The current recommendations for intravenous (i.v.) acyclovir dosing in obese patients suggest using ideal body weight (IBW) rather than total body weight (TBW). To our knowledge, no pharmacokinetic analysis has validated this recommendation. This single-dose pharmacokinetic study was conducted in an inpatient oncology population. Enrollment was conducted by 1:1 matching of obese patients (>190% of IBW) to normal-weight patients (80 to 120% of IBW). All patients received a single dose of i.v. acyclovir, 5 mg/kg, infused over 60 min. Consistent with current recommendations, IBW was used for obese patients and TBW for normal-weight patients. Serial plasma concentrations were obtained and compared. Seven obese and seven normal-weight patients were enrolled, with mean body mass indexes of 45.0 and 22.5 kg/m², respectively. Systemic clearance was substantially higher in the obese than normal-weight patients (mean, 19.4 ± 3.5 versus 14.3 ± 5.4 liters/h; \( P = 0.047 \)). Area under the concentration-time curve was lower in the obese patients (15.2 ± 2.9 versus 24.0 ± 9.4 mg·h/liter; \( P = 0.011 \)), as was maximum concentration (5.8 ± 0.9 versus 8.2 ± 1.3 mg/liter; \( P = 0.031 \)). Utilization of IBW for dose calculation of i.v. acyclovir in obese patients leads to lower systemic exposure than dosing by TBW in normal-weight patients. While not directly evaluated in this study, utilization of an adjusted body weight for dose determination appears to more closely approximate the exposure seen in normal-weight patients. (This study has been registered at ClinicalTrials.gov under registration no. NCT01714180.)
The Institutional Review Board of West Virginia University. Written informed consent was obtained before patient enrollment.

**Inclusion/exclusion criteria.** Patients at least 18 years of age requiring acyclovir as part of routine clinical care were screened for inclusion. Patients were excluded if they had a serum creatinine (Scr) level of >1.4 mg/dl, exhibited clinical instability (defined as ICU admission or receipt of vasoressor support in the prior 24 h), were receiving medications known to interact with acyclovir, or had received acyclovir or valacyclovir in the previous 24 h. Enrollment was conducted by 1:1 matching of MO patients (TBW greater than or equal to 190% of IBW, calculated by the Devine equation [12]) to NW patients (TBW from 80 to 120% of IBW) by gender and by an age of ≥10 years.

**Acyclovir administration.** Patients received i.v. acyclovir sodium, 5 mg/kg, utilizing IBW for MO patients and TBW for NW patients, consistent with current recommendations. Acyclovir was prepared in 100 ml of 0.9% sodium chloride and infused over 60 min via an infusion pump.

**Acyclovir concentration determination.** Blood samples were collected serially immediately prior to the first dose of i.v. acyclovir and at 30, 60, 75, 90, 120, 180, 300, 420, 540, and 720 min following initiation of the infusion. Blood samples were immediately placed on ice and subsequently centrifuged for isolation of plasma by the WVU Health Sciences Biospecimen Processing Core. Samples were stored at −20°C until analysis.

Samples were analyzed within 30 days of collection at NMs Labs (Wil- low Grove, PA) using liquid chromatography–tandem mass spectrometry that was internally validated. Briefly, labeled internal standard (acyclo- vid-d4) was added to diluted plasma samples, which were then deproteinized with trichloroacetic acid. The supernatant was analyzed by high-performance liquid chromatography separation using positive-ion electrospray tandem mass spectrometry (LC-MS/MS) for detection and quantitation. Calibration curves were constructed to quantify the concentration of acyclovir using a quadratic regression with 1/x weighting. The lower limit of quantitation for acyclovir was 0.020 μg/ml with additional calibration points at 0.050, 0.10, 0.50, 1.0, and 2.0 μg/ml. Within-run precision for the high, low, and lowest observable quantities assay was 2.6 to 5.2%, whereas between-run values were 3.7 to 6.8%. Similarly, assay accuracy for the high, low, and lowest observable quantities were 92 to 95% and 94 to 96% for within-run and between-run values, respectively. Plasma samples with initial concentrations greater than 2 μg/ml were diluted until the concentration was within the range of the assay. Two levels of quality control, 0.050 and 1.0 μg/ml, were prepared in bulk and frozen. These were utilized to evaluate each analytic run. Coefficients of variation for the assay were 7.69% and 10.20% for the high and low controls, respectively.

**Acyclovir pharmacokinetics.** Pharmacokinetic parameter estimates for each patient’s data set were generated by the standard two-stage approach using compartmental modeling (WinNonlin version 2.1; Phar- sight Corporation, Mountain View, CA) by the WVU Health Sciences Clinical Pharmacology Core. Selection of the most appropriate model for each patient’s data was primarily based on the Akaike information criterion. The absolute dose (nonnormalized) was utilized as the input variable for all analyses. Actual sample times obtained from initiation of the dose were calculated from the case report forms and used as primary input data. The optimal model was utilized to generate simulated data for estimation of the individual patients’ IC50 at steady state with 12-h dosing intervals. Evaluations of T > IC50 were conducted using IC50 of 0.5625 mg/liter for HSV and 1.125 mg/liter for VZV, which had been reported previously and correspond to more resistant strains (8, 13).

