}, LD_{50}, and LD_{0} of *A. baumannii* ATCC 17978, the murine peritoneal sepsis model was used. Mortality was dependent on the concentration of bacteria in the inoculum (data not shown). The MLD_{100}, LD_{50}, and LD_{0} of strain ATCC 17978 were 3.2 log_{10} CFU/ml, 2.75 log_{10} CFU/ml, and 2.3 log_{10} CFU/ml, respectively.

**Effects of *A. baumannii* on cytokine levels.** The effects of *A. baumannii* on cytokine levels were examined from 0 to 12 h after induction of the murine peritoneal sepsis model. At the early time points, *A. baumannii* infection induced large, transient changes in the cytokine levels. During the first 2 h after the mice were inoculated with bacteria, IL-10, TNF-α, and IL-6 levels significantly increased (P < 0.05) over control levels (Fig. 1A). IL-1β levels were not affected during the first 2 h after bacterial infection, but these levels were increased 8 h after bacterial inoculation (Fig. 1A).

**Effect of *A. baumannii* on the LPC level.** We next investigated whether these changes in cytokine levels were associated with changes in the level of LPC in serum. At early time points of the murine peritoneal sepsis model, the LPC levels in the infected mice decreased between 0 and 2 h (P = 0.02), coincidently with an increase of inflammatory cytokine levels (Fig. 1A), whereas the LPC levels were only slightly affected in the control mice (Fig. 1B). At 8, 12, and 24 h, a statistically significant decrease in LPC levels was observed in the infected mice in comparison with the control group (Fig. 1B).

**LPC toxicity, protective dose, and pharmacokinetic studies.** The LD_{50} and LD_{50} values of LPC were 50 mg/kg and 100 mg/kg, respectively. Adverse effects were observed in a dose-dependent manner at doses of 50 mg/kg or higher, including transitory movement disorders and muscle spasms. The PD_{50} against *A. baumannii* ATCC 17978 was 25 mg/kg. After a single dose of LPC (25 mg/kg), adverse effects were observed in a dose-dependent manner at 50 mg/kg or higher, including transitory movement disorders and muscle spasms. The PD_{50} against *A. baumannii* ATCC 17978 was 25 mg/kg.
mg/kg, i.p.), the pharmacokinetic parameters $C_{\text{max}}$, $t_{1/2}$, and AUC$_{0-\infty}$ were 73.17 μM, 0.48 h, and 29.25 μmol · h/liter, respectively. Serum LPC levels in healthy mice injected with LPC (25 mg/kg, i.p.) were increased 15% over the basal level 5 min after the dose (from 461.7 ± 17.24 μM to 534.87 ± 24.81 μM) (Fig. 2).

**LPC protects against *A. baumannii* peritoneal sepsis-induced infection and lethality.** We examined the effects of preemptive LPC therapy on bacterial tissue burdens and sterile blood cultures in the murine peritoneal sepsis model 2 days after bacterial inoculation of the mice. LPC reduced the lung and spleen bacterial concentrations by 2.37 and 3.9 log$_{10}$ CFU/g, respectively, compared to the controls ($P < 0.05$). The numbers of sterile blood cultures and survivors were increased in the LPC-pretreated group compared to the control group (Table 1). Interestingly, the MIC of LPC against *A. baumannii* ATCC 17978 is $>2$ mg/ml, which is $5 \times 10^4$-fold higher than the LPC $C_{\text{max}}$.

**LPC protects against *A. baumannii* pneumonia-induced infection and lethality.** We examined the effects of preemptive LPC therapy on bacterial burdens and sterile blood cultures in the murine pneumonia model for 2 days after bacterial inoculation. LPC reduced the lung bacterial concentration by 2.49 log$_{10}$ CFU/g compared to the control level ($P < 0.05$). The numbers of sterile blood cultures and survivors were increased in the LPC-treated group compared to the control group (Table 1).

**Effect of LPC on cytokine production by *A. baumannii*.** The effects of preemptive LPC therapy (25 mg/kg, i.p., 1 h prior to inoculation) on serum cytokine levels in the murine model of peritoneal sepsis caused by *A. baumannii* ATCC 17978 were examined from 0 to 12 h after inoculation (Fig. 3). At an early time point (4 h), LPC pretreatment induced decreases in the levels of the proinflammatory cytokines TNF-α and IL-6, whereas the level of the anti-inflammatory cytokine IL-10 was increased. The level of IL-1β, another proinflammatory cytokine, decreased in the group pretreated with LPC only at a late time point (12 h) in the experiment.

**DISCUSSION**

The present study provides new data highlighting the role of LPC, which is known to be a chemotactic factor, in the regulation of pro- and anti-inflammatory cytokine production in vivo. Furthermore, this study evaluated the in vivo therapeutic efficacy of LPC
against *A. baumannii* in the murine peritoneal sepsis and pneumonia models. Preemptive LPC therapy reduced the bacterial burdens in the lungs and spleen and increased the number of sterile blood cultures as well as the animal survival rate.

