

# Prevalence of SHV-12 among Clinical Isolates of *Klebsiella pneumoniae* Producing Extended-Spectrum $\beta$ -Lactamases and Identification of a Novel AmpC Enzyme (CMY-8) in Southern Taiwan

JING-JOU YAN,<sup>1</sup> SHIOU-MEI WU,<sup>2</sup> SHU-HUEI TSAI,<sup>1</sup> JIUNN-JONG WU,<sup>2\*</sup> AND IH-JEN SU<sup>1</sup>

Department of Pathology,<sup>1</sup> National Cheng Kung University Medical Center, and Department of Medical Technology,<sup>2</sup> National Cheng Kung University Medical College, Tainan, Taiwan

Received 19 October 1999/Returned for modification 26 January 2000/Accepted 29 February 2000

Twenty (8.5%) of 234 nonrepetitive clinical isolates of *Klebsiella pneumoniae* from southern Taiwan were found to produce extended-spectrum  $\beta$ -lactamases (ESBLs): 10 strains produced SHV-12, 4 produced SHV-5, 2 produced a non-TEM non-SHV ESBL with a pI of 8.3, 3 produced a novel AmpC  $\beta$ -lactamase designated CMY-8 with a pI of 8.25, and 1 produced SHV-12 and an unidentified AmpC enzyme with a pI of 8.2. The CMY-8 enzyme confers a resistance phenotype similar to CMY-1 and MOX-1, and sequence comparisons showed high homologies (>95%) of nucleotide and amino acid sequences among these three enzymes. Plasmid and pulse-field gel electrophoresis analyses revealed that all isolates harboring an SHV-derived ESBL were genetically unrelated, indicating that dissemination of resistance plasmids is responsible for the spread of SHV ESBLs among *K. pneumoniae* in this area. All three isolates carrying CMY-8 had identical genotypic patterns, suggesting the presence of an epidemic strain.

Resistance to the extended-spectrum cephalosporins among members of the family *Enterobacteriaceae* has become a growing worldwide problem (1, 10, 13, 16, 21, 24). Such resistance is often associated with transferable plasmid-encoded extended-spectrum  $\beta$ -lactamases (ESBLs) (13, 24). Most ESBLs from *Escherichia coli* and *Klebsiella pneumoniae* are derived from TEM- or SHV-type  $\beta$ -lactamases by one or more amino acid substitutions which confer resistance to extended-spectrum cephalosporins (13, 24). Resistance to cephalosporins and broad-spectrum cephalosporins has also arisen in *K. pneumoniae* and *E. coli* via acquisition of plasmids containing the chromosomally encoded AmpC  $\beta$ -lactamase found in *Enterobacter* spp., *Pseudomonas aeruginosa*, and *Citrobacter* spp. (3, 4, 9, 11, 30). Unlike class A ESBLs, the activities of these enzymes are not inhibited by clavulanic acid or tazobactam (3, 4, 21). In the Far-Eastern countries, two closely related plasmid-mediated AmpC enzymes have been found in *K. pneumoniae*: MOX-1 from Japan (9) and CMY-1 from Korea (4, 14). The prevalence rates of ESBL among clinical isolates of *K. pneumoniae* in Taiwan have been reported to range from 10.7 to 30% (12, 15). SHV-5 has been shown to be the most-common ESBL (15). The present study was conducted to determine the prevalence and types of ESBLs among *K. pneumoniae* in the southern part of the country. We also describe a novel AmpC enzyme in this report.

## MATERIALS AND METHODS

**Bacterial isolates.** Between April and September 1998, a total of 234 nonrepetitive clinical isolates of *K. pneumoniae* were consecutively collected in the Department of Pathology, National Cheng Kung University Hospital, a 900-bed university hospital in southern Taiwan. All isolates were identified by using the conventional techniques (8) and/or the API 20E system (bioMérieux, Marcy l'Etoile, France).

\* Corresponding author. Mailing address: Department of Medical Technology, National Cheng Kung University Medical College, No. 1 University Rd., Tainan, Taiwan 70101, Taiwan. Phone: 886-6-2353535, ext. 5775. Fax: 886-6-2363956. E-mail: jjwu@mail.ncku.edu.tw.

