Development of NS3/4A protease-based reporter assay suitable for efficiently assessing hepatitis C virus infection

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Running title:
A protease-based assay for screening of anti-HCV inhibitors

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Abstract

A cell culture system for the production of hepatitis C virus (HCV) whole virions has greatly accelerated studies of virus life cycle and discovery of anti-HCV agents. However, the quantification of the HCV titers in a whole virus infection/replication system currently relies mostly on reverse transcription-PCR (RT-PCR) or immunofluorescence assay (IFA), which would be cumbersome for high throughput drug screening. To overcome this problem, this study has generated a novel cell line, Huh7.5-EG(Δ4B5A)SEAP, that carries a dual reporter EG(Δ4B5A)SEAP. The EG(Δ4B5A)SEAP reporter is a viral protease-cleavable fusion protein in which the enhanced green fluorescence protein (EGFP) is linked to secreted alkaline phosphatase (SEAP) in-frame via Δ4B5A, a short peptide cleavage substrate for NS3/4A viral protease. This study demonstrates that virus replication/infection in the Huh7.5-EG(Δ4B5A)SEAP cells can be quantitatively indicated by measuring the SEAP activity in cell culture media. The levels of SEAP released from HCV-infected Huh7.5-EG(Δ4B5A)SEAP cells correlated closely with the amounts of HCV inoculums. The Huh7.5-EG(Δ4B5A)SEAP cells were also shown to be a suitable host for the discovery of anti-HCV inhibitors using known compounds that target multiple stages of the HCV life cycle. Z'-factor value of this assay ranged from 0.64 to 0.74 in 96-well plates; indicating that this reporter system is suitable for
high-throughput screening of prospective anti-HCV agents.
Introduction

Infection with Hepatitis C virus (HCV) is a major global health problem. More than 170 million individuals are infected worldwide. The infection is strongly associated with the risk of liver cirrhosis and hepatocellular carcinoma (42, 43). HCV is a single, positive-stranded RNA genome of approximately 9.6 kb, which encodes a polyprotein of about 3,000 amino acids (38). Cellular and viral proteases are involved in processing viral polyprotein into at least ten proteins (core, E1, E2, p7, non-structure protein 2 (NS2), NS3, NS4A, NS4B, NS5A, and NS5B) and the cleavages at the NS3-4A, NS4A-4B, NS4B-5A, and NS5A-5B junctions were mediated by NS3/4A protease (13, 24, 26). In the past few years, subgenomic HCV replicons have been employed extensively for studying viral replication, protein processing, virus-host interactions, and for discovering anti-HCV agents (4, 15, 16, 29, 34, 52). However, such subgenomic systems are not useful for studying the entry, assembly or release of viral particles because they lack structure proteins. In 2005, an infectious clone was isolated from a Japanese patient with fulminant hepatitis (JFH1) (44). The advent of the JFH1 isolate represented an important breakthrough for the investigation of the full viral life cycle as well as HCV interactions with host cells. Notably, this system is also useful for the testing of antiviral agents targeting viral attachment, entry, trafficking, assembly and release. Recently, JFH1-based intergenotypic HCV virions
were generated by replacing the JFH1 structure proteins region to that of other strains (12, 40, 46, 47, 50). These chimeras may be important to the development of effective antivirals and vaccines against all genotypes. Several robust virus-producing systems have also led to important advances in studies of HCV (5, 50, 51). In particular, a stably JFH1-cDNA-transfected human hepatoma cell line, Huh7-GL, is a valuable tool for the robust and durative production of infectious virus. The JFH1 virion generated from the Huh7-GL cells was utilized in this work.

To elucidate the viral life-cycle and the inhibitory effects of anti-HCV agents, several methods have been employed to measure the degree of HCV infection/replication. They include quantitative reverse transcription-PCR (qRT-PCR) to determine RNA levels or Western blot analysis to determine the expression levels of proteins (2, 12). These methods are time-consuming and labor-intensive because the need to prepare cell lysates and to purify RNA. Therefore, a high-throughput screening system for accelerating the discovery and development of anti-HCV therapeutics is urgently needed. Recombinant JFH1 viruses that contain luciferase or GFP reporter gene have recently been generated to investigate viral infection (1, 15, 25, 50). However, the associated common problem of these constructions is the low yield of viral progeny possibly due to the insertion of undesirable elements to impair RNA replication. We
have developed an efficient reporter-based assay for monitoring HCV protease activity and the degree of subgenomic replication (29-31). These studies involved the construction of a substrate vector, pEG(Δ4AB)SEAP, which contained a dual-reporter gene. The Enhanced Green Fluorescent Protein (EGFP) gene was separated from another SEcreated Alkaline Phosphatase (SEAP) reporter gene by fragment encoding a cleavable peptide substrate of the viral NS3/4A protease. These systems exhibited not only a high signal-to-noise ratio but also a close relationship between SEAP activity and HCV subgenomic replication.

In this study, we have established a stable reporter cell line, Huh7.5-EG(Δ4B5A)SEAP, that can be used to determine the JFH1 infection via the quantification of SEAP activity in culture medium. This system has allowed us to study detailed kinetic of viral infection in real-time without the preparation of cell lysates, isolation of RNA, and the following qRT-PCR experiments. Several known inhibitors that target various stages in the viral life cycle are used to validate that this novel Huh7.5 cell-based system can indeed by utilized to screen anti-HCV agents of different modes of action. Statistical analyses were performed with anti-HCV inhibitors treatment in a 96-well format and the results indicated that both the accuracy and sensitivity of this assay system are suitable for high-throughput
screening (HTS) of anti-HCV drugs.
Materials and Methods

Cells and virus

Huh7.5 cells were generously provided by Dr. C. Rice (Rockefeller University, New York, USA) and were maintained in Dulbecco’s modified Eagle’s medium (DMEM) supplemented with 10% fetal bovine serum (FBS, Life Technologies) plus penicillin (50 U/ml), streptomycin (50 mg/ml), and nonessential amino acids as a basal medium. Huh7.5 cells were a mutant line of Huh7 cells and were highly permissive for HCV RNA replication (3). Huh7-GL cells were kindly provided by Dr. Takaji Wakita (Tokyo Metropolitan Institute for Neuroscience, Tokyo, Japan) and were maintained in basal medium plus sodium pyruvate and 5 µg/ml blasticidin (Invitrogen). Huh7-GL cells contain a chromosomally integrated genotype 2a HCV cDNA and continuously generate infectious virus (JFH1)(5). In this study, stably transfected Huh7.5 cells, Huh7.5-EG(Δ4B5A)SEAP cells, expressing EG(Δ4B5A)SEAP fusion protein were maintained in basal medium plus 0.5 mg/ml G418.

