

## Horizontal Transfer of *parC* and *gyrA* in Fluoroquinolone-Resistant Clinical Isolates of *Streptococcus pneumoniae*

MARÍA JOSÉ FERRÁNDIZ,<sup>1</sup> ASUNCIÓN FENOLL,<sup>2</sup> JOSEFINA LIÑARES,<sup>3</sup>  
AND ADELA G. DE LA CAMPA<sup>1\*</sup>

Unidad de Genética Bacteriana (Consejo Superior de Investigaciones Científicas), Centro Nacional de Biología Fundamental,<sup>1</sup> and Servicio de Bacteriología, Centro Nacional de Microbiología,<sup>2</sup> Instituto de Salud Carlos III, 28220 Majadahonda, Madrid, and Servicio de Microbiología, Hospital Princesps d'Espanya, Ciutat Sanitària i Universitaria de Bellvitge, 08907 l'Hospitalet de Llobregat, Barcelona,<sup>3</sup> Spain

Received 30 July 1999/Returned for modification 4 November 1999/Accepted 27 December 1999

**We have analyzed genetically three clinical isolates (3180, 3870, and 1244) of *Streptococcus pneumoniae* with high-level ciprofloxacin resistance. Isolates 3180 and 3870 were atypical because of their insolubility in deoxycholate. However, they hybridized specifically with pneumococcal autolysin and pneumolysin gene probes and have typical pneumococcal *atpC* and *atpA* gene sequences. Analysis of the complete sequences of the *parC* and *gyrA* genes revealed total variations of 8 and 8.7% (isolate 3180) and 7.4 and 3.6% (isolate 3870), respectively, compared to the wild-type strain R6 sequence. The variations observed between the sequences of R6 and isolate 1244 were less than 0.9%. The structure of the *gyrA* and *parC* genes from isolates 3180 and 3870 was organized in sequence blocks that show different levels of divergence, suggesting a pattern of recombination. These results are evidence for recombination at the fluoroquinolone target genes in clinical isolates of *S. pneumoniae*. The genetically related viridans group streptococci could act as a reservoir for fluoroquinolone resistance genes.**

*Streptococcus pneumoniae* is the most common bacterial cause of community-acquired pneumonia, meningitis, otitis media, and sinusitis. The emergence of resistance to antimicrobial agents commonly used for the treatment of pneumococcal diseases (5, 17, 25, 38) has made very difficult the selection of optimal antimicrobial therapies for the treatment of pneumococcal infections. A parallel increasing resistance to penicillin and macrolide antibiotics has been also observed for the viridans group streptococci (1, 2, 7, 8). These microorganisms are, like *S. pneumoniae*, commensals of the oropharyngeal tract. Nevertheless, they are causative organisms of infective endocarditis (12, 44, 47) and are also a major cause of bacteremia in neutropenic cancer patients (3, 7, 8, 16).

There is considerable interest in the use of alternative antimicrobial agents, such as the new fluoroquinolones, with good activity against streptococci for the treatment of respiratory tract infections (6). The prevalence of ciprofloxacin resistance in *S. pneumoniae* has been found to be low in Spain (<3%) (32, 33); similar data have been reported in Canada (9). The prior administration of fluoroquinolones could be an important risk factor for quinolone-resistant strain selection, as has been observed for respiratory tract infections caused by ciprofloxacin-resistant (Cp<sup>r</sup>) *S. pneumoniae* (41). Likewise, fluoroquinolone resistance has been reported for blood isolates of viridans group streptococci from neutropenic cancer patients who received quinolone prophylaxis (22, 50). The prevalence of resistance to ciprofloxacin (MIC,  $\geq 4$   $\mu\text{g/ml}$ ) in viridans group streptococci consecutively isolated from different clinical sources from 1993 to 1998 at Hospital Princesps d'Espanya was as follows: 17.8% (135 of 756 clinical isolates) for *Streptococcus mitis*, 12.0% (10 of 83 isolates) for *Streptococcus salivarius*, 2.9% (11 of 378 clinical isolates) for *Streptococcus sanguis*, and

2.3% (13 of 575 isolates) for *Streptococcus anginosus* (unpublished data). These data are in accordance with those obtained in Canada, which showed a prevalence of resistance to ciprofloxacin of 11.4% (27 of 236 isolates) for the viridans group streptococci, *S. mitis* and *S. salivarius* being the most resistant (10).

The principal targets of the fluoroquinolones are DNA gyrase (gyrase) and DNA topoisomerase IV (topo IV), members of the topoisomerase family of enzymes that control bacterial DNA topology (15). Both enzymes function by passing one DNA double helix through another, using a transient double-strand break (35). Gyrase, an A<sub>2</sub>B<sub>2</sub> complex encoded by the *gyrA* and *gyrB* genes, catalyzes ATP-dependent negative supercoiling of DNA and is involved in DNA replication, recombination, and transcription (53); topo IV, a C<sub>2</sub>E<sub>2</sub> complex encoded by the *parC* and *parE* genes, is essential in chromosome partitioning (35). The deduced amino acid sequences of ParC and ParE are homologous to those of GyrA and GyrB, respectively (30). Genetic studies from a number of laboratories (23, 28, 36, 39, 49) have shown that topo IV is the primary target for ciprofloxacin in *S. pneumoniae* and that gyrase is the secondary target. Resistance mutations have been identified in a discrete region of ParC, ParE, GyrA, and GyrB termed the quinolone resistance-determining region (QRDR). We recently reported the same mechanism for viridans group streptococci (22): low-level Cp<sup>r</sup> strains had mutations altering one of the two subunits of topo IV.