**Statistics.** A sample size of 7 patients per group was estimated to provide an 80% power to detect a 19% difference in CL using a two-sided paired t test at significance level of 0.05 when the correlation coefficient is 0.15. In the data analysis of comparison on the primary and secondary outcomes, Wilcoxon signed-rank test was used for continuous variables with paired data and Wilcoxon rank sum test was used for continuous variables between two groups, while Fisher’s exact test was used in the data analysis between categorical variables. To explore optimal dosing strategies, CL was compared between MO and NW patients after normalizing by different measures of body size, including body surface area (BSA), TBW, IBW, lean body weight (LBW), and adjusted body weight (AjBW). AjBW was calculated as IBW plus 40% of TBW greater than IBW [AjBW = IBW + 0.4 × (TBW − IBW)] and LBW as previously described (14). Simple linear regression was performed to assess correlation of these body parameters with PK parameters. Analyses were considered statistically significant if P was <0.05. All statistical analyses were performed using Stata (Stata statistical software, release 13, 2014; StataCorp LP, College Station, TX) and R software (R Foundation for Statistical Computing, Vienna, Austria; [http://www.R-project.org/]).

## RESULTS

A total of 14 patients were enrolled and completed the study (7 patients per group). Baseline data were similar between groups with the exception of TBW, BMI, percentage of IBW, and BSA (Table 1). All patients in the MO group were classified as class III obesity (BMI, ≥40 kg/m2). All patients were receiving acyclovir for prophylaxis with chemotherapy regimens with an anticipated prolonged duration of neutropenia. None of the patients in the study were febrile or neutropenic or had any signs of viral infection at the time of sampling.

CL was significantly higher in MO than NW patients, while \( \text{AUC}_{\text{0-\infty}} \) and \( C_{\text{max}} \) were significantly lower in MO patients (Table 2; Fig. 1). This difference in \( \text{AUC}_{\text{0-\infty}} \) represents a 37% (95% con-

### Table 1 Baseline characteristics

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Value for patientsa</th>
<th>Normal wt</th>
<th>( P^b )</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (yr)</td>
<td>Morbidly obese (n = 7)</td>
<td>54.3 ± 9.6</td>
<td>53.0 ± 16.3</td>
</tr>
<tr>
<td></td>
<td>Caucasian race (%)</td>
<td>7 (100.0)</td>
<td>7 (100.0)</td>
</tr>
<tr>
<td></td>
<td>Female (%)</td>
<td>6 (85.7)</td>
<td>6 (85.7)</td>
</tr>
<tr>
<td>Weight (kg)</td>
<td>120.5 ± 15.7</td>
<td>61.2 ± 5.1</td>
<td>0.016</td>
</tr>
<tr>
<td>IBW (kg)</td>
<td>57.1 ± 8.8</td>
<td>58.5 ± 5.5</td>
<td>0.69</td>
</tr>
<tr>
<td>% of IBW</td>
<td>212.4 ± 15.4</td>
<td>105.2 ± 10.7</td>
<td>0.001</td>
</tr>
<tr>
<td>BMI (kg/m2)</td>
<td>45.0 ± 3.4</td>
<td>22.5 ± 2.2</td>
<td>0.016</td>
</tr>
<tr>
<td>BSA (m2)</td>
<td>2.3 ± 0.2</td>
<td>1.7 ± 0.1</td>
<td>0.016</td>
</tr>
<tr>
<td>SCR (mg/dl)</td>
<td>0.78 ± 0.26</td>
<td>0.76 ± 0.15</td>
<td>0.69</td>
</tr>
<tr>
<td>GFR (ml/min/1.73 m2)</td>
<td>93.4 ± 24.9</td>
<td>93.7 ± 25.7</td>
<td>0.94</td>
</tr>
</tbody>
</table>

a BMI, body mass index; BSA, body surface area; GFR, glomerular filtration rate; IBW, ideal body weight; Scr, serum creatinine.
b Data are means ± standard deviations unless otherwise noted.
c Determined by the Wilcoxon rank sum or Fisher exact test, as appropriate.