**Detection of ESBL producers and susceptibility testing.** ESBL production was detected by means of double-disk synergy tests (13) and phenotypic confirmatory tests as recommended by the National Committee for Clinical Laboratory Standards (NCCLS) (20). ESBL producers detected by either of these two tests were investigated further. The MICs of antibiotics were determined by using E Test strips including ampicillin, amoxicillin-clavulanic acid, cefotaxime, ceftazidime, and imipenem (AB Biodisk, Solna, Sweden), and the results were interpreted by using the NCCLS criteria (20).

**IEF analysis and enzyme inhibition assay.** Crude preparations of  $\beta$ -lactamases (5) from clinical isolates and their transconjugants were subjected to isoelectric focusing (IEF) analysis by the method of Matthew et al. (18) with an LKB Multiphor apparatus on prepared Ampholine PAG plate gels (pH 3.5 to 9.5; Pharmacia Biotech Asia Pacific, Hong Kong, China). Gels were developed with 0.5 mM nitrocefin (Oxoid, Basingstoke, United Kingdom). Inhibition assays were carried out by overlaying gels with 0.5 mM nitrocefin and 0.3 mM cloxacillin (Sigma, St. Louis, Mo.) or 0.3 mM clavulanic acid (SmithKline Beecham Pharma, Munich, Germany) (7).

**Conjugation experiments and plasmid analysis.** Conjugation experiments were performed as described previously (26) with streptomycin-resistant *E. coli* C600 as the recipient (2). Transconjugants were selected on tryptic soy agar plates supplemented by streptomycin (500  $\mu$ g/ml; Sigma) and ceftazidime (10  $\mu$ g/ml; Glaxo, Greenford, United Kingdom), cefotaxime (10  $\mu$ g/ml; Hoechst-Roussel Pharmaceuticals Inc., Somerville, N.J.) or cefoxitin (64  $\mu$ g/ml; Sigma). ESBL production in transconjugants was confirmed by the NCCLS confirmatory test. Plasmids from clinical isolates and transconjugants were extracted by a rapid alkaline lysis procedure (29). For the restriction enzyme analysis of transconjugant plasmids, *EcoRI* and *PstI* (Roche Molecular Biochemicals, Mannheim, Germany) were used. Digested and nondigested DNA samples were analyzed by electrophoresis on 0.8% agarose gels. The plasmid sizes of transconjugants were estimated by adding up restriction fragments.

**PCR amplification and DNA sequencing.** To amplify TEM- and SHV-related genes from clinical isolates and their transconjugants, the following oligonucleotide primers were used for PCR: for *bla*<sub>TEM</sub> genes, forward primer (5'-CCCC TATTTGTTTATTTTC-3') and reverse primer (5'-GACAGTTACCAATGC TTAATCA-3') (17), corresponding to nucleotides 112 to 130 and 1074 to 1053, respectively, of Sutcliffe (28); for *bla*<sub>SHV</sub> genes, forward primer (5'-GCCGGGT TATCTTATTTGTCGC-3') and reverse primer (5'-TCTTTCCGATGCCGC CGCCAGTCA-3') (22), corresponding to nucleotides 55 to 77 and 1067 to 1044, respectively, of Mercier and Levesque (19). The primers used for amplification of *bla*<sub>CMY-1</sub>-related genes are shown in Fig. 1. PCR amplification was performed as described elsewhere (14, 17, 22) and the amplicons were totally included in these three  $\beta$ -lactamase structural genes. The amplicons were purified with PCR Clean Up kits (Roche), and both strands were sequenced with the same primers for PCR and sequencing primers on an ABI PRISM 377 Sequencer Analyzer (Applied Biosystems, Foster City, Calif.). The sequencing primers for *bla*<sub>TEM</sub> and *bla*<sub>SHV</sub> genes were synthesized as described elsewhere (17, 27), and for *bla*<sub>CMY-1</sub>-related genes are also shown in Fig. 1.

