

## Susceptibilities of *Neisseria gonorrhoeae* Isolates Containing Amino Acid Substitutions in GyrA, with or without Substitutions in ParC, to Newer Fluoroquinolones and Other Antibiotics

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**We examined the antimicrobial susceptibilities of 85 *Neisseria gonorrhoeae* isolates, classified according to the presence of amino acid substitutions in the GyrA and ParC proteins, to 12 fluoroquinolones and 7 other antibiotics. Sitafloxacin and HSR-903 showed excellent activity against *N. gonorrhoeae*, including strains with both GyrA and ParC substitutions. Among the strains with various GyrA substitutions, strains with a serine-91-to-phenylalanine mutation required the highest MICs of all of the fluoroquinolones tested and were cross-resistant to structurally unrelated  $\beta$ -lactams.**

To date, fluoroquinolones have shown excellent clinical efficacy in the treatment of gonorrhea, but the recent emergence of clinical isolates resistant to fluoroquinolones has become a major problem in the treatment of gonococcal infections in several countries, including Japan (6, 9). Clinical failure of gonorrhea treatment has been encountered not only with ciprofloxacin (11) but also with sparfloxacin and pazufloxacin, which have been developed recently (9, 10). We therefore were concerned as to whether the newly constructed fluoroquinolones would have excellent clinical efficacy against gonorrhea caused by fluoroquinolone-resistant isolates in Japan. We also wondered whether the fluoroquinolone-resistant strains were cross-resistant to antimicrobial agents other than fluoroquinolones. In the present investigation, we tested the antimicrobial susceptibilities of *Neisseria gonorrhoeae* isolates, including wild-type strains and strains containing *gyrA* mutations with or without *parC* mutations, to various fluoroquinolones and other antibiotics.

A total of 85 *N. gonorrhoeae* isolates obtained between February 1993 and February 1997 were evaluated. Of the 85 strains, 43 had amino acid substitutions in the quinolone resistance-determining region (QRDR) within the GyrA protein alone, 22 had substitutions in the QRDRs within both the GyrA and ParC proteins, and the remaining 20 had no substitutions within the QRDR in either the GyrA or the ParC protein (wild type). None were posttreatment isolates or repeat isolates from the same patient. The isolates tested were epidemiologically unrelated. *N. gonorrhoeae* strains were identified on the basis of being gram-negative diplococci and by their oxidase reaction and sugar utilization patterns. The PCR and direct DNA sequencing were performed, as described previously (9), to identify mutations in the *gyrA* and *parC* genes of the gonococcal strains. The oligonucleotide primers for the PCR amplification were designed to amplify the genes corre-

sponding to the QRDR within the GyrA and ParC proteins (1, 9).

MICs for all isolates were determined by an agar dilution technique with a GC agar base (Becton Dickinson, Paramus, N.J.) containing 1% IsoVitalX (Becton Dickinson) (8). Plates were inoculated with 5  $\mu$ l of  $10^6$  CFU of each isolate per mL with a multipoint inoculator. World Health Organization reference *N. gonorrhoeae* strains A, B, C, D, and E and *N. gonorrhoeae* ATCC 49226 (8) were included as quality controls. The plates were incubated for 24 h at 35°C in a 5% CO<sub>2</sub> atmosphere.  $\beta$ -Lactamase production was assayed by using the chromogenic cephalosporin test (nitrocefin; Oxoid, Basingstoke, United Kingdom). The preexisting fluoroquinolones tested were norfloxacin, ciprofloxacin, levofloxacin, and sparfloxacin, and the newly constructed fluoroquinolones tested were pazufloxacin, prulifloxacin, grepafloxacin, trovafloxacin, gatifloxacin, sitafloxacin, moxifloxacin, and HSR-903. The non-fluoroquinolone antimicrobial agents were penicillin G, imipenem, ceftriaxone, cefixime, tetracycline, azithromycin, and spectinomycin. Antimicrobial susceptibilities of isolates to ciprofloxacin, penicillin G, tetracycline, ceftriaxone, and spectinomycin were judged by the breakpoint criteria as defined by the National Committee for Clinical Laboratory Standards (NCCLS) (8).

