

In Vitro and In Vivo Antibacterial Activities of OPC-20011, a Novel Parenteral Broad-Spectrum 2-Oxaisocephem Antibiotic

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OPC-20011, a new parenteral 2-oxaisocephem antibiotic, has an oxygen atom at the 2- position of the cephalosporin frame. OPC-20011 had the best antibacterial activities against gram-positive bacteria, including methicillin-resistant *Staphylococcus aureus* (MRSA), *Enterococcus faecalis*, and penicillin-resistant *Streptococcus pneumoniae*: MICs at which 90% of the isolates were inhibited were 6.25, 6.25, and 0.05 µg/ml, respectively. Its activity is due to a high affinity of the penicillin-binding protein 2' in MRSA, an affinity which was approximately 1,050 times as high as that for flomoxef. Against gram-negative bacteria, OPC-20011 also showed antibacterial activities similar to those of ceftazidime. The in vivo activities of OPC-20011 were comparable to or greater than those of reference compounds in murine models of systemic infection caused by gram-positive and -negative pathogens. OPC-20011 was up to 10 times as effective as vancomycin against MRSA infections in mice. This better in vivo efficacy is probably due to the bactericidal activity of OPC-20011, while vancomycin showed bacteriostatic activity against MRSA. OPC-20011 produced a significant decrease of viable counts in lung tissue at a dose of 2.5 mg/kg of body weight, an efficacy similar to that of ampicillin at a dose of 10 to 20 mg/kg on an experimental murine model of respiratory tract infection caused by non-ampicillin-susceptible *S. pneumoniae* T-0005. The better therapeutic efficacy of OPC-20011 was considered to be due to its potent antibacterial activity and low affinity for serum proteins of experimental animals (29% in mice and 6.4% in rats).

β-Lactam antibiotics are known to have strong, broad-spectrum antibacterial activity against both gram-positive and -negative bacteria and to be safe in clinical use. There is, however, a worldwide increase in bacteria highly resistant to β-lactams (4, 8, 22, 23). Approximately 50 to 80% of *Staphylococcus aureus* strains have become resistant to many kinds of antibiotics, with a few exceptions, such as vancomycin (VCM). Furthermore, resistance to penicillin among strains of *Streptococcus pneumoniae* has been spreading worldwide (9, 14, 15). In some regions, such as Spain, almost 50% of *S. pneumoniae* clinical isolates are penicillin resistant. These facts suggest that there may be few antibiotics which can be utilized against infections caused by these gram-positive pathogens in the future. Furthermore, the introduction of new antibiotics has been decreasing recently compared with the period from 1945 to 1970, when a variety of new antibiotics with new structures were discovered (3). The time has come to attempt to develop new agents which can solve the problem of resistant bacteria. Our efforts have been focused on the development of new antibiotics to solve these problems. In earlier work (6, 7), 2-oxaisocephem antibiotics were synthesized and evaluated as a new class of antibiotics by Doyle et al. They reported that 2-oxaisocephem lacked comprehensive antibacterial activity (7). In the course of our research, we identified OPC-20011 from a novel series of 2-oxaisocephems that showed good antibacterial activity against methicillin-resistant *S. aureus* (MRSA), *Enterococcus faecalis*, and penicillin-resistant *S. pneumoniae* and had a broad spectrum of antibacterial activity (13, 24). In this study we evaluated the antibacterial activities in vitro and in vivo of OPC-20011 and compared them with those of reference compounds. We also compared OPC-20011 with a 1-

sulfur analog with the same substituents as OPC-20011 to clarify the properties of 2-oxaisocephem antibiotics.

MATERIALS AND METHODS

Organisms. The clinical isolates used in this study were obtained from several hospitals in Japan from 1990 to 1995 and were stocked in our laboratory. The other strains used in this study were stock cultures from our laboratory.

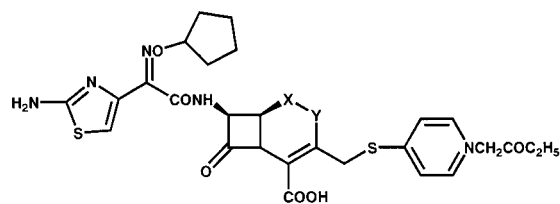
Antimicrobial agents. OPC-20011 and OPC-20026, a 1-sulfur analog with the same substituents as OPC-20011 (Fig. 1), were synthesized at the Microbiological Research Institute of Otsuka Pharmaceutical Co., Ltd., Tokushima, Japan. The other antibiotics, including cefotiam (CTM), flomoxef (FMOX), ceftazidime (CAZ), cefpirome (CPR), ampicillin (ABPC), imipenem (IPM), methicillin (DMPPC), and VCM, were purchased from commercial sources.