Interestingly, we found that serum LPC levels decreased during the first 2 h after infection of mice with *A. baumannii*, and this decrease was accompanied by increases in the levels of the proinflammatory cytokines TNF-α and IL-6. The decreased LPC concentration may reflect its enhanced and rapid conversion to lyso phosphatidic acid (LPA) by a plasmatic lysophospholipase D (23). LPA is known to induce a multitude of cellular responses, including LPA-driven effects on proinflammatory cytokine production (24). Clinically, it was reported that plasma LPC was significantly decreased in septic patients (25, 26), and patients with unfavorable sepsis outcomes had significantly lower plasma LPC-phosphorylcholine levels than patients who survived a septic episode (25, 26). These clinical findings support the results of our study.

Preemptive LPC therapy with 25 mg/kg LPC effectively protected the mice from *A. baumannii* infections and effectively reduced the bacterial burdens in the organs and the positive blood culture rate. At the dose used in this study, the Cmax of LPC in the sera of the mice was 2- to 3-fold higher than the 10 to 30 μM required to induce the chemotaxis of immune cells (24, 26), and this treatment did not induce any apparent toxic effects in the mice. In the peritoneal sepsis model, we observed that 60% of the LPC-treated mice were negative for bacteremia and that 40% of the LPC-treated mice survived. We suggest that the death of the nonbacteremic mice was due to the inflammatory responses caused by *A. baumannii* and to unresolved infection in the organs, such as the spleen and lungs, even though the mice were treated with LPC. Figure 3 shows that the LPC-treated mice had significantly but not totally decreased the production of the proinflammatory cytokines. Clinically, the occurrence of pneumonia without bacteremia is frequent, and a large percentage of patients with this infection die during hospitalization (27).

LPC is a chemotactic factor that stimulates immune cells and regulates the balance between the release of pro- and anti-inflammatory cytokines. The beneficial effects of pretreating bacterial infections with LPC in the short term that we have shown here far exceed the potential effects of bacterial infection, as LPC could prevent the release of proinflammatory cytokines and increase the release of anti-inflammatory cytokines. The next issue that must be addressed is to determine whether multiple doses of LPC, given as treatment in combination with antimicrobials, will have a major impact on bacterial

### Table 1 Therapeutic effect of LPC in murine peritoneal sepsis and pneumonia models of *A. baumannii* ATCC 17978 infection

<table>
<thead>
<tr>
<th>Infection model</th>
<th>Treatment group (n)</th>
<th>Survival (%)</th>
<th>Log₁₀ CFU/g spleen (mean ± SEM)</th>
<th>Log₁₀ CFU/g lung (mean ± SEM)</th>
<th>% sterile blood cultures</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pneumonia</td>
<td>Control (10)</td>
<td>0</td>
<td>ND</td>
<td>9.45 ± 0.4</td>
<td>20</td>
</tr>
<tr>
<td></td>
<td>25 mg/kg LPC (15)</td>
<td>68.75*</td>
<td>ND</td>
<td>6.96 ± 0.67*</td>
<td>44</td>
</tr>
<tr>
<td>Peritoneal sepsis</td>
<td>Control (13)</td>
<td>0</td>
<td>9.72 ± 0.09</td>
<td>8.74 ± 0.18</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>25 mg/kg LPC (15)</td>
<td>40</td>
<td>5.82 ± 0.76*</td>
<td>6.37 ± 0.77*</td>
<td>60*</td>
</tr>
</tbody>
</table>

* P < 0.05 with respect to the control group.

![FIG 3](http://aac.asm.org/) Effects of LPC on cytokine production in a murine peritoneal sepsis model of *A. baumannii* infection. Mice received preemptive therapy with LPC and were then inoculated with ATCC 17978. The levels of TNF-α, IL-6, IL-1β, and IL-10 in the serum were determined from 0 to 12 h. Representative results are shown, and the data are presented as means ± SEM. *, P < 0.05 for comparison of ATCC 17978 and LPC.
burdens and inflammatory responses, as a first step before evaluation in clinical studies. Moreover, in addition to this preclinical evaluation, we need to know which populations should be pretreated with LPC. Appropriate LPC clinical use should include patients at risk for severe A. baumannii infections, which have a high morbidity and mortality and usually are caused by multidrug-resistant strains, such as ventilator-associated pneumonia, bacteremia, and wound infections, among others (28). Additionally, the positive effect of LPC on bacterial burdens should be useful and may be used to complement treatment with antimicrobial agents for patients who are suddenly at risk for acquiring infection, such as patients who have extensive burns or trauma and patients admitted to an intensive care unit during an A. baumannii outbreak or epidemic.

In summary, the present study suggests the therapeutic application of LPC as a preemptive therapy for patients at risk of severe infections caused by A. baumannii.

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