Table 1 shows the antimicrobial susceptibilities of 85 *N. gonorrhoeae* isolates, classified by the presence of amino acid substitutions in GyrA and ParC, to various fluoroquinolones and other antibiotics. The MICs at which 50% of the isolates tested are inhibited (MIC<sub>50</sub>) and MIC<sub>90</sub> of the fluoroquinolones for the strains with both GyrA and ParC substitutions were much higher than those for the wild type and strains with GyrA substitutions alone. However, the MIC<sub>50</sub> and MIC<sub>90</sub> of sitafloxacin and HSR-903 for the strains with both GyrA and ParC substitutions were substantially lower than those of the other fluoroquinolones. All of the strains with GyrA substitutions alone and the wild type were susceptible to ciprofloxacin (MIC  $\leq$  0.06  $\mu$ g/ml), while 14 (63.6%) of the 22 isolates with both GyrA and ParC substitutions were resistant to ciprofloxacin (MIC  $\geq$  1  $\mu$ g/ml). The MIC<sub>90</sub> of the  $\beta$ -lactams for the strains with GyrA substitutions alone were four- or eightfold

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TABLE 1. Antimicrobial susceptibilities of *N. gonorrhoeae* isolates, classified by the presence of amino acid substitutions in GyrA and ParC, to fluoroquinolones and other antibiotics

Antibiotic	Type of substitution (no. of isolates)	MIC ( $\mu\text{g/ml}$ ) <sup>a</sup>		
		Range	MIC <sub>50</sub>	MIC <sub>90</sub>
Sitafloxacin	Wild type (20)	$\leq 0.001-0.004$	0.002	0.004
	GyrA alone (43)	$\leq 0.001-0.063$	0.008 (4 $\times$ )	0.031 (8 $\times$ )
HSR-903	GyrA and ParC (22)	0.008-0.25	0.063 (32 $\times$ )	0.25 (64 $\times$ )
	Wild type (20)	$\leq 0.001-0.004$	0.002	0.004
Gatifloxacin	GyrA alone (43)	$\leq 0.001-0.063$	0.008 (4 $\times$ )	0.031 (8 $\times$ )
	GyrA and ParC (22)	0.008-0.5	0.063 (32 $\times$ )	0.5 (128 $\times$ )
Grepafloxacin	Wild type (20)	0.002-0.008	0.008	0.008
	GyrA alone (43)	0.008-0.5	0.031 (4 $\times$ )	0.063 (8 $\times$ )
Sparfloxacin	GyrA and ParC (22)	0.031-2	0.5 (64 $\times$ )	2 (256 $\times$ )
	Wild type (20)	$\leq 0.001-0.063$	0.008	0.008
Moxifloxacin	GyrA alone (43)	0.008-0.5	0.031 (8 $\times$ )	0.125 (16 $\times$ )
	GyrA and ParC (22)	0.063-4	0.25 (64 $\times$ )	4 (512 $\times$ )
Trovafoxacin	Wild type (20)	$\leq 0.001-0.031$	0.008	0.008
	GyrA alone (43)	0.004-0.5	0.063 (8 $\times$ )	0.25 (32 $\times$ )
Prulifloxacin	GyrA and ParC (22)	0.063-8	0.5 (64 $\times$ )	8 (1,024 $\times$ )
	Wild type (20)	0.002-0.031	0.008	0.008
Ciprofloxacin (MIC $\geq 1 \mu\text{g/ml}$ ) <sup>b</sup>	GyrA alone (43)	0.008-0.5	0.063 (8 $\times$ )	0.5 (64 $\times$ )
	GyrA and ParC (22)	0.125-8	0.5 (64 $\times$ )	8 (1,024 $\times$ )
Levofloxacin	Wild type (20)	0.004-0.016	0.008	0.008
	GyrA alone (43)	0.008-0.5	0.063 (8 $\times$ )	0.5 (64 $\times$ )
Pazufloxacin	GyrA and ParC (22)	0.125-16	2 (256 $\times$ )	8 (1,024 $\times$ )
	Wild type (20)	0.008-0.063	0.016	0.016
Norfloxacin	GyrA alone (43)	0.031-1	0.125 (8 $\times$ )	0.5 (32 $\times$ )
	GyrA and ParC (22)	0.5-16	4 (256 $\times$ )	16 (1,024 $\times$ )
Penicillin G <sup>c</sup> (MIC $\geq 2 \mu\text{g/ml}$ ) <sup>b</sup>	Wild type (19)	0.008-0.063	0.031	0.063
	GyrA alone (38)	0.063-4	0.5 (16 $\times$ )	4 (64 $\times$ )
Imipenem	GyrA and ParC (22)	0.5-16	8 (256 $\times$ )	16 (256 $\times$ )
	Wild type (20)	0.031-1	0.063	0.5
Ceftriaxone (MIC $\leq 0.25 \mu\text{g/ml}$ ) <sup>d</sup>	GyrA alone (38)	0.016-2	0.25 (4 $\times$ )	2 (4 $\times$ )
	GyrA and ParC (22)	0.063-1	0.5 (8 $\times$ )	1 (2 $\times$ )
Cefixime	Wild type (20)	0.031-0.125	0.125	0.125
	GyrA alone (43)	0.031-0.5	0.25 (2 $\times$ )	0.5 (4 $\times$ )
Tetracycline (MIC $\geq 2 \mu\text{g/ml}$ ) <sup>b</sup>	GyrA and ParC (22)	0.063-0.5	0.25 (2 $\times$ )	0.5 (4 $\times$ )
	Wild type (20)	0.002-0.031	0.004	0.016
Azithromycin	GyrA alone (43)	0.002-0.25	0.016 (4 $\times$ )	0.125 (8 $\times$ )
	GyrA and ParC (22)	0.008-0.125	0.063 (16 $\times$ )	0.125 (8 $\times$ )
Spectinomycin (MIC $\geq 128 \mu\text{g/ml}$ ) <sup>b</sup>	Wild type (20)	0.002-0.031	0.008	0.016
	GyrA alone (43)	0.004-0.25	0.016 (2 $\times$ )	0.125 (8 $\times$ )
	GyrA and ParC (22)	0.004-0.125	0.063 (8 $\times$ )	0.063 (4 $\times$ )
	Wild type (20)	0.063-8	0.25	0.5
	GyrA alone (43)	0.063-2	0.5 (2 $\times$ )	1 (2 $\times$ )
	GyrA and ParC (22)	0.125-1	0.5 (2 $\times$ )	1 (2 $\times$ )
	Wild type (20)	0.016-0.5	0.125	0.5
	GyrA alone (43)	0.031-0.5	0.125 (1 $\times$ )	0.5 (1 $\times$ )
	GyrA and ParC (22)	0.016-0.25	0.063 (0.5 $\times$ )	0.25 (0.5 $\times$ )
	Wild type (20)	4-16	8	8
	GyrA alone (43)	4-16	8 (1 $\times$ )	8 (1 $\times$ )
	GyrA and ParC (22)	4-16	8 (1 $\times$ )	8 (1 $\times$ )