Susceptibility testing. MICs were determined by the twofold agar dilution method with Mueller-Hinton agar (Difco Laboratories, Detroit, Mich.), which was supplemented with 5% horse blood to support the growth of *S. pneumoniae*, *Streptococcus pyogenes*, and *Haemophilus influenzae*. The overnight cultures were diluted to approximately 10⁶ CFU/ml with fresh broth, and an inoculum of 10⁴ CFU per spot was applied with an inoculating apparatus (Microplanter; Sakuma Seisakusho, Tokyo, Japan) to agar plates containing graded concentrations of each compound. The MICs were defined as the minimum drug concentrations which completely inhibited the growth of bacteria after incubation at 37°C for 18 h.

Affinities of PBPs. The affinities of penicillin-binding proteins (PBPs) for OPC-20011 and the reference compounds were determined by competition assay with [¹⁴C]benzylpenicillin (Amersham Japan Co., Ltd., Tokyo, Japan) as described previously (20, 21). Membranes were collected by differential centrifugations (5,000 × g for 15 min and 100,000 × g for 60 min) of enzymatically (100 µg of lysostaphin/ml containing 10 µg of DNase/ml) disrupted cells of *S. aureus* FDA 209-P and MRSA 5143 or sonically disrupted cells of *S. pneumoniae* T0096 and *Escherichia coli* NIHJ JC-2 in 50 mM potassium phosphate buffer (pH 7.0). The concentrations of membrane proteins, based on γ-globulin levels, were adjusted to 8 mg/ml for *S. aureus* FDA 209-P, MRSA 5143, and *S. pneumoniae* T0096 and to 20 mg/ml for *E. coli* NIHJ JC-2. The binding reactions were done for 10 min with test compounds at each concentration followed by 15 min with [¹⁴C]benzylpenicillin at 37°C. The binding affinities of PBPs for the test compounds were expressed as the concentration required to prevent 50% of the binding of [¹⁴C]benzylpenicillin (50% inhibitory concentration [IC₅₀]) to each PBP, determined by the imaging analyzer BAS 2000 (Fuji Chemical Co. Ltd.).

In vivo effects. (i) **Systemic infections.** In vivo activities against systemic infections caused by gram-positive and -negative pathogens were determined. Ten male ICR mice weighing 20 ± 1 g were used for each dosage group. The mice were challenged intraperitoneally with 0.5 ml of approximately 10 to 100 times the 50% lethal doses of the respective pathogens. The bacterial suspensions,

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Compounds	X	Y
OPC-20011	CH ₂	O
OPC-20026	S	CH ₂

FIG. 1. Chemical structures of OPC-20011 and OPC-20026.

which were prepared from overnight cultures on Tryptic soy broth (Difco) for methicillin-susceptible *S. aureus* (MSSA) and MRSA, on brain heart infusion broth (Nissui) for *S. pneumoniae*, and on Mueller-Hinton broth for other pathogens at 37°C, were suspended in the fresh broth of overnight cultures containing gastric mucin (Difco) (for the concentrations, see Table 6). One hour after infection, various doses of each compound were administered to the mice subcutaneously. The number of mice surviving after each dose was determined on day 7 after infection, and the 50% effective dose (ED₅₀) was calculated by the probit method.

(ii) **Respiratory tract infection.** Groups of five rats (SD rats; age, 5 weeks), pretreated 3 days before infection with cyclophosphamide, were anesthetized with diethyl ether and challenged directly in the respiratory tract by instilling 0.2 ml of a suspension of MRSA 5143. The challenge doses were 2.75×10^7 CFU/ml. Antibiotics were administered subcutaneously at doses of 2.5, 5, and 10 mg/kg of body weight for OPC-20011 or 5, 10, and 20 mg/kg for VCM three times, at 24, 36, and 48 h following infection. The lungs were then removed aseptically under diethyl ether anesthesia 24 h after the final treatment and homogenized in 2 ml of distilled water, and the viable bacilli in the lungs of each rat were counted on Mueller-Hinton agar plates.

Groups of five mice (ICR mice; age, 4 weeks) were anesthetized with diethyl ether and challenged directly in the respiratory tract by instilling 0.1 ml of suspension of *S. pneumoniae* T0005, which is a pathogen insensitive to ABPC. The challenge doses were 2.75×10^4 CFU/ml. Antibiotics were administered subcutaneously at doses of 2.5, 5, and 10 mg/kg once at 24 h for OPC-20011 or at doses of 5, 10, and 20 mg/kg three times, at 24, 40, and 48 h after infection, for ABPC and VCM. The lungs were then removed aseptically under diethyl ether anesthesia 24 h after the final treatment and homogenized in 2 ml of distilled water, and the viable bacilli in the lungs of each mouse were counted on Mueller-Hinton agar plates which were supplemented with 5% horse blood. Statistical analysis was done by the following procedures. The viable count of bacilli in the lungs of each animal was expressed as log₁₀ CFU, and the differences among the groups was determined by using a two-tailed Dunnett's test.