The viridans group streptococci could be a reservoir of fluoroquinolone resistance genes if we assume that resistance in viridans group streptococci and *S. pneumoniae* arose from horizontal transfer, as has been observed with penicillin resistance (46). A number of observations suggest that this transfer between viridans group streptococci and *S. pneumoniae* could be a possible mechanism for the spread of fluoroquinolone resistance. The viridans group streptococci and *S. pneumoniae* share the same mechanism of resistance (22). The nucleotide sequences of their gyrase and topo IV genes show high identity (20, 22), and it is possible to transform *S. pneumoniae* cells to

\* Corresponding author. Mailing address: Unidad de Genética Bacteriana (Consejo Superior de Investigaciones Científicas), Centro Nacional de Biología Fundamental, Instituto de Salud Carlos III, 28220 Majadahonda, Madrid, Spain. Phone: (34) 91-5097904. Fax: (34) 91-5097919. E-mail: agcampa@isciii.es.

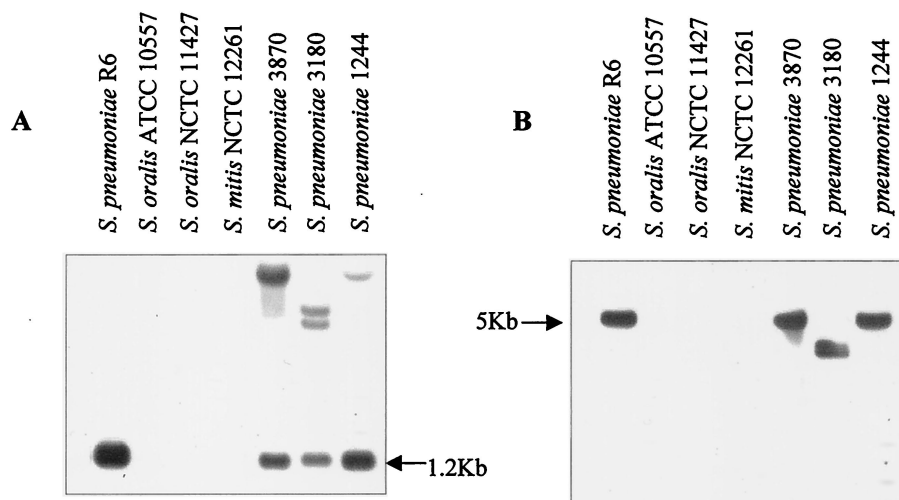


FIG. 1. Identification of *S. pneumoniae* isolates by hybridization with specific DNA probes. Chromosomal DNAs from the *S. oralis*, *S. mitis*, and *S. pneumoniae* strains indicated were cleaved with *Hind*III (A) or *Cla*I (B), and the fragments were separated in 1% agarose gels. The gels were blotted, and the blots were probed with biotinylated DNA as follows: A, insert of plasmid pCE3 containing the N terminus of the *lytA* gene; and B, insert of plasmid pJCP191 containing the *pnl* gene.

ciprofloxacin resistance with DNA from Cp<sup>r</sup> viridans group streptococci (22, 27). Additionally, nucleotide sequence comparisons of the DNA topoisomerase genes of the viridans group streptococci (20, 22) show a high level of intraspecies variation. These observations suggest that the viridans group streptococci could be considered a group of species that interchange genetic material between them and possibly with *S. pneumoniae*. In this report, we describe the characterization of Cp<sup>r</sup> *S. pneumoniae* isolates with a mosaic structure in their *parC* and *gyrA* genes, suggesting such interspecies recombination.

#### MATERIALS AND METHODS

**Southern blot identification of *S. pneumoniae* strains.** The ciprofloxacin-sensitive (Cp<sup>s</sup>) strain of *S. pneumoniae* used was the wild-type strain R6. The Cp<sup>r</sup> clinical isolates were obtained from sputum samples at the Hospital Princesps d'Espanya (Barcelona, Spain) in 1992 (strain 1244), 1994 (strain 3180), and 1996 (strain 3870). Plasmid pCE3 (18), containing a 0.65-kb fragment coding for the N terminus of the major pneumococcal autolysin (amidase), was used as a source of the *lytA* DNA probe. Plasmid pJCP191 (48), containing a 1.6-kb fragment coding for the complete pneumococcal pneumolysin gene, was used as a source of the *pnl* DNA probe and was kindly provided by S. Taira. The inserts of pCE3 and pJCP191 were isolated after digestion with *Hind*III-*Hinc*II and *Pvu*II, respectively. The resulting DNA inserts were labeled with the Phototope-Star Detection Kit (New England Biolabs). Southern blotting and hybridization were done by following the manufacturer's instructions.