Plasmid construction

In this study, we used pEGFP-C1 plasmid (Clontech) as a backbone vector to construct pEG(Δ4A4B)SEAP, pEG(Δ4B5A)SEAP, and pEG(Δ5A5B)SEAP. The SEAP gene was amplified by PCR using pSEAP2-control (Clontech) as a template. The forward primers, 5′-CGG GGT ACC GAT GAG ATG GAG GAA TGC GCC
TCT AGG GCG ATG CTG CTG CTG CTG CTG CTG CTG GGC CTG -3', 5'-CGG GGT ACC ACT GAG GAC TGC CCC ATC CCA TGC TCC GGA TCC TGG CTC CGC GAC GTG ATG CTG CTG CTG CTG CTG CTG GGC CTG-3', and 5'-CGG GGT ACC GAG GAC GAT ACC ACC GTG TGC TGC TCC ATG TCA TAC TCC TGG ACC GGG ATG CTG CTG CTG CTG CTG CTG CTG GGC CTG-3', contain the junction sites (genotype 2a HCV cDNA sequences) of NS4A/NS4B, NS4B/NS5A, and NS5A/NS5B, respectively, encoding in-frame amino acid sequences, Δ4A4B (DEMEECASRA), Δ4B5A (TEDCPICSGSWLDRV), and Δ5A5B (EDDTVCCMSYSWTG), as substrates of HCV NS3/4A protease. The reverse primer sequence was 5'-GTG ATT GGT ACC TTC ATG T CT GCT CG-3'. The PCR product was digested with KpnI and inserted into the KpnI site of pEGFP-C1.

**Transient transfection analysis of plasmid**

Huh7.5 cell lines were seeded at a density of 1 × 10^5 cells per well in 24-well plates and were incubated at 37°C overnight. Subsequently, cells were transfected with 0.2 µg of individual reporter plasmids by FuGENE 6 reagent (Roche). The transfection procedure was according to the manufacturer’s instruction. The plasmid-transfected Huh7.5 cells were infected with JFH1 virus at an MOI of 0.02 in each well for 6 h. At the end of infection, unbound viruses were removed by medium replacement and infected cells were further cultured in phenol red-free basal medium. At 6 days postinfection, SEAP activity was measured following one more medium replacement.
for 24 h. The expression of EGFP within reporter genes was served as an internal control and each experiment was normalized by the average number of EGFP-positive cells that detected in multiple foci.

**Stable cell line selection**

Huh7.5 cells were seeded at a density of $5 \times 10^4$ cells per well in a 24-well plate. After incubation at 37°C overnight, cells were transfected with 0.2 µg of pEG(Δ4B5A)SEAP plasmid in single well. After 2 day posttransfection, cells were maintained in basal medium plus 0.5 mg/ml G418 for selection of neomycin-resistance clones. After approximately one week selection, expended clones were examined for the expression of EG(Δ4B5A)SEAP fusion protein and the extracellular SEAP activity under the proteolytic digestion of NS3 protease using western blot analysis and Phospha-Light™ assay kit (Applied Biosystems), respectively. The stable cell line was designated Huh7.5-EG(Δ4B5A)SEAP.

**Western blot analysis**

In 24 well plates, Huh7.5-EG(Δ4B5A)SEAP cells were seeded at a density of $1 \times 10^5$ cells per well and infected with JFH1 virus at an MOI of 0.02 in each well for 6 h.

Huh7.5-EG(Δ4B5A)SEAP cells infected or non-infected by JFH1 were incubated for
3 days and washed with phosphate buffered saline (PBS). Cells were lysed in lysis buffer (50 mM Tris-HCl, 150 mM sodium chloride, 1% Nonidet P40, and 0.5% sodium-deoxycholate) and the cell lysates were collected and clarified by centrifugation at 15,000 × g for 10 min at 4°C. The protein concentration of cell extracts was measured by using a protein assay buffer (Bio-Rad). The equal amounts of protein samples were resolved by 12% SDS-polyacrylamide gel electrophoresis and then proteins in gel were transferred to a polyvinylidene difluoride membrane (Millipore). The membrane was blocked with NET buffer (0.25 g/l gelatin, 150 mM sodium chloride, 5 mM disodium ethylenediaminetetraacetate dihydrate, 50 mM Tris, and 0.05% Tween-20) for 1 hr at room temperature. The expression level of EG(Δ4B5A)SEAP fusion protein and free form EGFP were detected using mouse polyclonal anti-EGFP antibody. The intracellular NS3 protease of HCV-JFH1-infected cells was detected using mouse monoclonal anti-NS3 antibody (Virostat). The membrane was also probed with mouse monoclonal anti-actin antibody (Santa cruz biotechnology). The amounts of β-actin served as loading control. After primary antibody incubation, membrane was washed with TBST buffer (150 mM sodium chloride, 0.1% Tween-20, 50 mM Tris-HCl,) and probed with horseradish-peroxidase-conjugated goat anti-mouse IgG antibody. Finally, the membrane was treated with ECL reagent and exposed to X-ray film.
Detection of SEAP activity released from HCV-infected Huh7.5-EG(∆4B5A)SEAP cells

HCV-infected Huh7.5-EG(∆4B5A)SEAP cells were continually maintained in phenol red-free basal medium containing 0.5 mg/ml G418. At 5 days postinfection, culture medium was replaced and cells were incubated one more day. Subsequently, SEAP activity in culture medium was detected using Phospha-Light™ assay kit (Applied Biosystems) and cell viability was measured using MTS assay as previously described (8).