Time-kill curve. The activities of OPC-20011 and VCM against MRSA were determined by a time-kill study, as described previously (27). An overnight culture of MRSA was diluted with fresh Mueller-Hinton broth to a density of approximately 10^4 CFU/ml, and the dilution was preincubated in L-tubes at 37°C for 1 h with shaking. Various concentrations, close to the MICs, of the test compounds were added to the culture. At fixed time intervals, 0.1-ml portions of the contents of each tube were removed to 0.9 ml of fresh saline and spread on drug-free Mueller-Hinton agar plates after being suitably diluted (10^1 to 10^7 times). The colonies were counted after incubation for 18 h at 37°C.

Affinity for serum proteins of different animal species. The binding percentages of OPC-20011 to serum proteins of different animal species were determined by the following procedure. Sera were obtained from different species (ICR mice, SD rats, NEZ rabbits, beagle dogs, cynomolgus monkeys, and healthy human donors). Each serum was mixed with 1/10 volume of 500 µg of OPC-20011/ml and incubated for 1 h at 37°C. The incubated samples were then ultrafiltered with Centriscart Cutoff 20000 (Sartorius). The binding concentrations of the compound in the filtrates were measured by high-performance liquid chromatography (SPD-10A chromatograph [Shimadzu Co. Ltd.]; UV wavelength, 300 nm; solvent, 25% solution of CH₃CN containing 0.05% trifluoroacetate).

RESULTS

Susceptibility of clinical isolates. The activity of OPC-20011 against clinical isolates was compared with that of OPC-20026,

CTM, CAZ, FMOX, CPR, ABPC, IPM, DMPPC, and VCM. The MICs at which 90% of the isolates are inhibited (MIC₉₀s) of test compounds for the strains tested are shown in Tables 1 and 2. In the first study, the MIC₉₀s of OPC-20011 were 2 to 16 times lower than those of OPC-20026. In particular, a 16-fold difference was observed between the MIC₉₀ of OPC-20011 and that of OPC-20026 for *E. faecalis* (Table 1). The MIC₉₀s of OPC-20011 for MSSA, DMPPC-susceptible *Staphylococcus epidermidis*, and non-ABPC-susceptible *S. pneumoniae* were 0.78, 1.56, and 0.05 µg/ml, respectively, the lowest MICs among the test compounds. For MRSA, the MIC₉₀ of OPC-20011 was 6.25 µg/ml, which was lower than those of other β-lactam antibiotics tested but 2 dilutions higher than that of VCM. The MIC₉₀s of OPC-20011 for DMPPC-resistant *S. epidermidis* and *E. faecalis* were 3.13 and 6.25 µg/ml, respectively, which were almost equal to those for VCM and ABPC, respectively, and 2 to 32 times lower than those of the other reference antibiotics tested.

Against *E. coli* and *Klebsiella pneumoniae*, OPC-20011 inhibited 90% of the strains at 1.56 and 3.13 µg/ml, respectively, which were 2 to 32 times higher than the MIC₉₀s of the reference compounds. Against *H. influenzae*, OPC-20011 inhibited 90% of the strains at 0.2 µg/ml, which was equal to or two to eight times lower than the MIC₉₀s of the reference compounds. Against *Serratia marcescens*, *Proteus mirabilis*, and *Providencia rettgeri*, OPC-20011 inhibited 90% of the strains at 6.25, 1.56, and 1.56 µg/ml, respectively, at least 64 times more active than CTM and FOMX. Weaker activity was observed with all of the test compounds against *Pseudomonas aeruginosa*: the MIC₉₀s were 50 to 100 µg/ml.

PBP affinity. In order to explain the antibacterial activities of OPC-20011, the binding of OPC-20011 to PBPs in *S. aureus* FDA 209-P, MRSA 5143, *S. pneumoniae* C0096, and *E. coli* NIHJ JC-2 was compared with those of the reference compounds. The results are shown in Tables 3 to 5. The IC₅₀s of OPC-20011 for PBP2, -3, and -4 in *S. aureus* FDA 209-P were generally 3 to >64 times lower than those of CTM, CAZ, FMOX, and CPR. For PBP2' in MRSA 5143, the IC₅₀ of OPC-20011 was 2.0 µg/ml, which was approximately 370 and 1,050 times as high as those of CPR and FMOX, respectively. The IC₅₀s of OPC-20011 for PBP1A and -1B in *S. pneumoniae* C0096 were 9 to 50 times lower than those of ABPC. PBP1B, -2 and -3 in *E. coli* NIHJ JC-2 showed the highest affinity for OPC-20011 compared with FMOX and CPR, but OPC-20011

TABLE 1. Comparison of antibacterial activities (MIC₉₀s) of OPC-20011 and OPC-20026

Clinical isolate	No. of strains	MIC ₉₀ (µg/ml)		Ratio ^a
		OPC-20011	OPC-20026	
MSSA	47	0.78	1.56	2
MRSA	33	6.25	25	4
DMPPC-susceptible <i>S. epidermidis</i>	44	1.56	3.13	2
DMPPC-resistant <i>S. epidermidis</i>	34	3.13	6.25	2
<i>E. faecalis</i>	25	6.25	100	16
Non-ABPC-susceptible <i>S. pneumoniae</i>	18	0.05	0.1	2
<i>E. coli</i>	54	1.56	6.25	4
<i>K. pneumoniae</i>	26	3.13	12.5	4
<i>S. marcescens</i>	27	6.25	12.5	2
<i>P. mirabilis</i>	41	1.56	6.25	4
<i>P. rettgeri</i>	40	1.56	6.25	4

^a MIC₉₀ of OPC-20026/MIC₉₀ of OPC-20011.