**Amplification and analysis of genes.** Genes were amplified from genomic DNA by the PCR as described previously (22). The *atpCA* region and the *parC* and *gyrA* genes were amplified with the following primers, based on published sequences (4, 19, 36, 40): *atpCUP* (5'-dAAAGGAGAATTTGTTATGAA-3'), corresponding to nucleotides -15 to +5 of *atpC*, and *atpB56* (5'-dGACGGGC TTCTTCAGCTCTGTC-3'), complementary to nucleotides 147 to 169 of *atpB*; *parCUP* (5'-dGAACACGCCCCTAGATACTGTG-3'), corresponding to nucleotides -103 to -83 of *parC*, and *parCDOWN* (5'-dCGTTACTGTTCATATCC ACTCC-3'), complementary to nucleotides 120 to 142 downstream of *parC*; and *gyrAUP1* (4) and *gyrADOWN* (4). DNA fragments were purified with Micro-Spin S400 HR columns (Pharmacia) and were sequenced on both strands by use of an Applied Biosystems Prism 377 sequencer with the primers used for PCR amplification and with internal primers. For nucleotide sequence comparisons, in addition to the Cp<sup>s</sup> *S. pneumoniae* strain R6 (GenBank accession no. AF170996 and AF053121 for *parC* and *gyrA*, respectively), two other sequences were used: the sequences of the Cp<sup>s</sup> strain 7785 (accession no. Z67739 and AJ005815 for *parC* and *gyrA*, respectively) and of another, unknown isolate, which we call AB (accession no. for *gyrA*, AB010387).

**Nucleotide sequence accession numbers.** The new DNA sequences reported in this paper have been assigned the following GenBank accession no.: AF170996 to AF170999 (*parC* genes), AF170993 to AF170995 (*gyrA* genes), and AF171000 to AF171002 (*atpCA* regions).

#### RESULTS AND DISCUSSION

**Identification of strains.** Three clinical isolates, 3180, 3870, and 1244, were analyzed in this work. These isolates were previously described as *Streptococcus oralis* 3180 (22), *S. oralis* 3870 (22), and *S. pneumoniae* 1244 (36). Initial characterization of the three isolates by colony morphology on blood agar and optochin susceptibility identified them as pneumococcal strains. However, strains 3180 and 3870 were insoluble in deoxycholate, while strain 1244 was soluble. Phenotypic characterization of isolates 3180 and 3870 by the API 32 Strep system classified them as *S. oralis* (22). Given the unreliability of this method for the identification of *S. pneumoniae* (20, 31), the isolates were studied by hybridization with two pneumococcal probes. One of the probes coded for the N terminus of the major pneumococcal autolysin (*lytA*), and the other coded for the complete pneumococcal pneumolysin (*pnl*). The *S. pneumoniae* strain R6 showed, as expected, hybridization with the *lytA* probe in a 1.2-kb *Hind*III chromosomal fragment (21), while *S. oralis* and *S. mitis* type strains did not (Fig. 1A). Strains 3180, 3870, and 1244 showed high-molecular-weight hybridization bands with the *lytA* probe, in addition to the 1.2-kb *Hind*III fragment (Fig. 1A). These bands could have resulted from hybridization with homologous *lytA* genes of pneumococcal prophages, which have been described to be very frequent in pneumococcal clinical isolates (43). Hybridization with the pneumolysin probe detected a single 5-kb band in *S. pneumoniae* R6 *Cla*I-digested DNA (Fig. 1B), as expected from the physical map of the *pnl* chromosomal region (48, 52), while no hybridization was observed with the *S. oralis* and *S. mitis* type strains. The three Cp<sup>r</sup> *S. pneumoniae* clinical isolates (3180, 3870, and 1244) all hybridized with the *pnl* probe. Because both LytA and pneumolysin proteins have been demonstrated to be species specific (18, 24, 29, 42, 45, 51), these results identified the three isolates as *S. pneumoniae*.

Sequencing of the *atpC* and *atpA* genes allowed further characterization of the strains. The *atpC* gene is responsible for the characteristic optochin susceptibility phenotype of pneumococci (19, 37). The sequences of a region spanning 960 nucleotides, including *atpC* and *atpA*, showed high homogeneity (data not shown). The three Cp<sup>r</sup> *S. pneumoniae* strains