<sup>a</sup> Numbers in parentheses indicate fold change compared with wild type.<sup>b</sup> NCCLS criteria for resistance (8).<sup>c</sup> Only non-PPNG strains are shown.<sup>d</sup> NCCLS criteria for sensitivity (8).

TABLE 2. Relationship between amino acid substitutions in GyrA and ParC and antimicrobial susceptibilities of *N. gonorrhoeae* to various fluoroquinolones and other antibiotics

Amino acid substitution(s)		No. of isolates	Mean MIC ( $\mu\text{g/ml}$ ) of antibiotics <sup>a</sup> :						
GyrA	ParC		SIFX	HSR-903	GFLX	GPFX	SPFX	MOFX	TVFX
Wild type	Wild type	20	0.002	0.002	0.006	0.007	0.005	0.01	0.009
A67S	Wild type	1	0.004	0.004	0.016	0.008	0.016	0.016	0.016
A75S	Wild type	5	0.009	0.009	0.019	0.016	0.016	0.028	0.044
A84P	Wild type	1	0.004	0.001	0.016	0.008	0.016	0.031	0.016
S91F	Wild type	21	0.025 (13 $\times$ )	0.021 (11 $\times$ )	0.052 (9 $\times$ )	0.093 (13 $\times$ )	0.09 (18 $\times$ )	0.088 (9 $\times$ )	0.14 (16 $\times$ )
S91C	Wild type	1	0.001	0.002	0.008	0.008	0.008	0.016	0.016
D95G	Wild type	8	0.008	0.009	0.035	0.033	0.059	0.063	0.055
D95N	Wild type	5	0.009	0.007	0.031	0.025	0.047	0.057	0.041
S91F-D95G	Wild type	1	0.063	0.031	0.5	0.5	0.5	1	0.5
S91Y	R116H	1	0.016	0.016	0.063	0.063	0.125	0.125	0.125
S91F	A92G	1	0.031	0.031	0.063	0.25	0.125	0.125	0.5
S91F	A86N	4	0.02	0.014	0.071	0.19	0.19	0.125	0.75
S91F	S88P-E91G	1	0.008	0.016	0.063	0.25	0.25	0.063	0.5
S91F-D95N	E91G	1	0.13	0.063	0.5	2	1	1	0.5
S91F-D95N	S88P	9	0.089	0.072	0.6	0.72	0.69	1.2	0.99
S91F-D95N	S87I-S88P	1	0.25	0.5	1	8	4	4	8
S91F-D95N	S88P-E91K	2	0.25	0.5	2	6	6	4	6
S91F-D95N	S88P-E91G	1	0.125	0.25	1	2	4	2	1
S91F-D95N	S88P-E91Q	1	0.125	0.063	1	0.5	1	0.5	0.125