Quantitative, Reverse-Transcription Polymerase Chain Reaction

Total cellular RNA and virus RNA were extracted using TRIZOL reagent (Invitrogen) and QIAamp® Viral RNA mini kit (Qiagen), respectively. One microgram of total RNA was performed the reverse transcription reaction using M-MLV RT kit (Promega). Briefly, RNA sample was mixed with 1µM of random Hexamer primer and allowed to unfold the secondary structure of RNA by heating for 8 min at 65°C. The Reverse transcription reaction was carried out by following condition: annealing at 25°C for 10 min, extension at 42°C for 90 min and denaturation at 95°C for 5 min. A 20 µl of reaction volume contained 200 U of M-MLV-RT RNase H(-) reverse
transcriptase, 1 mM each deoxynucleotide triphosphate, and 20 U of RNase OUT reagent (Invitrogen).

Two microliter of reverse transcription reaction was used in Quantitative PCR (Q-PCR) that was performed on a LightCycler (Roche) with a LightCycler® DNA master SYBR Green I kit, according to the manufacturer’s instructions. Primers for HCV (genotype 2a) cDNA were forward, 5'-CCC GTT GAG ACA CTC G-3'; reverse reverse primer, 5'-CCT AAT GTT GGG ATT GAT GC-3'. Primer for GAPDH were forward, 5'-GAA GGT GAA GGT CGG AGT C-3'; reverse, 5'-GAA GAT GGT GAT GGT GGG ATT TC-3'. The Q-PCR reaction condition was as follows: denaturation at 95°C for 600 s, 45 cycles of amplification at 95°C for 0 s, 64°C for 5 s, and 72°C for 15 sec. At the end of Q-PCR procedure, the PCR cycle numbers of each sample was converted to the relative copy number of HCV RNA by according to the calibration curve. The calibration curve was made from serial 10-fold dilutions of reference DNA that consists of a full-length HCV (genotype 2a) cDNA. The copy number of intracellular HCV RNA in each sample was normalized by dividing the corresponding GAPDH copy numbers. All experiments were performed in triplicates to determine statistical parameters.

**Enzymatic assays for the HCV NS3/4A protease**
At 6 days postinfection, HCV-infected Huh7.5-EG(Δ4B5A)SEAP cells were washed with PBS and lysed in lysis buffer without protease inhibitor. The cell lysates were collected and clarified by centrifugation at 15,000 × g for 10 min at 4 °C. The protein concentration of cell extract was measured and 50 µg of total protein was performed on HCV NS3/4A protease assay. The HCV NS3/4A protease activity was determined by using an internally quenched fluorogenic depsipeptide (HCV Protease FRET Substrate (RET S1)), Ac-DED(EDANS)EEαAbuψ[COO]ASK(DABCYL)-NH₂, (AnaSpec Inc.) in a black F96 MicroWell™ plate (Nunc) (11). The cell extract was incubated with reaction buffer (50 mM Tris-HCl (pH 7.5), 50 % glycerol, and 30 mM dithiothreitol). The reaction was started by the addition of 1.5 µM RET-S1 substrate and incubated at 25°C. The reaction volume was 100 µl. The assay was run in a kinetics model for 10 min and product release was detected using a Fluoroskan Ascent FL plate reader (Thermo Scientific) with excitation at 355 nm and emission at 510 nm. The protease activity was determined by the linear slope of product release with time.

Preparation of HCV-JFH1 stock

Approximately 30-40% confluence of Huh7-GL cells in a T150 flask were maintained
in basal medium without 0.5 ug/ml blasticidin. After incubation for 4 days, culture medium was collected and cleared by using low-speed centrifugation (1,000 × g). The supernatant was passed through a 0.45-µm low-protein binding filter. The filtrate was concentrated 1/50 using an Amicin Ultra-15 Centrifugal Filter, 100 kDa, (Millipore) (44) and the remained filtrate was aliquoted for storage at -80°C in 15% glycerol. The amount of infectious HCV-JFH1 in virus stock was measured by immunofluorescence assay.

**Titration of infectious HCV-JFH1**

In this study, we estimated the amount of infectious HCV-JFH1 in viral stock by immunofluorescence assay (51). Briefly, Viral stock was serially diluted 10-fold in DMEM and then used to infected 8 × 10^3 Huh7.5 cells per well in 96-well plates. Virus inoculum after 6 h, the supernatants were removed and culture medium were supplemented. After 4 days, the cells were fixed with 4% paraformaldehyde and permeabilized with 0.5% Triton X-100. Subsequently, fixed cells were blocked with 5% bovine serum albumin for 1 h at room temperature. The intracellular HCV NS3 proteins in infected cells were detected by incubation with mouse monoclonal anti-NS3 antibody (1:20 dilution) (Virostat) for 1 h at room temperature. Rhodamine
(TRITC)-conjugated AffiniPure Donkey anti-mouse IgG (H+L) (1:200 dilution) (obtained from optical biology core lab in core facilities of NHRI) was applied to visualize the positive cells. The number of HCV foci in each dilution is counted and viral titer is expressed as focus-forming units per milliliter (FFU/ml).

**Effects of anti-CD81 antibody and H-89 on HCV-JFH1 infection**

Huh7.5-EG(Δ4B5A)SEAP cells were seeded at a density of $8 \times 10^3$ cells per well in 96 well plates. After incubation overnight, cells were treated with increasing concentrations of anti-CD81 antibody (Santa Cruz Biotechnology, Inc.) or H-89 (Calbiochem) in basal medium for 1 h and infected with JFH1 virus at an MOI of 0.02 per well for 6 h in the presence of inhibitor. At the end of infection, unbound virus/inhibitors were removed by medium replacement and HCV-infected cells were continually cultured. At 5 days postinfection, the culture medium in each well was replaced with phenol red-free basal medium and cells were cultured one more day. Subsequently, SEAP activity in culture medium was measured and cell viability was estimated by MTS assay in the meanwhile. Each experiment was performed in triplicates for statistical analysis.
Effects of naringenin, BILN2061, and IFN-α on intracellular activity of HCV

To estimate the levels of secreted HCV-JFH1 from various compounds-treated Huh7-GL cells, a density of $1.5 \times 10^4$ cells per well in 96-well plates were treated with increasing concentrations of reference compounds, as naringenin (Sigma), BILN2061 (Acme Bioscience, Inc.), and IFN-α (Sigma) for 3 days. At the end of treatment, the supernatants containing secreted JFH-1 were collected and cell viability was estimated by MTS assay. The level of secreted JFH-1 was detected by infecting Huh7.5-EG(Δ4B5A)SEAP cells and HCV-infected cells were further culture. At 5 days postinfection, the culture medium in each well was performed medium replacement. Subsequently, SEAP activity and cell viability in each well was measured at 6 days postinfection.