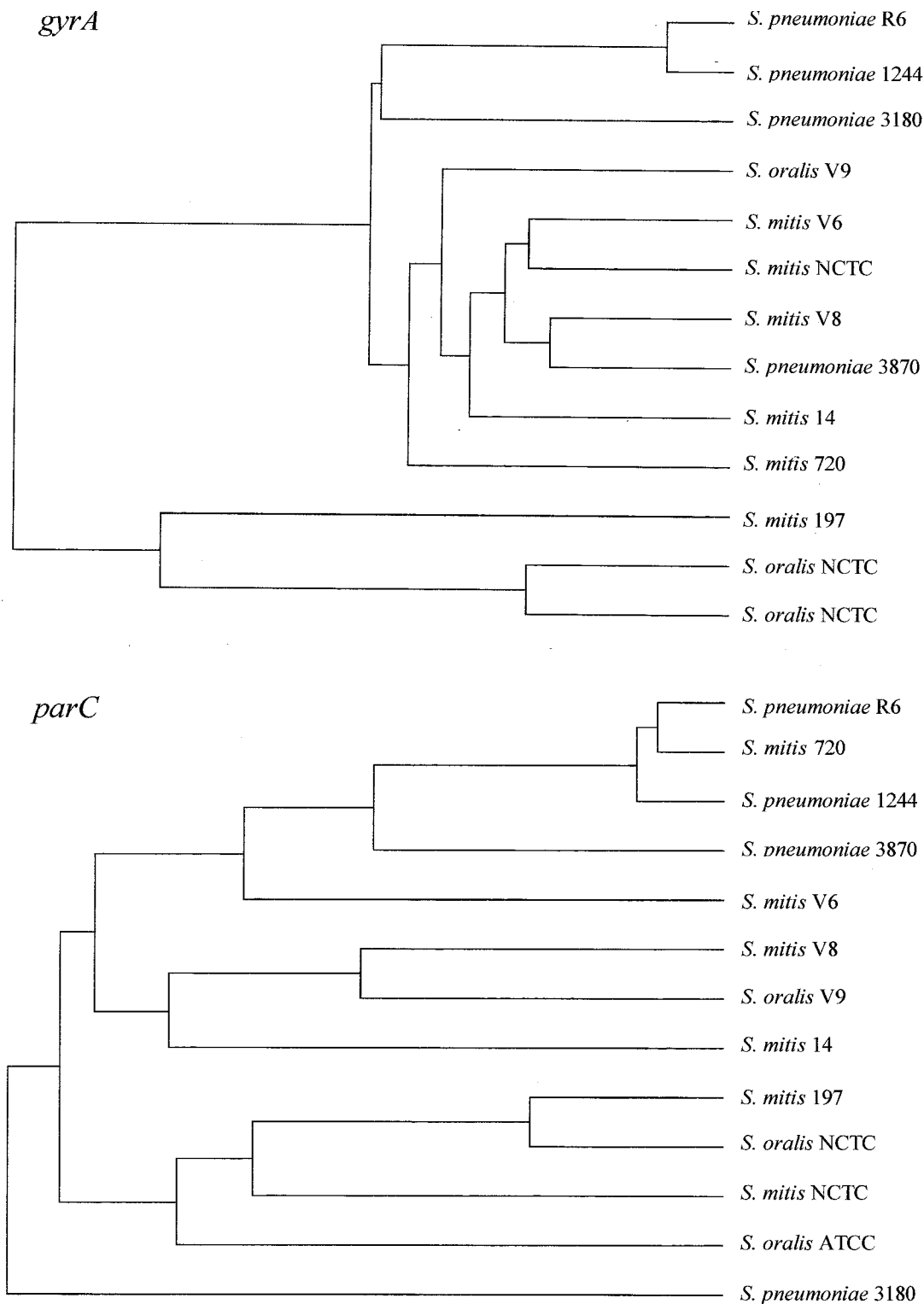


FIG. 2. Trees of nucleotide sequences of *parC* and *gyrA* QRDRs. The 185-nucleotide *parC* sequence included positions 213 to 397, and the 280-nucleotide *gyrA* sequence included positions 175 to 454. Nucleotides are numbered by taking the first *gyrA* and *parC* nucleotides as nucleotide 1. The trees were compiled by using the CLUSTAL multiple-alignment program from PCGENE with default parameters. The nucleotide sequences used have been previously reported (20, 22, 36).

showed less than 0.6% variation, compared with the wild-type strain R6, while the *S. oralis* NCTC 11427 type strain showed 20% variation. These values are in agreement with the results of comparisons of the amyломaltase gene sequences:  $\leq 0.5\%$  *S.*

*pneumoniae* intraspecies variation (14) and 4 to 6% divergence between *S. pneumoniae* and *S. oralis* (13).

The results of comparisons of the *atpCA* region are consistent with the results of Southern blot hybridization and iden-