TABLE 2. Antibacterial activities of OPC-20011 and reference compounds against clinical isolates

Organism (no. of strains)	Compound	MIC (µg/ml)			
		Range	MIC ₅₀	MIC ₉₀	
MSSA (47)	OPC-20011	0.1–3.13	0.39	0.78	
	CTM	0.025–12.5	0.78	3.13	
	FMOX	0.39–6.25	0.78	1.56	
	CAZ	3.13–50	12.5	25	
	CPR	0.39–25	1.56	1.56	
	IPM	0.006–1.56	0.006	0.1	
	DMPPC	1.56–6.25	3.13	6.25	
	VCM	0.78–1.56	0.78	1.56	
MRSA (33)	OPC-20011	0.78–6.25	6.25	6.25	
	CTM	100–>100	100	100	
	FMOX	0.78–100	25	100	
	CAZ	>100–>100	100	100	
	CPR	3.13–100	50	100	
	IPM	0.1–>100	50	100	
	DMPPC	>100–>100	100	100	
	VCM	0.78–1.56	1.56	1.56	
DMPPC-susceptible <i>S. epidermidis</i> (44)	OPC-20011	0.2–1.56	0.39	1.56	
	CTM	0.39–3.13	0.78–3.13		
	FMOX	0.39–12.5	1.56	12.5	
	CAZ	3.13–25	6.25	25	
	CPR	0.39–6.25	0.78	3.13	
	DMPPC	1.56–6.25	3.13	6.25	
	VCM	0.78–3.13	1.56	3.13	
	DMPPC-resistant <i>S. epidermidis</i> (34)	OPC-20011	0.78–6.25	1.56	3.13
CTM		1.56–12.5	3.13	6.25	
FMOX		3.13–100	25	50	
CAZ		12.5–>100	25	100	
CPR		3.13–50	6.25	50	
DMPPC		12.5–>100	100	100	
VCM		1.56–3.13	1.56	3.13	
<i>E. faecalis</i> (27)		OPC-20011	0.78–6.25	3.13	6.25
	CTM	100–>100	100	100	
	FMOX	100–>100	100	100	
	CAZ	>100	100	100	
	CPR	12.5–100	50	100	
	ABPC	1.56–3.13	1.56	3.13	
	ABPC-susceptible <i>S. pneumoniae</i> (27)	OPC-20011	<0.006–	<0.006	<0.006
		CTM	0.025–0.78	0.2	0.78
FMOX		0.025–0.39	0.2	0.39	
CAZ		0.0125–6.25	0.2	0.39	
CPR		<0.006–0.78	0.0125	0.1	
ABPC		<0.006–0.1	0.025	0.1	
VCM		0.05–0.2	0.1	0.2	
Non-ABPC-susceptible <i>S. pneumoniae</i> (18)		OPC-20011	<0.006–0.05	0.025	0.05
	CTM	3.13–6.25	6.25	6.25	
	FMOX	1.56–6.25	3.13	6.25	
	CAZ	1.56–6.25	3.13	6.25	
	CPR	0.2–0.78	0.39	0.78	
	ABPC	1.56–3.13	3.13	3.13	
	VCM	0.05–0.2	0.1	0.2	
	<i>E. coli</i> (54)	OPC-20011	0.39–1.56	0.78	1.56
CTM		0.1–0.78	0.2	0.78	
FMOX		0.05–0.39	0.1	0.2	
CAZ		0.1–0.78	0.2	0.78	
CPR		0.025–0.2	0.05	0.2	
<i>K. pneumoniae</i> (26)		OPC-20011	0.1–6.25	1.56	3.13
	CTM	0.1–1.56	0.2	0.39	
	FMOX	0.1–0.78	0.1	0.1	
	CAZ	0.1–25	0.2	0.39	
	CPR	0.05–3.13	0.1	0.1	
	<i>H. influenzae</i> (20)	OPC-20011	0.025–0.2	0.1	0.2
CTM		0.1–0.78	0.2	0.39	
FMOX		0.78–3.13	0.78	1.56	
CAZ		0.05–0.39	0.2	0.2	
CPR		0.05–0.39	0.1	0.2	