Calculation of S/B, S/N, and Z’ factor

To evaluate the performance of this developed assay for HTS, S/B, S/N, and Z’ factor values were calculated using the method of Zhang et al (49). In HCV-JFH1 entry inhibition, Huh7.5-EG(Δ4B5A)SEAP cells were seeded at a density of $8 \times 10^3$ cells per well in a single 96-well plate overnight. H-89 was added to the 30 wells for 1 h at
a volume of 100 µl and a final concentration of 10 µM in 1% DMSO (v/v) as a background set. One % DMSO (v/v) was added to the other 30 wells as a signal set. Subsequently, cells were infected with JFH1 virus at an MOI of 0.02 per well for 6 h in the presence of H-89 or 1% DMSO. At the end of infection, unbound virus and compounds were removed by medium replacement and HCV-infected cells were further cultured. At 6 days postinfection, medium was collected from each well for SEAP activity analysis and cell viability was measured. In the intracellular HCV-JFH1 activity, Huh7-GL cells were seeded at a density of $1.5 \times 10^4$ cells per well in a single 96-well plate. Cells in the 30 wells were treated with 100 nM BILN2061 in 1% DMSO (v/v), as a background set, for 3 days while Cells in the other 30 wells were added 1% DMSO (v/v), as a signal set, for 3 days. Three days later, supernatant media were collected and cell viability was measured. Subsequently, Huh7.5-EG(Δ4B5A)SEAP cells at a density of $8 \times 10^3$ cells per well in a 96-well plate treated with collected supernatant media for 6 h by plate to plate manner. At the end of treatment, reporter cells were further culture for 6 days followed by detection of SEAP activity and cell viability. Finally, the data were collected and S/B, and S/N, and $Z'$ factor were determined.
Data analysis and statistics

Data analysis for SEAP activity and cell viability were performed with the SigmaPlot software (SPSS Inc.). The SEAP signal data were presented as mean ± SD. Statistical significant effect between groups was examined using the paired Student’s t-test. Statistical significance was defined as P < 0.01.
Results

Characterization of HCV-JFH1 infection in Huh7.5-EG(Δ4B5A)SEAP cells

To develop a convenient tool for monitoring HCV-JFH1 infection, several reporter plasmids were constructed based on our earlier reporter-based assay system that involves the HCV Con1 strain of genotype 1b subgenomic replicon (30), in which the EGFP was fused to SEAP via three prospective peptide linkers corresponding to the JFH1 genotype 2a NS3 protease cleavage sites at various junctions. The amount of SEAP that is secreted into the medium depends on the expression of NS3 protease following HCV infection. The evaluated cleavage peptides of JFH genotype 2a include DEMEEC-ASRA; (designated Δ4A4B), TEDCPIPC-SGSWLRDV; (designated Δ4B5A), and EDDTTVCC-SMSYSWGT; (designated Δ5A5B) (Fig. 1A). Initially, those reporter constructs were transiently transfected into Huh7.5 cells to examine whether they could be efficiently cleaved by NS3 protease following HCV infection. At 6 days postinfection, cell culture media were harvested and analyzed for SEAP activity. Surprisingly, elevated SEAP levels were observed only in media from cells that were transfected with the EG(Δ4B5A)SEAP construct but not in media from the other two constructs (Fig. 1B). This result differed substantially from that in a previous investigation in which the linked Δ4A4B peptide sequence (DEMEEC-ASHL) of the HCV genotype 1b was cleavable by NS3 protease in HCV
replicon cells harboring HCV subgenome of genotype 1b (29). Indeed, we observed that the SEAP activity also can be significantly detectable with EG(ΔA4A4B)SEAP construct containing peptide sequence (DEMEEC-ASHL) following HCV-JFH1 infection (data not shown), suggesting that interference of the cleavage may be caused by different protein conformations when the various peptide sequences were inserted into the chimera fusion protein. The JFH1 viruses were collected from Huh7-GL cells that contained a chromosomally integrated full-length JFH1 HCV cDNA (5).

To confirm that the EG(ΔA5B5A)SEAP fusion protein was indeed cleaved by NS3 protease, an EG(ΔA5B5A)SEAP stably transfected Huh7.5 cell line, designated as Huh7.5-EG(ΔA5B5A)SEAP, was infected with JFH1 virus (Each well of cells in a 24-well plate received a viral inoculum with an MOI of 0.02) for 6 days. Cell lysates were analyzed by Western blot with anti-EGFP antibody and anti-NS3 antibody, in which the levels of β-actin were served as the loading controls. In Figure 2A, high levels of intact EG(ΔA5B5A)SEAP fusion protein were observed in Huh7.5-EG(ΔA5B5A)SEAP reporter cells (lanes 3 and 4, upper panel) but not in the parental Huh7.5 cells (lanes 1 and 2). Upon infection of the Huh7.5 control cells (lane 2) and the Huh7.5-EG(ΔA5B5A)SEAP reporter cells by JFH1 (lane 4), viral replication is revealed by the expression of NS3 viral protease, as determined by the Western blot.
analysis. A distinct band that corresponds to free EGFP was clearly present in lane 4, indicating that JFH1-derived NS3 protease effectively cleaved the EG(Δ4B5A)SEAP into EGFP and SEAP.