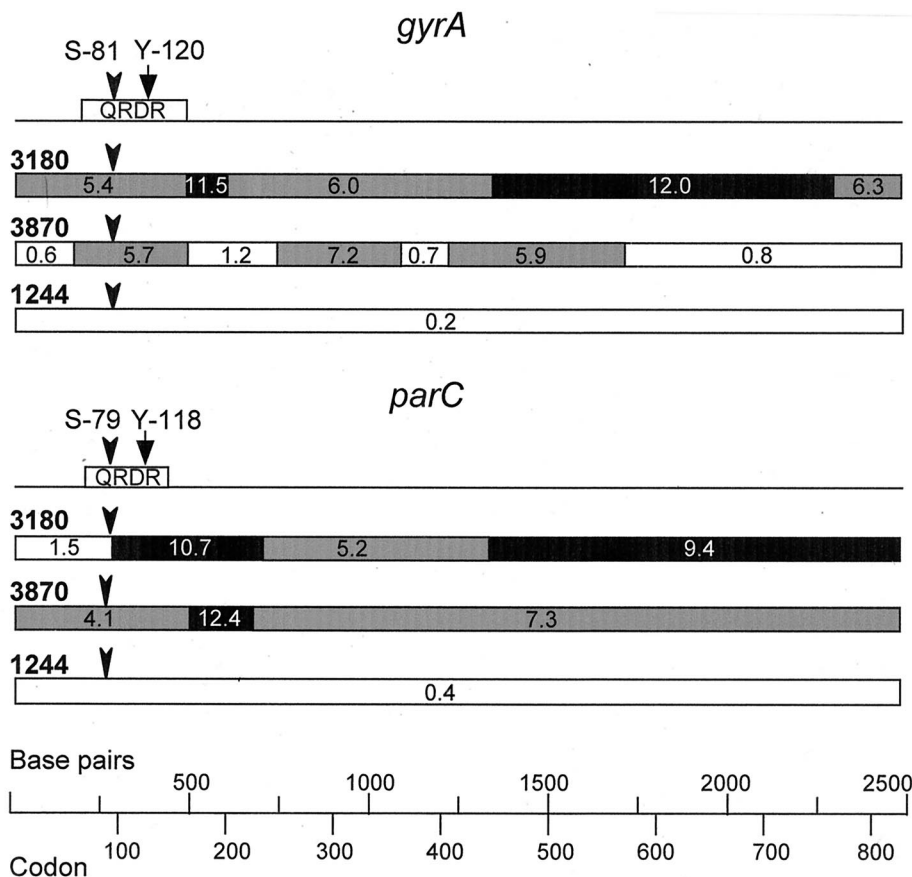


FIG. 5. Mosaic structure of the *gyrA* and *parC* genes of Cp<sup>+</sup> *S. pneumoniae* isolates 3180 and 3870. The locations of the QRDRs are represented above the *gyrA* and *parC* sequences. The positions of the active Tyr residues (Y-120 in GyrA and Y-118 in ParC) that bind DNA and of the Ser residues that are changed in the Cp<sup>r</sup> strains (S-81 in GyrA and S-79 in ParC) and are involved in resistance are marked. Blocks showing the percent sequence divergence from the corresponding regions in Cp<sup>s</sup> pneumococci are indicated. White boxes, regions of sequence that differ by  $\leq 1.5\%$ ; grey boxes, regions that differ by more than 1.5% but less than 9%; black boxes, regions that differ by  $>9\%$ .

(TTC) in isolate 3870. The other change, present in both isolates, was Gly-144 to Ser (AGT to GGT). The analysis of Cp<sup>r</sup> transformants obtained with DNA from the *gyrA* QRDRs of isolates 3180 and 3870 showed that the amino acid changes at Ser-81 were indeed involved in resistance (22).

A comparison of the nucleotide sequences of the *parC* and *gyrA* QRDRs of isolates 3180, 3870, and 1244, *S. pneumoniae* R6, and several *S. oralis* and *S. mitis* strains is shown in Fig. 2. While isolate 1244 was grouped with *S. pneumoniae* R6 within the *parC* or *gyrA* tree, the location of isolates 3180 and 3870 varied depending on the gene considered. These results suggested a recombinational origin for the genes encoding the fluoroquinolone target proteins of isolates 3180 and 3870. Such a situation would be due to genetic transformation with DNA from other bacterial species, probably the genetically closed related viridans group streptococci. The recombination machinery requires approximately 80% sequence identity between two homologous DNA molecules (19, 26). At least 87% identity was found between the *parC* and *gyrA* QRDRs of isolates 3180 and 3870 and those of the viridans group streptococci (Fig. 2). Similarly, horizontal transfer of altered penicillin-binding protein genes between *S. pneumoniae* and viridans group streptococci (46) has been observed.

To test this hypothesis, the nucleotide sequences of the complete *parC* and *gyrA* genes of the two isolates (3180 and 3870) that showed high divergence in their *parC* and *gyrA*

QRDRs and one isolate (1244) that did not show this divergence were determined. The results of sequence comparisons of Cp<sup>s</sup> and Cp<sup>r</sup> strains (Fig. 3 and 4 and Table 1) clearly showed three groups of strains. One group, with a nucleotide sequence variation of  $<1\%$ , was formed by the sensitive strains and isolate 1244. Isolates 3180 and 3870 each formed separate groups, since the nucleotide sequence variations between the sequences of the two isolates were 7.6% for their *parC* sequences and 8% for their *gyrA* sequences. The average variations in the *parC* and *gyrA* sequences between isolate 3180 and the first group of strains (sensitive strains and isolate 1244) were  $\geq 8\%$ , while these values for isolate 3870 were about 7% for *parC* and about 3% for *gyrA* (Table 1). The variations found in isolates 3180 and 3870 could be organized in blocks (Fig. 5) with different degrees of relatedness. The limits of the blocks were determined by inspection, with the only limitation being at least a 4% difference in divergence between two contiguous blocks.

From these results, we can assume that the first group of strains (sensitive strains and isolate 1244) are nonrecombinant isolates, while isolates 3180 and 3870 show a recombinational pattern probably resulting from gene transfer events. The *gyrA* sequence of isolate 3870 clearly shows four blocks of low divergence ( $\leq 1.2\%$ ) and three blocks of high divergence ( $\geq 5.7\%$ ). This result suggests that the low-divergence blocks represent regions in which interspecies recombination events

had occurred. Another block with a 1.5% divergence was observed in *parC* of isolate 3180, although in this case the second recombination point should be located outside the gene. Likewise, recombination outside the genes would be the origin of the *gyrA* gene of isolate 3180 and of the *parC* gene of isolate 3870.

Because both gyrase and topo IV are tetrameric proteins, an interchange of *parC* would need an accompanying interchange of *parE*. Since both genes are contiguous in the pneumococcal chromosome, we cannot exclude the possibility of a recombinational event involving both genes. On the other hand, an interchange of both *gyrA* and *gyrB* genes would involve two independent recombinations, since the genes are separated by at least 90 kb in the chromosome (36).