RESULTS

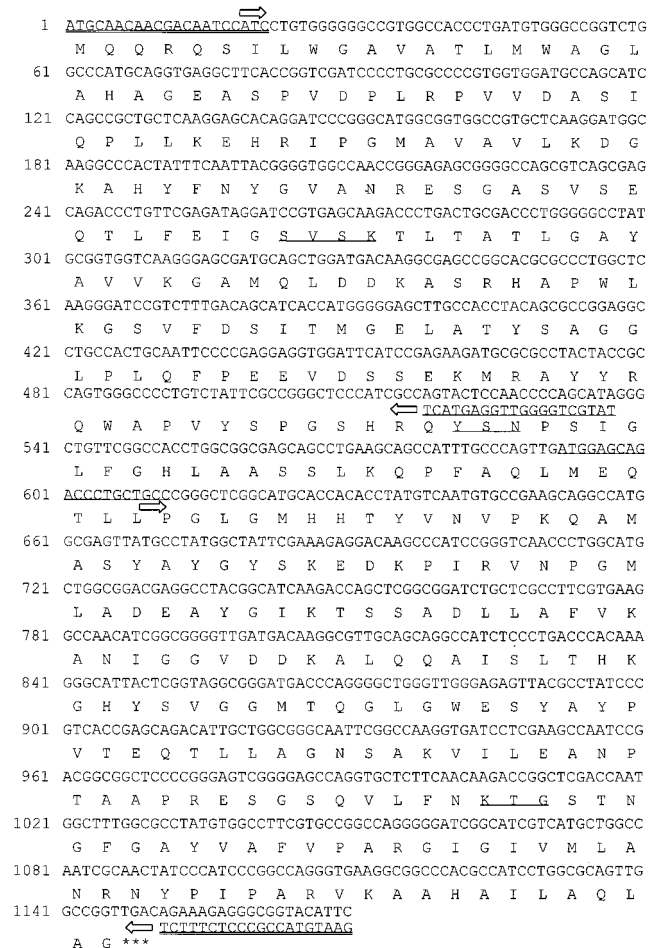


FIG. 1. Complete nucleotide sequence and predicted amino acid sequence of the *bla*<sub>CMY-8</sub> β-lactamase gene. The primers for amplification of the gene were synthesized according to the nucleotide sequence of CMY-1 (4, 14) and are double underlined. DNA sequencing was performed with a sequencing primer (underlined) and the PCR primers as well. Arrows indicate the directions of DNA sequencing. The stop codon is indicated with three asterisks. The β-lactamase active site S-V-S-K, the conserved triad K-T-G, and the class C typical motif Y-X-N are underlined.

**PFGE analysis.** Genomic DNAs prepared by the procedure of Piggot et al. (25) were digested overnight with 10 U of *Xba*I (New England Biolabs, Beverly, Mass.) and subjected to pulsed-field gel electrophoresis (PFGE) with the Pulsaphor plus system (Pharmacia) as described previously (6). DNA fragments were separated in a 1% agarose gel in 0.5× Tris-borate-EDTA (TBE) buffer at 150 V for 30 h, with pulse times ranging from 5 to 35 s.

**Nucleotide sequence accession number.** The sequence of a novel AmpC β-lactamase has been submitted to GenBank database under the accession no. AF167990.

**Prevalence of ESBL-producing isolates in *K. pneumoniae*.** Among 234 nonrepetitive clinical isolates of *K. pneumoniae*, production of ESBL was inferred in 20 (8.5%) isolates by the NCCLS confirmatory test for ESBL. The double-disk synergy test showed clavulanic acid synergy in 15 (75%) of the 20 ESBL-producing isolates.

**Resistance phenotypes.** On the basis of susceptibilities to cefotaxime, ceftazidime and cefoxitin, the ESBL producers were classified into four resistance phenotypes (Table 1). The resistance phenotype groups I and II involved high-level resistance to ceftazidime and cefotaxime, respectively. Group III involved high-level resistance to cefoxitin and cefotaxime, while group IV showed high-level resistance to cefoxitin and ceftazidime.

**IEF analysis and enzyme inhibition test.** The results of IEF are summarized in Table 2. On IEF gels, the β-lactamases with a pI of 8.25 were inhibited by cloxacillin but not by clavulanic acid, suggesting that they were AmpC enzymes. The pI 8.2 β-lactamase of the isolate in group IV, O787, was not inhibited by 0.3 mM cloxacillin and 0.3 mM clavulanic acid. All other enzymes with pIs of 5.4, 7.6, 8.2, or 8.3 were inhibited by clavulanic acid but not cloxacillin.