The SEAP levels in the culture media were also measured. In Huh7.5 cells without the EG(Δ4B5A)SEAP reporter, only basal levels of SEAP activity were detected in the culture media (Fig. 2B, columns 1 and 2). The SEAP level in the culture medium of the Huh7.5-EG(Δ4B5A)SEAP cells was slightly higher (column 3). The SEAP level in the culture medium of Huh7.5-EG(Δ4B5A)SEAP cells infected with JFH1 virus was many times higher than that of non-infected cells (column 4) presumably because of proteolytic processing of EG(Δ4B5A)SEAP by the NS3 protease.

To optimize the assay conditions, Huh7.5-EG(Δ4B5A)SEAP cells were infected with JFH1 at increasing MOIs, and SEAP activity in each culture medium was analyzed at various time points postinfection. After inoculation with the JFH1 virus, culture media were harvested daily to measure the activities of SEAP. As shown in Figure 3A, increasing MOIs in the inoculum progressively increased the SEAP signals in a time-dependent manner. The SEAP signals became detectable at 3 days postinfection. When Huh7.5-EG(Δ4B5A)SEAP cells were infected with JFH1 virus at an MOI of
0.02, at 6 days postinfection, the SEAP activity was approximately 14.9 ± 0.7-fold higher than that at 2 days. With a low MOI (0.04), or higher MOI (>0.04), at 6 days postinfection, the SEAP activity also reached approximately 15-fold higher than that at 2 days. Such a kinetic of SEAP signals is consistent with the infection kinetics as observed by Zhong et al. using immunofluorescence staining on naïve JFH1-infected Huh7.5.1 cells (51). Notably, the time course of SEAP detection was similar to the results recently described by Yu et al. using a NS3-based Fluorescence Resonance Energy Transfer (FRET) assay for monitoring HCV infection (48). Furthermore, we examined the relationship between intracellular HCV RNA levels and NS3 protease expression during virus infection. It was interesting to found that the induction of SEAP correlated directly with increased NS3 protein levels, but not with HCV RNA (Fig. 3B). We suggest that the slow onset of the SEAP signal may be caused by the lag time for HCV to overcome the innate infection in the reporter cells before HCV start to protein expression efficiently.

Correlation between levels of secreted SEAP activity and HCV-JFH1 replication inside cells

Whether the levels of SEAP activity in culture supernatants were correlated with the extent of HCV replication inside infected cells was examined. The
Huh7.5-EG(Δ4B5A)SEAP cells were inoculated with increasing MOIs and incubated for 6 days. Subsequently, the culture supernatants were analyzed for SEAP activity. The cell lysates were analyzed for both NS3 expression levels and protease activity using Western blotting with anti-NS3 antibody and Fluorescence Resonance Energy Transfer (FRET) assay with an internally quenched fluorogenic depsipeptide of NS3/4A protease substrate, as described in Materials and Methods, respectively. As shown in Figure 4A, a linear relationship exists between the SEAP levels and the amount of the HCV inoculums (panel a). A similar linear relationship was also observed between the NS3/4A protease activity and the titers of virus inoculums (panel a). In panel b of Figure 4A, the viral NS3/4A protease expression levels clearly increased with the amount of the HCV inoculum. The square of the correlation constant ($R^2$) is 0.962, as calculated by linear regression (Fig. 4B). This value indicates that the secretion of SEAP was closely correlated with the abundance of the viral proteins, as reflected by the NS3 expression levels and relative activity. Altogether, in a manner similar to the representation of viral replication using the replicon-based reporter system that we have established previously (29, 30), the secreted SEAP reporter level suitably represented the amount of virus inoculums.

**Evaluations of various kinds of HCV inhibitors in Huh7.5-EG(Δ4B5A)SEAP cells**
that were infected with JFH1 virus

To verify the utility of the JFH1-infected Huh7.5-EG(∆4B5A)SEAP cells as a quantitative antiviral assay system, several known HCV inhibitors that target multiple stages of the HCV life cycle were examined. These HCV inhibitors include 1) anti-CD81 antibody, an entry blocker that interferes with the CD81-E2 interaction (37), 2) H-89, a protein kinase A (PKA) inhibitor that reduces viral receptor activity by inducing the redistribution of CLDN1 at the plasma membrane (23), 3) BILN2061, an NS3 protease inhibitor (28), 4) interferon alpha (IFN-α), a clinical drug against viral replication (41), and 5) naringenin, a blocker of apolipoprotein B-dependent hepatitis C virus secretion (36). These inhibitors were added to JFH1-infected Huh7.5-EG(∆4B5A)SEAP cells at concentrations that did not cause cytotoxicity. Then, the levels of SEAP activity in culture supernatants were measured to reflect anti-HCV activity of the compounds.

In the virus neutralizing test, Huh7.5-EG(∆4B5A)SEAP cells were incubated with the neutralizing anti-CD81 antibody at concentrations of 0, 0.08, 0.4, 2 10, and 50 µg/ml for 1 hour before the addition of JFH1 virus. SEAP activity was measured 6 days later. As shown in Figure 5A, the anti-CD81 Ab showed anti-HCV activity with an EC50 at 0.25 ± 0.03 µg/ml, which was comparable to the results recently shown by Iro et al.
using a similar NS3-based reporter assay (21). Furthermore, the treatment of Huh7.5-EG(Δ4B5A)SEAP cells with another entry inhibitor, H-89, also resulted in the dose-dependent reduction of SEAP activity with an EC$_{50}$ at 4.8 ± 0.2 µM (Fig. 5B), which was similar to the result of Farquhar et al (10).

To clarify whether this reporter system is useful for analyzing inhibitors against HCV replication in the later stages of the viral life cycle, the Huh7-GL cells, a cell line can constitutively produce the infectious virus, were incubated with various concentrations of BILN2061 and IFN-α for 72 hours. Subsequently, the culture supernatant containing secreted JFH1 virions were employed to infect Huh7.5-EG(Δ4B5A)SEAP reporter cells. As revealed by SEAP activity, BILN2061 showed an EC$_{50}$ value of 6.9 ± 0.8 nM (Fig. 5(C)). IFN-α also significantly reduced SEAP activity in a dose-dependent manner (Fig. 5(D)).