Two different processes could lead to the acquisition of fluoroquinolone resistance: spontaneous mutation and transformation. A comparison of the frequencies of these two processes reveals that transformation could be several orders of magnitude more frequent than mutation. The frequency of mutation to  $Cp^r$  in *S. pneumoniae* has been shown to be in the range of  $10^{-8}$  to  $10^{-9}$  (39). However, the frequencies of transformation to  $Cp^r$  with chromosomal DNAs from  $Cp^r$  *S. pneumoniae* strains were in the range of  $10^{-2}$  (36, 49) for monogenic transformation (low-level  $Cp^r$ ) (36, 49). The acquisition of low-level  $Cp^r$  via transformation could then be  $10^6$  to  $10^7$  times more frequent than that via spontaneous mutation. Likewise, it has been shown that the frequency of transformation of *S. pneumoniae* competent cells to low-level  $Cp^r$  with DNA from  $Cp^r$  *S. mitis* is in the range of  $10^{-3}$  (22, 27). Interspecies transformation could thus be  $10^5$  to  $10^6$  more frequent than spontaneous mutation. These differences are even higher when the acquisition of high-level resistance is considered. The frequencies of transformation with two unlinked markers that gave rise to high-level  $Cp^r$  were  $10^{-4}$  when both donor and recipient cells were *S. pneumoniae* (36, 49) and  $10^{-6}$  when competent *S. pneumoniae* cells were transformed with *S. mitis* DNA (27). However, two spontaneous mutations are necessary to obtain a high level of resistance (i.e., the frequency could be  $10^{-14}$  to  $10^{-16}$ ). Nevertheless, these estimates are not necessarily true since, as pointed out above, an interchange of *parC* would need an accompanying interchange of *parE* and an interchange of *gyrA* would need an accompanying interchange of *gyrB*. Other factors to be considered for transformation in the natural environment are the availability of DNA and the competence state of the recipient cells.

#### ACKNOWLEDGMENTS

We thank P. A. Lazo for allowing us to use the PCGENE program and A. Rodríguez-Bernabé for excellent technical assistance.

M.J.F. has a fellowship from Comunidad Autónoma de Madrid. This work was supported by grant 97/2026 from Fondo de Investigación Sanitaria.

#### REFERENCES

- Alcaide, F., J. Carratalá, J. Liñares, F. Gudiol, and R. Martín. 1996. In vitro activities of eight macrolide antibiotics and RP-59500 (quinupristin-dalfopristin) against viridans group streptococci isolated from blood of neutropenic cancer patients. *Antimicrob. Agents Chemother.* **40**:2117–2120.
- Alcaide, F., J. Liñares, R. Pallarés, J. Carratalá, M. A. Benítez, F. Gudiol, and R. Martín. 1995. In vitro activity of 22  $\beta$ -lactam antibiotics against penicillin-resistant and penicillin-susceptible viridans group streptococci isolated from blood. *Antimicrob. Agents Chemother.* **39**:2243–2247.
- Awada, A. P., P. Van der Auwera, P. Meunier, D. Daneau, and J. Klasterksy. 1992. Streptococcal and enterococcal bacteremia in patients with cancer. *Clin. Infect. Dis.* **15**:33–48.
- Balas, D., E. Fernández-Moreira, and A. G. de la Campa. 1998. Molecular characterization of the gene encoding the DNA gyrase A subunit of *Streptococcus pneumoniae*. *J. Bacteriol.* **180**:2854–2861.
- Baquero, F. 1996. Epidemiology and management of penicillin-resistant pneumococci. *Curr. Opin. Infect. Dis.* **9**:372–379.
- Bartlett, J. G., R. F. Breiman, L. Mandell, and T. M. File. 1998. Community-acquired pneumonia in adults: guidelines for management. *Clin. Infect. Dis.* **26**:811–838.
- Bochud, P. Y., P. H. Eggiman, T. Calandra, G. Van Melle, L. Saghafi, and P. Francioli. 1994. Bacteremia due to viridans streptococcus in neutropenic patients with cancer: clinical spectrum and risk factors. *Clin. Infect. Dis.* **18**:25–31.
- Carratalá, J., F. Alcaide, A. Fernández-Sevilla, X. Corbell, J. Liñares, and F. Gudiol. 1995. Bacteremia due to viridans streptococci that are highly resistant to penicillin: increase among neutropenic patients with cancer. *Clin. Infect. Dis.* **20**:1169–1173.
- Chen, D. K., A. McGeer, J. C. De Azevedo, and D. E. Low. 1999. Decreased susceptibility of *Streptococcus pneumoniae* to fluoroquinolones in Canada. *N. Engl. J. Med.* **341**:233–239.