Continued

TABLE 2—Continued

Organism (no. of strains)	Compound	MIC (µg/ml)		
		Range	MIC ₅₀	MIC ₉₀
<i>S. marcescens</i> (27)	OPC-20011	1.56–12.5	1.56	6.25
	CTM	3.13–>100	12.5	100
	FMOX	0.78–50	0.78	6.25
	CAZ	0.2–25	0.39	0.78
	CPR	0.05–0.78	0.1	0.39
<i>P. mirabilis</i> (41)	OPC-20011	0.05–>100	0.39	1.56
	CTM	0.2–>100	0.78	100
	FMOX	0.2–>100	0.39	100
	CAZ	0.05–100	0.2	6.25
	CPR	<0.006–>100	0.1	1.56
<i>P. rettgeri</i> (40)	OPC-20011	0.39–3.13	0.78	1.56
	CTM	0.05–>100	0.39	100
	FMOX	0.05–6.25	0.1	3.13
	CAZ	0.05–6.25	0.2	1.56
	CPR	<0.006–0.39	0.025	0.2
	<i>P. aeruginosa</i> (27)	OPC-20011	12.5–>100	25
CTM		>100	100	100
FMOX		>100	100	100
CAZ		1.56–50	3.13	50
CPR		3.13–100	12.5	50

did not bind to PBP1A, -4, and -5 at concentrations as high as 62.5 µg/ml.

Protective effects in vivo. The therapeutic effects of OPC-20011 and the reference compounds against acute systemic infections in mice are shown in Table 6. The efficacies of OPC-20011 against *S. aureus* Smith, MRSA 5120, *S. pneumoniae* Type III, and *S. pneumoniae* C0096 were approximately 2 to over 100 times greater than those of the other compounds tested. OPC-20011 showed protective effects equivalent to those of VCM against MRSA 5143 and to those of ABPC against *E. faecalis*, and it was approximately 8 to over 40 times more effective than the other reference compounds. OPC-20011 also had ED₅₀s ranging from 0.06 to 8.4 mg/kg for the infections caused by gram-negative pathogens, except *P. aeruginosa*. Against *P. aeruginosa* T0017, the efficacy of OPC-20011 was nearly equal to that of aztreonam (AZT) but less than that of CAZ, though the other β-lactams, CTM and FMOX, did not exhibit any protective effects.

The therapeutic effects of OPC-20011 and VCM against experimental respiratory tract infection caused by MRSA 5143 are shown in Table 7. A significant decrease in the number of viable cells in the lungs was observed at concentrations of ≥5 mg/kg for OPC-20011 and ≥10 mg/kg for VCM. The efficacy of

TABLE 3. Binding of OPC-20011 and reference compounds to PBPs in *S. aureus* FDA 209-P

Compound	IC ₅₀ (µg/ml) ^a			
	PBP1	PBP2	PBP3	PBP4
OPC-20011	ND ^b	0.068	0.053	0.065
OPC-20026	ND	0.303	0.501	0.199
CTM	10.946	0.273	0.282	0.283
CAZ	ND	0.360	>3.13	0.327
FMOX	32.545	0.212	0.370	1.027
CPR	ND	0.163	3.086	0.022

^a Binding affinities of test compounds for PBPs are expressed as the concentration (in micrograms per milliliter) required to prevent 50% of the binding of [¹⁴C]benzylpenicillin to each PBP.

^b ND, not detected.

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TABLE 4. Binding of OPC-20011 and reference compounds to PBPs in *S. pneumoniae* C0096

Compound	IC50 ($\mu\text{g/ml}$) ^a				
	PBP1A	PBP1B	PBP2X	PBP2	PBP3
OPC-20011	0.14	0.08	1.28	>6.4	<0.025
ABPC	7.16	0.70	0.65	>6.4	<0.025

^a Binding affinities of test compounds for PBPs are expressed as the concentration (in micrograms per milliliter) required to prevent 50% of the binding of [¹⁴C]benzylpenicillin to each PBP.

OPC-20011 at 5 mg/kg was almost equal to that of VCM at 20 mg/kg.

The therapeutic effects of OPC-20011, ABPC, and VCM against experimental respiratory tract infection caused by *S. pneumoniae* T0005 are shown in Table 8. A significant decrease in the number of viable cells in the lungs was observed at concentrations of ≥ 2.5 mg/kg for OPC-20011 and 20 mg/kg for ABPC with a single treatment. The efficacy of OPC-20011 at 2.5 mg/kg was almost equal to that of ABPC at 10 to 20 mg/kg. When three doses were given, no viable bacilli were observed in any of the OPC-20011-treated groups. With VCM, a significant decrease was observed when three doses were given, though no decrease was observed with a single treatment.

Bactericidal activity. The bactericidal activity of OPC-20011 was compared with that of VCM against MRSA 5143 and MRSA 5120 (Fig. 2 and 3). Against MRSA 5143, both OPC-20011 (at twice the MIC) and VCM (at four times the MIC) showed bactericidal activity. Against MRSA 5120, however, OPC-20011 showed bactericidal activity at one-fourth the MIC within 8 h and produced a rapid decrease in the number of viable cells at the same concentration, while VCM did not show any bactericidal activity even at a concentration of eight times the MIC. The MICs against the obvious regrowth of MRSA 5120 exposed to concentrations of OPC-20011 below the MIC for 24 h were not changed.