To verify if HCV secretion inhibitor could also be discovered using this reporter system, Huh7-GL cells were cultured in the presence of naringenin, with increasing concentrations for 72 hours. Then, the secreted JFH1 viruses were collected to infect Huh7.5-EG(Δ4B5A)SEAP reporter cells for 6 h. At 6 days postinfection, the culture media were assayed for SEAP activity after they had been replaced. As
shown in Figure 5E, naringenin was facilely shown to reduce HCV secretion by 87.2% ± 3.2% at a concentration of 250 µM. There results were similar to those obtained by Nahmias et al. using the rather cumbersome immunofluorescence staining of HCV core protein (36). Taken together, the EC$_{50}$ values are comparable to previous reports, demonstrating that SEAP activity really reflects the activity of the anti-HCV agents of different modes-of-action.

**Statistical evaluation of reporter-based HCV-JFH1 infection assay for robust analysis**

To assess whether these protease-based Huh7.5-EG(Δ4B5A)SEAP reporter cells can be adopted in a high-throughput screening assay for novel anti-HCV agents, assay parameters, including signal to noise ratio (S/N), signal to background ratios (S/B), and Z'-factor, were measured in the presence and absence of H-89 or BILN-2061. As a control for screening of viral entry inhibitors, Huh7.5-EG(Δ4B5A)SEAP cells were treated with 10 µM of H-89 for 1 hour. Six days after JFH1 virus inoculation, the levels of SEAP activity following replacement of media were shown graphically in Figure 6A. The S/N ratio and S/B ratio were 34.1 and 3.9, respectively. Using statistic calculation defined by Zhang’s formula (49), a Z' value of 0.74 was obtained. Similarly, the Huh7.5-EG(Δ4B5A)SEAP cells were evaluated for the screening of
inhibitors working at viral life cycle after the entry and uncoating of vision. Toward this end, Huh7-GL cells were firstly treated with 100 nM BILN2061 for 3 days. Then, the viruses produced in the supernatants were harvested to inoculate Huh7.5-EG(Δ4B5A)SEAP cells. The SEAP activity was measured at 6 days postinfection. As shown in Figure 6B, the S/N and S/B ratios were 21.2 and 3.8, respectively. The Z' factor was 0.64. Collectively, both of different models of virus infection exhibited higher S/B ratios, lower S/N ratios, and more than 0.5 of Z'-factor, which indicated that the signal-to-noise scatter in the assay plates was highly reproducible with little variation for robust determination of inhibitors against viral infection.
Discussion

Currently, the quantification of the levels of HCV RNA or viral proteins relies mainly on real-time RT-qPCR or immunofluorescence assay (IFA). However, these methods are rather time-consuming and labor-intensive. In recent years, several protease-based approaches have been successfully established for developing inhibitors for a number of different diseases including hypertension, diabetes, thrombosis, osteoporosis, and infectious diseases, etc. (7, 14, 35, 45). To discover novel HCV inhibitors, we have previously developed a rapid and reliable assay system in the HCV subgenomic replicon cells based on the principle employed in this study (17-19, 29, 32). The utility of such an assay system has been validated by the fact that we have indeed discovered several bioactive compounds against HCV replication using the reporter system.

The study of HCV in cell culture has been greatly facilitated by the advent of the JFH1 clone (33, 44, 51). In this study, a facile reporter system was established for the evaluation of HCV infectivity in cells. Several engineered reporter fusion proteins were made with several NS3/4A-cleavable peptides encoded in the polyprotein of the JFH1 clone. One important finding was that, out of the three prospective constructs examined, only the EG(Δ4B5A)SEAP reporter was functional
Notably, the EG(Δ4B5A)SEAP reporter described in this study is also a functional reporter for monitoring HCV 1b replication in a sub-genomic replicon cell line (results not shown). Whether the EG(Δ4B5A)SEAP fusion protein can be employed as a reporter for other sub-types of HCV is currently being evaluated in our laboratory.

The use of EG(Δ4B5A)SEAP for HCV replication is advantageous because this reporter does not need to be engineered into the viral genome. Several chimeric JFH1 reporter viruses have been developed for analyzing virus infection (1, 15, 22, 25, 39, 50). One common strategy is to modify HCV cDNA by inserting GFP or luciferase gene into the viral genome in a monocistronic or bicistronic reporter construction. However, genetic modification of a viral genome may be cumbersome. Furthermore, the engineered viral genome rendered more deviation from the authentic RNA structure (27, 39), and the enlarged genome size might slow RNA synthesis and compromise the packaging of viral progeny (9, 20, 25, 27). Schaller et al., also found that expression of the GFP reporter gene rapidly disappeared during prolonged passage of monocistronic HCV reporter virus (39). Accordingly, the higher risk of detrimental mutation associated with the high mutation rate of virus-encoded RNA-dependent RNA polymerase (RdRp) is another caveat to use the monocistronic
HCV reporter genome. In the case of bicistronic reporter genome, Schaller et al. and Koutsoudakis et al. also observed that the kinetics of RNA replication and the release of infectious particles were lower than those of the parental genome (27, 39). The addition of a controlling element such as the internal ribosome entry site (IRES) of encephalomyocarditis virus (EMCV) or the foot-and-mouth disease virus (FMDV) 2A protease also reduces the specificity of anti-HCV drug screening as the prospective hits may interfere these off targets to cause the emergence of false positives (27, 39, 50).