- de Azevedo, J. C. S., L. Trpeski, S. Pong-Porter, S. Matsumura, T. C. B. S. Network, and D. E. Low. 1999. In vitro activity of fluoroquinolones against antibiotic-resistant blood culture isolates of viridans group streptococci across Canada. *Antimicrob. Agents Chemother.* **43**:2299–2301.
- Díaz, E., R. López, and J. L. García. 1992. Role of the major pneumococcal autolysin in the atypical response of a clinical isolate of *Streptococcus pneumoniae*. *J. Bacteriol.* **174**:5508–5515.
- Douglas, C. W. L., J. Heath, K. K. Hampton, and F. E. Preston. 1993. Identity of viridans streptococci isolated from cases of infective endocarditis. *J. Med. Microbiol.* **39**:179–182.
- Dowson, C. G., A. Hutchinson, N. Woodford, A. P. Johnson, R. C. George, and B. G. Spratt. 1990. Penicillin-resistant viridans streptococci have obtained altered penicillin-binding protein genes from penicillin-resistant strains of *Streptococcus pneumoniae*. *Proc. Natl. Acad. Sci. USA* **87**:5858–5862.
- Dowson, C. G., A. Hutchison, J. A. Brannigan, R. C. George, D. Hansman, J. Liñares, A. Tomasz, J. Maynard Smith, and B. G. Spratt. 1989. Horizontal transfer of penicillin-binding protein genes in penicillin-resistant clinical isolates of *Streptococcus pneumoniae*. *Proc. Natl. Acad. Sci. USA* **86**:8842–8846.
- Drlica, K., and X. Zhao. 1997. DNA gyrase, topoisomerase IV, and the 4-quinolones. *Microbiol. Mol. Biol. Rev.* **61**:377–392.
- Elting, L. S., G. P. Bodey, and B. H. Keefe. 1992. Septicemia and shock syndrome due to viridans streptococci: a case-control study predisposing factors. *Clin. Infect. Dis.* **14**:1201–1207.
- Fenoll, A., I. Jado, D. Vicioso, A. Pérez, and J. Casal. 1998. Evolution of *Streptococcus pneumoniae* serotypes and antibiotic resistance in Spain: update (1990–1996). *J. Clin. Microbiol.* **36**:3447–3454.
- Fenoll, A., J. Martínez-Suárez, R. Muñoz, J. Casal, and J. L. García. 1990. Identification of atypical strains of *Streptococcus pneumoniae* by a specific DNA probe. *Eur. J. Clin. Microbiol. Infect. Dis.* **9**:396–401.
- Fenoll, A., R. Muñoz, E. García, and A. G. de la Campa. 1994. Molecular basis of the optochin-sensitive phenotype of pneumococcus: characterization of the genes encoding the  $F_0$  complex of the *Streptococcus pneumoniae* and *Streptococcus oralis*  $H^+$ -ATPases. *Mol. Microbiol.* **12**:587–598.
- Ferrández, M. J., J. Oteo, B. Aracil, J. L. Gómez-Garcés, and A. G. de la Campa. 1999. Drug efflux and *parC* mutations are involved in fluoroquinolone resistance in viridans group streptococci. *Antimicrob. Agents Chemother.* **43**:2520–2523.
- García, P., J. L. García, E. García, and R. López. 1986. Nucleotide sequence and expression of the pneumococcal autolysin gene from its own promoter in *Escherichia coli*. *Gene* **43**:265–272.
- González, I., M. Georgiou, F. Alcaide, D. Balas, J. Liñares, and A. G. de la Campa. 1998. Fluoroquinolone resistance mutations in the *parC*, *parE*, and *gyrA* genes of clinical isolates of viridans group streptococci. *Antimicrob. Agents Chemother.* **42**:2792–2798.
- Gootz, T. D., R. Zaniewski, S. Haskell, B. Schmieder, J. Tankovic, D. Girard, P. Courvalin, and R. J. Polzer. 1996. Activity of the new fluoroquinolone trovafloxacin (cp-99,219) against DNA gyrase and topoisomerase IV mutants of *Streptococcus pneumoniae* selected in vitro. *Antimicrob. Agents Chemother.* **40**:2691–2697.
- Guillespie, S., C. Ullman, M. D. Smith, and V. Emery. 1994. Detection of *Streptococcus pneumoniae* in sputum samples by PCR. *J. Clin. Microbiol.* **32**:1308–1311.
- Hoffman, J., M. S. Cetron, M. M. Farley, W. S. Baughman, R. R. Facklam, J. A. Elliot, K. A. Deaver, and R. F. Breiman. 1995. The prevalence of drug-resistant *Streptococcus pneumoniae* in Atlanta. *N. Engl. J. Med.* **333**:481–486.
- Humbert, O., M. Prudhomme, R. Hakenbeck, C. G. Dowson, and J.-P. Claverys. 1995. Homologous recombination and mismatch repair during transformation in *Streptococcus pneumoniae*: saturation of the Hex mismatch repair system. *Proc. Natl. Acad. Sci. USA* **92**:9052–9056.
- Janoir, C., I. Podglajen, M. D. Kitzis, C. Poyart, and L. Gutmann. 1999. In vitro exchange of fluoroquinolone resistance determinants between *Streptococcus pneumoniae* and viridans streptococci and genomic organization of the *parE-parC* region in *S. mitis*. *J. Infect. Dis.* **180**:555–558.
- Janoir, C., V. Zeller, M.-D. Kitzis, N. J. Moreau, and L. Gutmann. 1996.