Affinity for serum proteins of different animal species. The affinities of the filters (Centrisart Cutoff 20000) used in this binding study for OPC-20011 were checked. The rate of binding of OPC-20011 to the filters was less than 1%. The binding percentage (\pm standard deviation [SD]) of OPC-20011 to serum proteins prepared from ICR mice, SD rats, NEZ rabbits, beagle dogs, cynomolgus monkeys, and healthy human donors are shown in Table 9. The rates of binding of OPC-20011 to serum proteins from humans and mice were almost equal.

TABLE 5. Binding of OPC-20011 and reference compounds to PBPs in *E. coli* NIHJ JC-2

Compound	IC50 ($\mu\text{g/ml}$) ^a						
	PBP1A	PBP1B	PBP2	PBP3	PBP4	PBP5	PBP6
OPC-20011	>62.5	2.56	<0.1	<0.1	>62.5	>62.5	18.84
OPC-20026	1.52	1.57	1.74	<0.1	>62.5	>62.5	>62.5
FMOX	0.10	1.26	>62.5	<0.1	1.22	0.14	0.14
CPR	10.13	2.78	4.04	<0.1	38.43	>62.5	>62.5

^a Binding affinities of test compounds for PBPs were expressed as the concentration (in micrograms per milliliter) required to prevent 50% of the binding of [¹⁴C]benzylpenicillin to each PBP.

TABLE 6. Protective effects of OPC-20011 and reference compounds against experimental systemic infection models in mice

Organism (% of mucin used)	Compound	MIC ($\mu\text{g/ml}$)	Challenge dose (cells/mouse)	ED ₅₀ (mg/kg)	Confidence limit (mg/kg)
<i>S. aureus</i> Smith (5)	OPC-20011	0.2	1.32×10^7	0.22	0.11–0.37
	CTM	0.39		0.61	0.006–1.56
	CAZ	12.5		12.12	6.54–19.66
	FMOX	0.39		0.53	0.17–0.87
	CPR	0.39		1.72	1.29–2.40
MRSA 5120 (5)	OPC-20011	6.25	7.6×10^7	3.74	2.58–5.12
	CTM	100		1.00	0.60–1.57
	CAZ	100		>100	>100
	FMOX	25		8.52	4.41–17.3
	CPR	50		27.7	12.92–105
MRSA 5143 (5)	OPC-20011	6.25	1.31×10^7	11.5	7.12–17.2
	CTM	>100		6.63	4.48–10.8
	CAZ	>100		>200	>200
	FMOX	50		74.5	37.9–230
	CPR	100		>200	>200
<i>E. faecalis</i> C0063 (2.5)	OPC-20011	0.78	7.0×10^7	4.66	3.30–6.36
	CTM	100		>200	>200
	CAZ	>100		>200	>200
	FMOX	100		>200	>200
	CPR	12.5		>100	>100
<i>S. pneumoniae</i> Type III (none)	ABPC	1.56	4.2×10^2	5.2	3.86–7.00
	OPC-20011	<0.0015		5.54	3.56–11.9
	CTM	0.1		27	17.94–56.3
	CAZ	0.2		172	122–341
	FMOX	0.2		11.2	5.26–41.4
<i>S. pneumoniae</i> C0096 (none)	CPR	0.013	2.0×10^4	26.72	19.5–42.1
	ABPC	0.025		13.2	8.36–25.3
	OPC-20011	0.006		0.56	0.31–0.90
	CTM	6.25		29.5	17.4–49.5
	CAZ	3.13		18.1	11.8–33.5
<i>E. coli</i> 29 (2.5)	FMOX	6.25	1.35×10^6	18.6	10.1–63.7
	CPR	0.2		1.68	1.08–2.55
	ABPC	3.13		8.23	5.47–12.8
	OPC-20011	0.39		0.15	0.12–0.19
	CTM	0.10		1.10	0.78–1.53
<i>E. coli</i> T0020 (2.5)	CAZ	0.39	6.8×10^5	0.17	0.12–0.22
	FMOX	0.10		0.43	0.31–0.57
	CPR	0.025		0.64	0.46–0.86
	OPC-20011	0.013		0.06	0.039–0.090
	CTM	0.05		0.052	0.036–0.072
<i>K. pneumoniae</i> T0011 (2.5)	CAZ	0.05	5.3×10^2	0.21	0.126–0.298
	FMOX	0.025		0.59	0.435–0.775
	CPR	0.013		0.65	0.486–0.871
	OPC-20011	0.2		8.42	5.93–12.4
	CTM	0.1		10.7	8.14–14.3
<i>S. marcescens</i> T0004 (2.5)	CAZ	0.05	1.6×10^6	0.87	0.24–1.45
	FMOX	0.1		12.7	9.80–16.4
	CPR	0.013		0.67	0.45–0.91
	OPC-20011	0.78		1.41	0.622–2.30
	CTM	3.13		26.7	8.34–56.2
<i>P. mirabilis</i> T0003 (2.5)	CAZ	0.2	2.5×10^6	0.14	0.065–0.300
	FMOX	0.78		2.34	1.20–3.79
	CPR	0.1		0.11	0.0063–0.209
	OPC-20011	0.2		3.38	1.51–6.59
	CTM	0.39		10.6	5.73–26.6
<i>P. aeruginosa</i> T0017 (2.5)	CAZ	0.05	4.8×10^4	1.42	8.92–59.3
	FMOX	0.39		22.6	0.686–2.53
	OPC-20011	6.25		130	95.7–171
	CTM	>100		>200	>200
	CAZ	0.78		11.3	5.44–22.9
	FMOX	>100	>200	>200	>200
	AZT	3.13		78.6	50.3–133