<sup>a</sup> SIFX, sitafloxacin; GFLX, gatifloxacin; GPFX, grepafloxacin; SPFX, sparfloxacin; MOFX, moxifloxacin; TVFX, trovafloxacin; PUFX, prulifloxacin; CPFX, ciprofloxacin; LVFX, levofloxacin; PZFX, pazufloxacin; NFLX, norfloxacin; PCG, penicillin G; IPM, imipenem; CTRX, ceftriaxone; CFIX, cefixime; TC, tetracycline; AZM, azithromycin; SPCM, spectinomycin. Numbers in parentheses indicate fold change compared with wild type, and for nonfluoroquinolones numbers are shown when threefold or more. NS, data not shown because all strains were PPNG.

higher than those for the wild type. The MIC<sub>90</sub> of the  $\beta$ -lactams for strains with both GyrA and ParC substitutions were almost identical to those for the strains with GyrA substitutions alone. In general, however, the isolates with both GyrA and ParC substitutions were less susceptible to  $\beta$ -lactams at the MIC<sub>50</sub> than the strains with GyrA mutations alone. Five (5.9%) isolates were penicillinase-producing *N. gonorrhoeae* (PPNG). Seven (8.2%) and three (3.5%) isolates had chromosomally mediated resistance to penicillin (MIC  $\geq 2 \mu\text{g/ml}$ ) and tetracycline (MIC  $\geq 2 \mu\text{g/ml}$ ), respectively. However, all of the isolates were susceptible to ceftriaxone (MIC  $\leq 0.25 \mu\text{g/ml}$ ) and spectinomycin (MIC  $\leq 32 \mu\text{g/ml}$ ).

We then investigated the relationship between antimicrobial susceptibility to fluoroquinolones and amino acid substitutions in GyrA and ParC in the gonococcal isolates. Among the strains with the various single GyrA substitutions, those containing a serine (Ser)-to-phenylalanine (Phe) mutation at position 91 (Ser-91 in *N. gonorrhoeae* GyrA corresponds to Ser-83 in *Escherichia coli* [1]) required the highest MICs of all of the fluoroquinolones (Table 2). The strains containing the Ser-91-to-Phe mutation in GyrA also exhibited resistance to structurally unrelated  $\beta$ -lactams (Table 2). All of the seven strains with chromosomally mediated resistance to penicillin contained the Ser-91-to-Phe mutation in GyrA. Five strains with the alanine-75-to-Ser mutation in GyrA required threefold-greater MICs of the cepheims and azithromycin than those required by the wild type (Table 2). These five were all PPNG strains and may have originated in the same clone.

Among the various fluoroquinolones tested, the newly developed sitafloxacin and HSR-903 were more potent than other fluoroquinolones against not only isolates with GyrA alterations alone but also the strains with both GyrA and ParC alterations. The MIC<sub>50</sub> and MIC<sub>90</sub> of sitafloxacin against the strains containing both GyrA and ParC substitutions were 0.063 and 0.25  $\mu\text{g/ml}$ , respectively, while those of HSR-903 were 0.063 and 0.5  $\mu\text{g/ml}$ , respectively. Pharmacokinetic studies of these two fluoroquinolones have demonstrated that, af-