- High-level fluoroquinolone resistance in *Streptococcus pneumoniae* requires mutations in *parC* and *gyrA*. *Antimicrob. Agents Chemother.* **40**:2760–2764.
29. Kalin, M., K. Klancierski, M. Granstrom, and R. Mólby. 1987. Diagnosis of pneumococcal pneumonia by enzyme-linked immunosorbent assay of antibodies to pneumococcal hemolysin (pneumolysin). *J. Clin. Microbiol.* **25**:226–229.
  30. Kato, J., Y. Nishimura, R. Imamura, H. Niki, S. Higara, and H. Suzuki. 1990. New topoisomerase essential for chromosome segregation in *E. coli*. *Cell* **63**:393–404.
  31. Kikuchi, K., T. Enari, K. Totsuka, and K. Shimizu. 1995. Comparison of phenotypic characteristics, DNA-DNA hybridization results, and results with a commercial rapid biochemical and enzymatic reaction system for identification of viridans group streptococci. *J. Clin. Microbiol.* **33**:1215–1222.
  32. Liñares, J., A. G. de la Campa, and R. Pallarés. 1999. Fluoroquinolone resistance in *Streptococcus pneumoniae*. *N. Engl. J. Med.* **341**:1546–1548.
  33. Liñares, J., F. Tubau, and M. A. Domínguez. 1999. Antibiotic resistance in *Streptococcus pneumoniae* in Spain: an overview in the 1990s, p. 399–407. In A. Tomasz (ed.), *Streptococcus pneumoniae*. Molecular biology and mechanisms of disease—update for the 1990s. Mary Ann Liebert Inc., New York, N.Y.
  34. López, R., E. García, P. García, and J. L. García. 1997. The pneumococcal cell wall degrading enzymes: a modular design to create new lysins? *Microb. Drug Resist.* **3**:199–211.
  35. Luttinger, A. 1995. The twisted life of DNA in the cell: bacterial DNA topoisomerases. *Mol. Microbiol.* **15**:601–606.
  36. Muñoz, R., and A. G. de la Campa. 1996. ParC subunit of DNA topoisomerase IV of *Streptococcus pneumoniae* is a primary target of fluoroquinolones and cooperates with DNA gyrase A subunit in forming resistance phenotype. *Antimicrob. Agents Chemother.* **40**:2252–2257.
  37. Muñoz, R., E. García, and A. G. de la Campa. 1996. Quinine specifically inhibits the proteolipid subunit of the F<sub>0</sub>F<sub>1</sub> H<sup>+</sup>-ATPase of *Streptococcus pneumoniae*. *J. Bacteriol.* **178**:2455–2458.
  38. Pallarés, R., J. Liñares, M. Vadillo, C. Cabellos, F. Manresa, P. F. Viladrich, R. Martín, and F. Gudiol. 1995. Resistance to penicillin and cephalosporin and mortality from severe pneumococcal pneumonia in Barcelona, Spain. *N. Engl. J. Med.* **333**:474–480.
  39. Pan, X.-S., J. Ambler, S. Mehtar, and L. M. Fisher. 1996. Involvement of topoisomerase IV and DNA gyrase as ciprofloxacin targets in *Streptococcus pneumoniae*. *Antimicrob. Agents Chemother.* **40**:2321–2326.
  40. Pan, X.-S., and L. M. Fisher. 1996. Cloning and characterization of the *parC* and *parE* genes of *Streptococcus pneumoniae* encoding DNA topoisomerase IV: role in fluoroquinolone resistance. *J. Bacteriol.* **178**:4060–4069.
  41. Pérez-Trallero, E., J. M. García-Arenzana, J. A. Jiménez, and A. Peris. 1990. Therapeutic failure and selection of resistance to quinolones in a case of pneumococcal pneumonia treated with ciprofloxacin. *Eur. J. Clin. Microbiol. Infect. Dis.* **9**:905–906.
  42. Pozzi, G., M. R. Oggioni, and A. Tomasz. 1989. DNA probe for the identification of *Streptococcus pneumoniae*. *J. Clin. Microbiol.* **27**:370–372.
  43. Ramirez, M., E. Severina, and A. Tomasz. 1999. A high incidence of prophage carriage among natural isolates of *Streptococcus pneumoniae*. *J. Bacteriol.* **181**:3618–3625.
  44. Roberts, R. B., A. G. Krieger, N. L. Schiller, and K. C. Gross. 1979. Viridans streptococcal endocarditis: the role of various species, including pyridoxal-dependent streptococci. *Rev. Infect. Dis.* **1**:955–965.
  45. Rudolph, K. M., A. J. Parkison, C. M. Black, and L. W. Mayer. 1993. Evaluation of polymerase chain reaction for diagnosis of pneumococcal pneumonia. *J. Clin. Microbiol.* **31**:2661–2666.
  46. Spratt, B. G. 1994. Resistance to antibiotics mediated by target alterations. *Science* **264**:388–393.
  47. Sussman, J. I., J. Baron, M. J. Tenenbaum, M. H. Kaplan, R. R. Greenspan, R. R. Facklam, M. B. Tyburski, M. A. Goldman, B. F. Kanzer, and R. A. Pizzarello. 1986. Viridans streptococcal endocarditis: clinical, microbiological, and echocardiographic correlations. *J. Infect. Dis.* **154**:597–603.
  48. Taira, S., E. Jalonen, J. C. Paton, M. Sarvas, and K. Runeberg-Nyman. 1989. Production of pneumolysin, a pneumococcal toxin, in *Bacillus subtilis*. *Gene* **77**:211–218.
  49. Tankovic, J., B. Perichon, J. Duval, and P. Courvalin. 1996. Contribution of mutations in *gyrA* and *parC* genes to fluoroquinolone resistance of mutants of *Streptococcus pneumoniae* obtained in vivo and in vitro. *Antimicrob. Agents Chemother.* **40**:2505–2510.
  50. Venditti, M., P. Baiocchi, C. Barandimarte, P. Serra, G. Gentile, C. Girmenia, and P. Martino. 1989. Antimicrobial susceptibilities of *Streptococcus* species that cause septicemia in neutropenic patients. *Antimicrob. Agents Chemother.* **33**:580–582.
  51. Virolainen, A., P. Salo, J. Jero, P. Karma, J. Eskola, and M. Leinonen. 1994. Comparison of PCR assay with bacterial culture for detecting *Streptococcus pneumoniae* in middle ear fluid of children with acute otitis media. *J. Clin. Microbiol.* **32**:2667–2670.
  52. Walker, J. A., R. L. Allen, P. Falgame, M. K. Johnson, and G. J. Boulnois. 1987. Molecular cloning, characterization, and complete nucleotide sequence of the gene for pneumolysin, the sulfhydryl-activated toxin of *Streptococcus pneumoniae*. *Infect. Immun.* **55**:1184–1189.
  53. Wang, J. C. 1985. DNA topoisomerases. *Annu. Rev. Biochem.* **54**:665–697.