TABLE 7. Therapeutic effects of OPC-20011 and VCM in an experimental murine model of respiratory tract infection caused by MRSA 5143^a

Compound	Dose (mg/kg)	Log viable bacilli in lungs ^b (± SD)
Control	0	7.81 ± 0.75
OPC-20011	2.5	5.92 ± 1.23
	5	3.58 ± 0.47**
	10	2.21 ± 0.76**
VCM	5	7.14 ± 1.32
	10	5.35 ± 1.47*
	20	4.48 ± 0.51**

^a Challenge dose was 5.5×10^6 CFU/mouse.
^b Viable bacilli in lungs were counted at 24 h after final treatment. The difference between the control and each treated group was determined by using a two-tailed Dunnett's test (*, $P < 0.05$; **, $P < 0.01$).

DISCUSSION

OPC-20011 is a 2-oxaisocephem antibiotic that has never been introduced to clinical use. In a previous report about 2-oxaisocephems, it was shown to have limited antibacterial activity (7). Then what is the advantage of a 2-oxaisocephem? The answers were obtained in this study. The antibacterial activities and binding to PBPs of OPC-20011 and OPC-20026, a 1-sulfur analog with the same substituents as OPC-20011, were compared. The antibacterial activity of the 2-oxaisocephem was 2 to 16 times more potent than that of the 1-sulfur cephem. This potency of the 2-oxaisocephem was considered to be due to the high affinity of the PBP in *S. aureus*, including MRSA.

None of the commercially available cephem antibiotics that were tested had any activity against *E. faecalis*; OPC-20011, the oxaisocephem antibiotic, showed antibacterial activity similar to that of ABPC, although the 1-sulfur analog with the same substituents as OPC-20011 was not active against this organism.

MRSA has an additional PBP2', which is involved in resistance to β-lactams (26). There have also been many recent reports about other factors related to DMPPC resistance (2, 5, 10, 11, 17). The mechanism of resistance in MRSA is more complex than PBP2' alone. Some other factors, such as *femA*, the glycine content of peptidoglycan, and the enhancement of autolysis, have been reported to be essential for expression of

TABLE 8. Therapeutic effects of OPC-20011, ABPC, and VCM in an experimental murine model of respiratory tract infection caused by *S. pneumoniae* T0005^a

Compound ^c	Dose (mg/kg)	Log viable bacilli in lungs ^b (± SD)	
		Single	Three times
Control	0	6.42 ± 0.82	7.21 ± 0.5
OPC-20011	2.5	4.0 ± 1.5*	<1**
	5	3.08 ± 1.18*	<1**
	10	1.73 ± 0.89**	<1**
	20	0.84 ± 1.03**	<1**
ABPC	2.5	6.02 ± 1.08	6.23 ± 1.51
	5	5.55 ± 1.52	3.67 ± 0.73**
	10	3.76 ± 2.28	2.09 ± 0.96**
	20	2.98 ± 1.29*	1.73 ± 0.95**
VCM	5	5.09 ± 1.19	4.11 ± 2.1*
	10	5.43 ± 1.81	<1**
	20	4.40 ± 1.2	<1**
	40	3.72 ± 2.09	<1**

^a Challenge dose was 2.75×10^3 CFU/mouse.
^b Viable bacilli in lungs were counted at 24 h after final treatment. Single, single treatment at 24 h after infection; three times, treatments at 24, 40, and 48 h after infection. The difference between the control and each treated group was determined by using a two-tailed Dunnett's test (*, $P < 0.05$; **, $P < 0.01$).
^c MICs of OPC-20011, ABPC, and VCM against *S. pneumoniae* T0005 were 0.003, 0.2, and 0.39 μg/ml, respectively.

DMPPC resistance. However, PBP2' is an essential factor for DMPPC resistance (16, 19, 25), and inhibiting the activity or the expression of PBP2' is the best way to solve the resistance problem. PBP2' bound OPC-20011 approximately 370 to 1,050 times better than the other β-lactams. This may be the reason for the MIC for OPC-20011 against all of the clinically isolated MRSAs, which was less than 6.25 μg/ml.