In this report, we also showed that 6 days infection assay is the optimal assay condition for achieving detectable SEAP activity in culture media (Fig. 3). Such a time course of SEAP induction in our assay system is consistent with a very recent study (48). In a protease-based HCV infection assay, Yu et al have demonstrated that the kinetics of HCV NS3/4A protease protein accumulation correlate well with FRET signal detection in Huh7 cell infected with HCV at various MOIs, and greater FRET signal is detectable up to day 6 postinfection, which is similar to our observations (Fig. 3 and 4). However, by use of a similar dual reporter EGFP-SEAP protein linked by an octapeptide DEDEDEDE followed by the HCV genotype 1b NS4A/4B recognition sequence (DEMEEC-ASHL) of NS3/4A protease, Iro et al. defined a significant
increase of SEAP at day 4 postinfection. In addition, the levels of induction of SEAP are greater than our system (21). The reason may be due to the inclusion of the DEDEDEDEDE peptide in Iro’s study. Chou et al. have demonstrated that the DEDEDEDE residues prior to the natural 4A/4B cleavage have markedly enhanced the cleavage efficiency (6). However, we cannot exclude the possibility that there are different copy numbers of chromosomally integrated fusion reporter gene in selected reporter cell clones.

Another interesting observation from this study is that, upon JFH1 infection of Huh7.5-EG(Δ4B5A)SEAP cells, the expression levels of HCV NS3 viral proteins kept on increasing up to day 6 postinfection while the viral RNA level seemed to reach plateau at day 5 (Fig. 3B). This indicates that cellluar anti-HCV immunity may exist in the Huh7.5-based reporter cells to mollify HCV replication during the earlier days postinfection. It will be important to further engineer the host cells so that they can become more permissive to HCV infection; that is, efficient expression of HCV viral proteins would take place immediately upon viral infection. Alternatively, mutant JFH1 clones with various replication competence in host cells may be established to provide a cell culture system that can be more reflective of HCV infection in humans.
Finally, as shown in Table 1, compared to the Ava5-EG(Δ4AB)SEAP system that we have established previously (29), Huh7.5-EG(Δ4B5A)SEAP reporter cells can be facilely utilized for investigating all stages of the HCV life cycle including viral entry, uncoating, replication, virion assembly, and secretion. This novel reporter-based assay system has great potential to be used for screening novel anti-HCV agents. The SEAP activity in culture media may reflect the level of HCV replication, and the viability of cells can be analyzed simultaneously.
Acknowledgments

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Figure legends

Figure 1. Diagram of dual-functional reporter vectors performed in HCV NS3/4A protease activity assay. (A) The various junction regions, NS4A/NS4B, NS4B/NS5A, and NS5A/NS5B, as substrates of HCV NS3/4A protease were located between the egfp gene and the seap gene by an in-frame fusion. The amino acid sequences of the junction regions were Δ4A4B (DEMEECASRA), Δ4B5A (TEDCPIPCSGWLRDV), and Δ5A5B (EDDTVCCSMSYSWTG). Expression of the reporter genes are controlled by human cytomegalovirus (CMV) immediate early promoter and the selection marker for the generation of stable cell line is neomycin phosphotransferase (neo). (B) Huh7.5 cells were seeded at a density of 1 × 10^5 cells per well in 24-well plates and transfected with 0.2 µg of individual plasmids. The expression of EGFP within reporter genes was served as an internal control for the normalization of transfection efficiency. Transiently transfected Huh7.5 cells were infected with or without JFH1 virus at an MOI of 0.02 per well. At 6 days postinfection, the medium was harvested and SEAP activity was measured. Each experiment was performed in triplicate, and error bars reflecting standard deviations are shown. ** P<0.01.

Figure 2. Characterization of Huh7.5-EG(Δ4B5A)SEAP cells stably expressing EG(Δ4B5A)SEAP fusion protein in Huh7.5. (A) Cells were seeded at a density of 1 × 10^5 cells per well in 24-wells and infected with JFH1 virus at an MOI of 0.02 per well.
The lysates of HCV-JFH1-infected and non-infected cells were harvested at 6 days postinfection. Expression of EG(Δ4B5A)SEAP and NS3 protease were analyzed using western blotting with anti-EGFP (upper panel) and anti-NS3 antibody (middle panel), respectively. Lane 1: Huh7.5 cells, lane 2: JFH1-infected Huh7.5 cells, lane 3: Huh-7.5-EG(Δ4B5A)SEAP cells, lane 4: JFH1-infected Huh-7.5-EG(Δ4B5A)SEAP cells. β-actin was the loading control (low panel). (B) SEAP activity in culture medium was measured at 6 days postinfection. Column 1: Huh7.5 cells, Column 2: JFH1-infected Huh7.5 cells, Column 3: Huh7.5-EG(Δ4B5A)SEAP cells, Column 4: JFH1-infected Huh7.5-EG(Δ4B5A)SEAP cells. Each experiment was performed in triplicate, and error bars reflecting standard deviations are shown. ** P<0.01.

**Figure 3.** Kinetics of HCV-JFH1 infection in Huh-7.5-EG(Δ4B5A)SEAP cells. (A) A virus stock generated from Huh7-GL was diluted to infect Huh-7.5-EG(Δ4B5A)SEAP (1 × 10^5 cells per well in 24 well-plates) with various MOIs (0, 0.02, 0.04, 0.09, 0.18, and 0.35). The supernatant medium was collected and SEAP activity was measured at the indicated time points postinfection. The SEAP activity in HCV-JFH1 infection with various MOIs increased in a time-dependent manner. (B) Huh-7.5-EG(Δ4B5A)SEAP cells with HCV-JFH1 inoculation at an MOI of 0.02 were analyzed the intracellular HCV NS3 protein and viral RNA at the indicated time
points postinfection. The HCV NS3 and β-actin protein levels were analyzed by Western blot (upper panel). The intracellular HCV RNA level was quantified by qRT-PCR (lower panel). Each infection experiment was performed in triplicate, and error bars reflecting standard deviations are shown.