ter administration of a single 200-mg dose of sitafloxacin and HSR-903 to healthy volunteers, their serum drug concentrations peak at 1.86 and 0.86  $\mu\text{g/ml}$ , respectively (reference 7 and K. Uemura, K. Mizuno, and M. Nakashima, Abstr. 36th Intersci. Conf. Antimicrob. Agents Chemother., abstr. F60, p. 100, 1996). These data corroborate that a single 200-mg-or-more dose of sitafloxacin or HSR-903 is likely to show excellent clinical efficacy against gonorrhea caused by fluoroquinolone-resistant *N. gonorrhoeae* carrying both GyrA and ParC substitutions. The increment in MIC of norfloxacin associated with the Ser-91-to-Phe mutation in GyrA is higher than is usually seen and suggests the possibility that other unidentified additional mutations such as efflux-type mutations (5) or porin gene mutations (4) contribute to resistance in these strains.

Interestingly, cross-resistance between the fluoroquinolones and the structurally unrelated  $\beta$ -lactams was observed in the gonococcal isolates. Other investigations have also reported such cross-resistance (2, 3). Among strains with the various substitutions, strains containing the Ser-91-to-Phe mutation showed significant resistance to penicillin G, imipenem, and cepheims. The antimicrobial susceptibilities of isolates with other substitutions in GyrA alone to the antibiotics were comparable to those of the wild type. The susceptibilities of the strains with multiple substitutions to  $\beta$ -lactams were lower than those of the wild type but comparable to those of the isolates with the single Ser-91-to-Phe alteration. These results indicate that the Ser-91-to-Phe mutation may be important in cross-resistance to other structurally unrelated agents. We were unable to explain why strains containing the Ser-91-to-Phe mutation show cross-resistance to structurally unrelated  $\beta$ -lactams. Some of the strains with the Ser-91-to-Phe alteration in GyrA may have had efflux-type mutations (5) or porin gene mutations (4) that affect responses to quinolones and other drugs. Further study is necessary to investigate cross-resistance between fluoroquinolones and  $\beta$ -lactams.

TABLE 2—Continued

Mean MIC ( $\mu\text{g/ml}$ ) of antibiotics <sup>a</sup>											
PUFX	CPFX	LVFX	PZFX	NFLX	PCG	IPM	CTRX	CFIX	TC	AZM	SPCM
0.008	0.007	0.012	0.016	0.028	0.23	0.11	0.008	0.01	0.69	0.17	7.6
0.008	0.016	0.031	0.063	0.125	0.016	0.063	0.002	0.008	0.063	0.063	8
0.025	0.031	0.069	0.069	0.25	NS	0.25	0.061 (8 $\times$ )	0.031 (3 $\times$ )	1.3	0.5 (3 $\times$ )	7.2
0.016	0.016	0.031	0.063	0.125	0.063	0.063	0.004	0.004	0.063	0.125	8
0.22 (28 $\times$ )	0.24 (34 $\times$ )	0.34 (28 $\times$ )	0.48 (30 $\times$ )	1.6 (57 $\times$ )	1.1 (5 $\times$ )	0.41 (4 $\times$ )	0.092 (12 $\times$ )	0.074 (7 $\times$ )	0.71	0.19	7.8
0.016	0.008	0.016	0.031	0.063	0.031	0.031	0.004	0.004	1	0.063	8
0.051	0.029	0.071	0.071	0.2	0.067	0.059	0.012	0.007	0.14	0.086	11
0.063	0.052	0.088	0.11	0.35	0.088	0.13	0.007	0.009	0.25	0.1	7.2
0.25	0.5	0.5	1	2	0.125	0.063	0.008	0.008	0.5	0.25	4
0.125	0.125	0.25	0.5	0.5	0.25	0.063	0.063 (8 $\times$ )	0.008	0.25	0.063	8
0.25	0.25	0.5	1	2	1 (4 $\times$ )	0.5 (5 $\times$ )	0.063 (8 $\times$ )	0.063 (6 $\times$ )	1	0.25	8
0.5	0.22	0.38	1	1.8	0.75 (3 $\times$ )	0.19	0.079 (10 $\times$ )	0.055 (6 $\times$ )	0.88	0.063	8
0.25	0.25	0.25	0.5	1	0.5	0.25	0.063 (8 $\times$ )	0.063 (6 $\times$ )	0.5	0.125	8
0.5	2	2	2	4	0.25	0.25	0.016	0.016	0.5	0.25	8
1.4	1.8	3.9	6	9.8	0.78 (3 $\times$ )	0.28 (3 $\times$ )	0.056 (7 $\times$ )	0.063 (6 $\times$ )	0.64	0.11	9.3
8	16	8	16	16	0.5	0.25	0.063 (8 $\times$ )	0.063 (6 $\times$ )	0.5	0.125	8
8	12	8	16	16	1 (4 $\times$ )	0.19	0.094 (12 $\times$ )	0.063 (6 $\times$ )	0.31	0.063	8
4	4	16	16	16	0.5	0.25	0.031 (4 $\times$ )	0.031 (3 $\times$ )	0.5	0.063	8
2	2	4	8	8	0.063	0.125	0.008	0.031 (3 $\times$ )	0.5	0.063	8