Although VCM is probably the best drug that can be used for MRSA infections at present, VCM-intermediate MRSA has been isolated in Japan (12). Though the in vitro anti-MRSA activity of OPC-20011 was less than that of VCM, the in vivo efficacy of OPC-20011 in MRSA infection models was almost equal to or greater than that of VCM. In particular, OPC-20011 showed a therapeutic effect two to four times as potent as that of VCM in the model of respiratory tract infection, even though OPC-20011 showed activity almost equal to that of VCM in systemic infection in mice caused by the same organism, MRSA 5143. One reason may be related to the bactericidal activities of these compounds for MRSA. The bactericidal activity of OPC-20011 for MRSA 5120 was greater

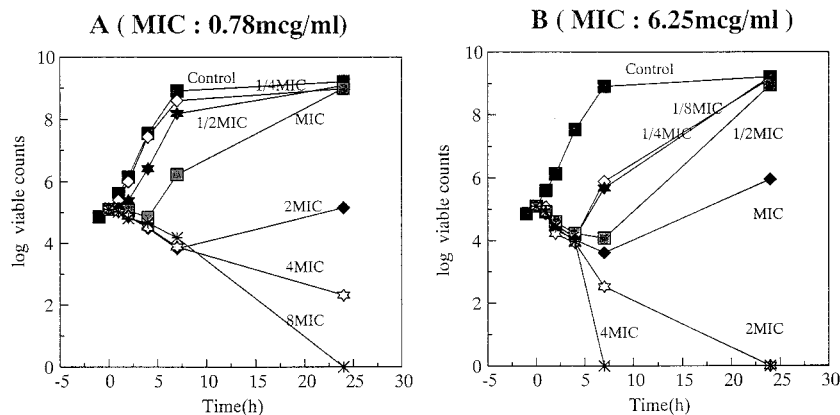


FIG. 2. Bactericidal activities of VCM (A) and OPC-20011 (B) against MRSA 5143.

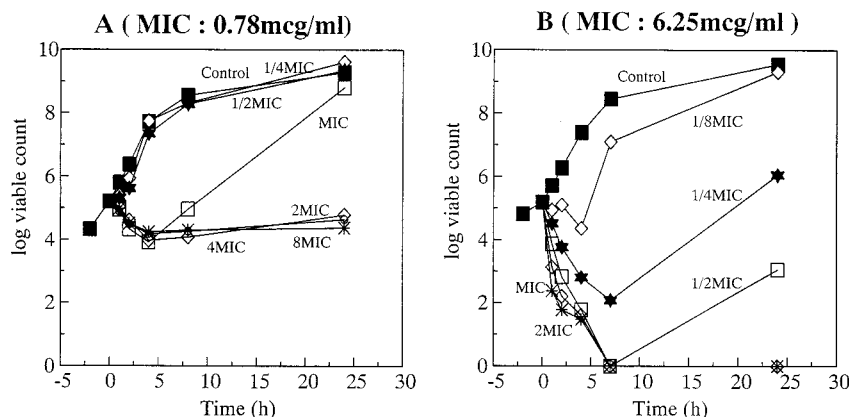


FIG. 3. Bactericidal activities of VCM (A) and OPC-20011 (B) against MRSA 5120.

than that for MRSA 5143, and these bactericidal activities were well correlated to the efficacy in vivo. Also, the distribution of OPC-20011 in lung tissue was approximately 15 µg/ml after a single administration at a dose of 20 mg/kg (data not shown). It is possible that VCM cannot reach sufficient tissue concentrations due to its poor distribution, as the sputum level of VCM in patients is 2 to 3 µg/ml. This is considered to be consistent with the results of our study comparing the therapeutic effects of OPC-20011 and VCM on respiratory tract infection in rats.

The rate of bacterial eradication by VCM in combination with other β-lactams was reported to be approximately 60% in polymicrobial infections (18). Furthermore, it has been reported that during clinical use, VCM induces microbial substitution (1) when used alone, because of its narrow spectrum. Since OPC-20011 has a broad antibacterial spectrum covering not only gram-positive but also gram-negative pathogens, even if used alone, its induction rate of microbial substitution may be less than that of VCM.

OPC-20011 was the most effective of the compounds tested in this study against *S. pneumoniae*. The reason for this potent activity was considered to be the high PBP affinity. Though *S. pneumoniae* C0096 is insensitive to ABPC because of the drug's weak binding to PBP1A, this PBP showed higher affinity for OPC-20011. Although we tried to examine the therapeutic efficacy in a respiratory tract infection model, to our disappointment, we could not establish the model with this organism. However, the efficacy of OPC-20011 was significantly better than those of ABPC and VCM in a corresponding model with the non-ABPC-susceptible *S. pneumoniae* T0005.

In conclusion, OPC-20011 has broad-spectrum antibacterial activity, not only against MRSA, but also against penicillin-resistant *S. pneumoniae* and *E. faecalis*. It is expected to demonstrate activity against gram-negative pathogens equivalent

to those of the selected reference compounds. OPC-20011 is thus considered to be an antibacterial agent which can overcome the shortcomings of some of the existing β-lactam antibiotics.

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TABLE 9. Binding of OPC-20011 to serum proteins from different animal species

Animal	% Binding to serum proteins (± SD)
Mouse.....	29.0 ± 3.0
Rat.....	6.4 ± 2.5
Rabbit.....	<1
Dog.....	5.8 ± 1.5
Monkey.....	1.7 ± 3.1
Human.....	23.2 ± 1.7

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