**Figure 4.** Correlation between intracellular NS3/4A protease activity and extracellular SEAP activity in HCV-JFH1 infected Huh7.5-EG(Δ4B5A)SEAP cells. In 24-well plates, 1 × 10^5 Huh7.5-EG(Δ4B5A)SEAP cells per well were infected with HCV-JFH1 virus at various MOIs (0, 0.001, 0.003, 0.006, 0.013, and 0.026) for 6 h. Six days postinfection, the cell lysates and supernatant media were harvested to measure NS3/4A protease activity and SEAP activity, respectively. (A) In panel a, linear regression analyzed the relationship between the various numbers of HCV-JFH1 copies and intracellular NS3/4A protease activities or extracellular SEAP activities. In panel b, the cell lysates were analyzed by western blot with anti-NS3 antibody. β-actin is the loading control. (B) Linear regression analysis of SEAP activities and NS3/4A protease activities in HCV-JFH1-infected Huh7.5-EG(Δ4B5A)SEAP cells. Each point was obtained in triplicate, and error bars reflecting standard deviations are shown.
Figure 5. Attenuation of HCV-JFH1 infectivity by reference compounds. (A) and (B) for inhibition of HCV-JFH1 entry, Huh7.5-EG(Δ4B5A)SEAP cells at a density of $8 \times 10^3$ cells per well in 96-well plates were pre-incubated with increasing concentrations of anti-CD81 antibody ($\alpha$CD81) or H-89 for 1 h, and then infected with HCV-JFH1 at an MOI of 0.02 per well in the presence of the inhibitor. SEAP activity and cell viability in each well were measured at 6 days postinfection. The 50% effective concentrations of $\alpha$CD81 and H-89 were 0.34 µg/ml and 2.26 µM, respectively. (C), (D), and (E) for inhibition of HCV-JFH1 intracellular activity, Huh7-GL cells at a density of $1.5 \times 10^4$ cells per well in 96-well plates were treated with BILN2061, IFN-α, and naringenin for 3 days. Following the treatment, the supernatant media containing secreted JFH1 were collected and used to infect Huh7.5-EG(Δ4B5A)SEAP cells. Cell viability was estimated in the meanwhile. Huh7.5-EG(Δ4B5A)SEAP cells were seeded at a density of $8 \times 10^3$ cells per well in 96-well plates and infected with the virus. Six days postinfection, SEAP activity and cell viability were measured. The 50% effective concentrations of BILN2061, IFN-α, and naringenin were 6.98 nM, 21.91 IU/ml, and 8.33 µM, respectively. Each infection experiment was performed in triplicate, and error bars reflecting standard deviations are shown. ** P<0.01.

Figure 6. Estimate of statistical parameters of screening system. (A) HCV-JFH1 entry
system. Huh7.5-EG(Δ4B5A)SEAP cells at a density of $8 \times 10^3$ cells per well in a 96-well plate were utilized to determine the statistical parameters. Cells were pretreated with 10 µM H-89 (open circle), as a background set, and 1% DMSO (closed circle), as a signal set, for 1 h, before being infected with HCV-JFH1 at an MOI of 0.02 per well for 6 h. Six days postinfection, SEAP activity and cell viability were measured. The signal-to-noise ratio (S/N), signal-to-background ratio (S/B), and Z’ factor were 34.1, 3.9, and 0.74, respectively. 

(B) In the HCV-JFH1 secretion system. Huh7-GL cells at a density of $1.5 \times 10^4$ cells per well in a 96-well plate were treated with 100 nM BILN2061 (open circles), as a background set, and 1% DMSO (closed circles), as a signal set, for 3 days. Subsequently, supernatant media were inoculated in Huh7.5-EG(Δ4B5A)SEAP cells ($8 \times 10^3$ cells per well) for 6 h in a plate to plate manner. Six days postinfection, SEAP activity and cell viability were measured. S/N, S/B, and the Z’ factor were 21.2, 3.8, and 0.64, respectively.

Table1. Comparison of reporter-based replicon system and HCV-JFH1 infectious system in drug discovery.

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<td>Replicon system was applied as described by J.-C. Lee et al. (2004) (29).</td>
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aCD81 indicates anti-CD81 antibody.

bNot Applicable.

cApplicable.
References


11. **Gallinari, P., D. Brennan, C. Nardi, M. Brunetti, L. Tomei, C. Steinkuhler,


44. Wakita, T., T. Pietschmann, T. Kato, T. Date, M. Miyamoto, Z. Zhao, K.


Pan et al., Figure 1A

A

pUC ori $P_{CMV\text{IE}}$ EGFP SEAP $SV_{40}$ poly A $P_{SV_{40}E}$ $Kan^{r}/Neo^{r}$ $HSV\ TK$ poly A

△4A4B: (DEMEEC-ASRA)
△4B5A: (TEDCPIPC-SGSWLRDV)
△5A5B: (EDDTVCC-SMSYSWTG)
Pan et al., Figure 1B

Fold increase SEAP activity
0
1
2
3
4
5
6
Huh7.5/Mock
Huh7.5/JFH1
**
pEG(cleavage site)SEAP
∆5A5B ∆4B5A ∆4A4B

B

Fold increase SEAP activity

Huh7.5/Mock
Huh7.5/JFH1

∆4A4B ∆4B5A ∆5A5B

pEG(cleavage site)SEAP

**
A

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<td>+</td>
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Pan et al., Figure 2A
Pan et al., Figure 3

Days post infection (MOI: 0.02)

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<td>Mock</td>
<td>NS3 protease</td>
<td>β-actin</td>
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Intracellular HCV RNA (copies/ug total RNA)

Pan et al., Figure 3
Pan et al., Figure 4

A

a. NS3 protease activity

b. MOI 0 0.001 0.003 0.007 0.013 0.026

B

Fold increase NS3 protease activity

R² = 0.962

MOI

Fold increase SEAP activity

0 0.000 0.005 0.010 0.015 0.020 0.025 0.030

 folded increase NS3/4A Protease activity

0 2 4 6 8 10 12 14

NS3 protease activity

SEAP activity

β-actin

Pan et al., Figure 4

MOI 0 0.001 0.003 0.007 0.013 0.026

NS3 protease

β-actin
Pan et al., Figure 6

A

S/N = 34.1
S/B = 3.9
Z' = 0.74

B

S/N = 21.2
S/B = 3.8
Z' = 0.64
### Drug Screening Systems

<table>
<thead>
<tr>
<th>Antiviral targets</th>
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<th>Replicon system</th>
<th>HCV-JFH1 Virus system</th>
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<td>Entry</td>
<td>αCD8, H-89</td>
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