### Table 2 Comparison of mean pharmacokinetic parameters

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Value for patientsa</th>
<th>Normal wt</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dose (mg)</td>
<td>Morbidly obese (n = 7)</td>
<td>285 ± 44</td>
</tr>
<tr>
<td>( C_{\text{max}} ) (mg/liter)</td>
<td>5.8 ± 0.9</td>
<td>8.2 ± 1.3</td>
</tr>
<tr>
<td>( \text{AUC}_{\text{0-\infty}} ) (mg.hr/liter)</td>
<td>15.2 ± 2.9</td>
<td>24.0 ± 9.4</td>
</tr>
<tr>
<td>Time &gt; 0.5625 mg/liter (min)</td>
<td>402.6 ± 204.2</td>
<td>524.3 ± 253.0</td>
</tr>
<tr>
<td>Time &gt; 1.125 mg/liter (min)</td>
<td>264.9 ± 54.5</td>
<td>373.1 ± 181.6</td>
</tr>
<tr>
<td>CL (liters/h)</td>
<td>19.4 ± 5.3</td>
<td>14.3 ± 5.4</td>
</tr>
<tr>
<td>V (liters)</td>
<td>31.8 ± 9.9</td>
<td>25.9 ± 10.4</td>
</tr>
</tbody>
</table>

a \( \text{AUC}_{0-\infty} \) area under the curve from time zero to infinity; CL, systemic clearance; \( C_{\text{max}} \) maximum concentration; V, volume of distribution.
b Determined by the Wilcoxon signed-rank test.
DISCUSSION

The objective of this study was to evaluate the currently recommended dosing strategy for i.v. acyclovir in MO patients. The PK parameters observed in the current study for NW patients are similar to those previously reported for healthy, nonobese patients. Average CL for NW patients in our study (14.6 liters/h/1.73 m²) was similar to that previously reported by Laskin and colleagues (16.1 liters/h/1.73 m²) and Blum and colleagues (19.6 liters/h/1.73 m²) (15, 16). Laskin and colleagues reported a similar AUC of 23.2 mg · h/liter; P = 0.49). While these data are based on this single-dose analysis, we expect that dosing over several days would provide similar results.

While previous data (10, 11) have shown that using TBW for dose determination leads to excessive acyclovir exposure in obese patients, our study found that dosing by IBW will provide substantially lower exposure than in nonobese controls. Using patient-specific PK parameters in the MO patients, utilizing an AjBW to dose acyclovir would result in similar exposure (AUC∞) compared to our NW patients. While we agree with Davis and colleagues (10) that TBW leads to excess exposure in MO patients, dosing by AjBW (correction factor of 0.4) may more closely approximate drug exposure in NW patients. Utilizing BSA to dose obese patients may also result in exposure similar to that in NW patients; however, dosing by BSA is uncommon with antimicrobials and may lead to additional difficulties, as the dosing recommendation for i.v. acyclovir in NW patients is based on weight.

While the difference did not reach statistical significance, we found MO patients to have lower T>IC₅₀ than NW patients (Table 2). With dosing every 12 h, a similar number of MO and NW patients achieved the T>IC₅₀ goal of 50% in plasma for both HSV (57.1% and 71.4% for MO and NW patients; P = 1.0) and VZV (14.3% and 42.9% for MO and NW patients; P = 0.56). Note that achievement of this target is based on samples collected from blood and is likely different from PD parameters in other body compartments.

Limited studies evaluating dosing of acyclovir and valacyclovir for treatment of genital herpes suggest that AUC and T>IC₅₀ are both associated with efficacy (4–7). It should be noted that genital herpes is commonly treated with oral rather than i.v. acyclovir. Unfortunately, trials evaluating the PD of i.v. acyclovir in treating
more invasive infections, such as HSV encephalitis, are lacking, and extrapolation of PD targets may not be appropriate.

Several important limitations exist in this study. First, our study was conducted with lower doses of i.v. acyclovir (5 mg/kg). While escalating doses result in linear increases in AUC and similar V (15, 17), caution should be exercised in extrapolating these data to other dosing regimens. Second, these patients were all receiving acyclovir for prophylaxis and were not actively infected or critically ill. Altered PK are often present in patients that are critically ill (18, 19). Third, our study evaluated the PK from blood samples only. Concentrations at different body sites, such as in cerebral spinal fluid, were not evaluated in this study and the PK parameters obtained in our study cannot be easily transposed to these body sites. Fourth, we evaluated only obese patients with BMIs of \( \geq 40 \) kg/m\(^2\) (class III obesity) and did not include any patients with BMIs from 25.0 to 39.9 kg/m\(^2\). Finally, these were cancer patients receiving active chemotherapy treatments, but we have no reason to believe this would affect the PK characteristics with i.v. administration compared to nononcology patients.

Conclusions. Our data suggest that MO (BMI, \( \geq 40 \) kg/m\(^2\)) patients treated with i.v. acyclovir dosed by IBW experience substantially decreased overall exposure compared to NW patients dosed by TBW. While not directly evaluated in this study, utilization of an AjBW (IBW + 0.4 \( \times \) (TBW − IBW)) appears to more closely approximate the exposure seen in NW patients. Future research is needed to verify this finding and to explore appropriate dosing in this population and in other classes of obesity.

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