## REFERENCES

- Belland, R. J., S. G. Morrison, C. A. Ison, and W. M. Huang. 1994. *Neisseria gonorrhoeae* acquires mutations in analogous regions of *gyrA* and *parC* in fluoroquinolone-resistant isolates. *Mol. Microbiol.* **14**:371–380.
- Deguchi, T., M. Yasuda, M. Nakano, S. Ozeki, T. Ezaki, I. Saito, and Y. Kawada. 1996. Quinolone-resistant *Neisseria gonorrhoeae* correlation of alterations in the GyrA subunit of DNA gyrase and the ParC subunit of topoisomerase IV with antimicrobial susceptibility profiles. *Antimicrob. Agents Chemother.* **40**:1020–1023.
- Fox, K. K., J. S. Knapp, K. K. Holmes, E. W. Hook III, F. N. Judson, S. E. Thompson, J. A. Washington, and W. L. Whittington. 1997. Antimicrobial resistance in *Neisseria gonorrhoeae* in the United States, 1988–1994: the emergence of decreased susceptibility to the fluoroquinolones. *J. Infect. Dis.* **175**:1396–1403.
- Gill, M. J., S. Simjee, K. Al-Hattawi, B. D. Robertson, C. S. F. Easmon, and C. A. Ison. 1998. Gonococcal resistance to  $\beta$ -lactams and tetracycline involves mutation in loop 3 of the porin encoded at the *penB* locus. *Antimicrob. Agents Chemother.* **42**:2799–2803.
- Hagman, K. E., and W. M. Shafer. 1995. Transcriptional control of the *mnt* efflux system of *Neisseria gonorrhoeae*. *J. Bacteriol.* **177**:4162–4165.
- Ison, C. A., J. A. R. Dillon, and J. W. Tapsall. 1998. The epidemiology of global antibiotic resistance among *Neisseria gonorrhoeae* and *Haemophilus ducreyi*. *Lancet* **351**(Suppl. III):8–11.
- Nakashima, M., T. Uematsu, K. Kosuge, K. Umemura, H. Hakusui, and M. Tanaka. 1995. Pharmacokinetics and tolerance of DU-6859a, a new fluoroquinolone, after single and multiple oral doses in healthy volunteers. *Antimicrob. Agents Chemother.* **39**:170–174.
- National Committee for Clinical Laboratory Standards. 1998. Performance standards for antimicrobial susceptibility testing. Eighth informational supplement. Document M100-S8, vol. 18, no. 1. National Committee for Clinical Laboratory Standards, Wayne, Pa.
- Tanaka, M., T. Matsumoto, M. Sakumoto, K. Takahashi, T. Saika, I. Kobayashi, J. Kumazawa, and The Pazufloxacin STD Group. 1998. Reduced clinical efficacy of pazufloxacin against gonorrhea due to high prevalence of quinolone-resistant isolates with the GyrA mutation. *Antimicrob. Agents Chemother.* **42**:579–582.
- Tanaka, M., H. Nakayama, M. Haraoka, T. Nagafuji, T. Saika, and I. Kobayashi. 1998. Analysis of quinolone resistance mechanisms in a sparfloxacin-resistant clinical isolate of *Neisseria gonorrhoeae*. *Sex. Transm. Dis.* **25**:489–493.
- Tapsall, J. W., E. A. Limnios, C. Thacker, B. Donovan, S. D. Lynch, L. J. Kirby, K. A. Wise, and C. J. Carmody. 1995. High-level quinolone resistance in *Neisseria gonorrhoeae*: a report of two cases. *Sex. Transm. Dis.* **22**:310–311.