**Transfer of resistance.** Resistance to extended-spectrum β-lactams was successfully transferred from 17 of 20 ESBL producers to *E. coli*. The transconjugants revealed similar resistance phenotypes to their donors. The susceptibility tests of the transconjugants are summarized in Table 1. It is noteworthy that the transconjugant of isolate O787 in group IV was resistant to both cefoxitin and ceftazidime, but only the pI 8.2 β-lactamase, which was not inhibited by clavulanic acid and cloxacillin, was detected on an IEF gel. The sizes of the resistance plasmids transferred to *E. coli* are shown in Table 2.

**Sequence analysis.** The results of PCR amplification and nucleotide sequencing are summarized in Table 2. The *bla*<sub>SHV</sub> genes were amplified from all clinical isolates of *K. pneumoniae* and 12 of 17 transconjugants. Of 15 isolates producing a pI 8.2 enzyme, 4 carried an SHV-5 enzyme and 11 harbored an SHV-12 enzyme. Except for isolate O787 with group IV resistance phenotype, all isolates carrying SHV-5 or SHV-12 were distributed in group I. The SHV genotypes of the *E. coli* transconjugants were consistent with those of their donors. All clinical isolates in groups II and III carried SHV-11, which was consistent with the β-lactamase with a pI of 7.6, but their transconjugants gave a negative PCR result. The *bla*<sub>TEM</sub> genes were amplified from all 16 ESBL-producing isolates carrying a pI 5.4 β-lactamase and from none of their *E. coli* transconjugants. DNA sequencing of the amplicons showed that all of them were TEM-1.

A DNA fragment of approximately 1 kb was amplified with the primer pair specific for *bla*<sub>CMY-1</sub>-related genes from all three isolates in group III and their *E. coli* transconjugants, all

TABLE 1. MICs for *K. pneumoniae* strains with different resistance phenotypes and their transconjugants<sup>a</sup>

Resistance phenotype group	No. of clinical isolates	MIC (μg/ml) of drug for clinical isolates						No. of transconjugants	MIC (μg/ml) of drug for transconjugants					
		AMP	AMC	CAZ	CTX	FOX	IPM		AMP	AMC	CAZ	CTX	FOX	IPM
I	14	>256	4-8	≥256	3-16	3-16	0.125-0.25	11	>256	4-8	≥256	6-16	2-6	0.125-0.25
II	2	>256	4-8	2-24	128-256	8-16	0.125	2	>256	4-8	2-3	128-256	8-16	0.125
III	3	>256	16	96	>256	>256	0.25-0.5	3	>256	24	32-64	>256	>256	0.25-0.5
IV	1	>256	12	>256	48	>256	0.5	1	>256	12	>256	8	>256	0.125

<sup>a</sup> MICs were determined by the E test. Abbreviations: AMP, ampicillin; AMC, amoxicillin-clavulanic acid combination; CAZ, ceftazidime; CTX, cefotaxime; FOX, cefoxitin; IPM, imipenem.

TABLE 2. Results of IEF, genotyping of β-lactamases, plasmid analysis, and PFGE

Resistance phenotype group and strain	Clinical isolate					Transconjugant <sup>c</sup>				
	pI(s)	SHV	TEM	AmpC	Plasmid profile	PFGE pattern	pI	SHV	AmpC	Approx plasmid size (kb)
<b>Group I</b>										
X386	8.2, 7.6	5			A	I	8.2	5		100
C097	8.2, 7.6, 5.4	5	1		B	II	8.2	5		100
Q381	8.2, 7.6, 5.4	5	1		C	III	8.2	5		70
T923 <sup>a</sup>	8.2, 7.6, 5.4	5	1		D	IV				
T386	8.2, 7.6, 5.4	12	1		E	V	8.2	12		70
R436	8.2, 7.6, 5.4	12	1		E	VI	8.2	12		100
S276	8.2, 7.6, 5.4	12	1		F	VII	8.2	12		100
T986	8.2, 7.6, 5.4	12	1		G	VIII	8.2	12		70
B262 <sup>a</sup>	8.2, 7.6, 5.4	12	1		H	IX				
T255 <sup>a</sup>	8.2, 7.6, 5.4	12	1		I	X				
V293	8.2, 7.6, 5.4	12	1		G	XI	8.2	12		70
D155	8.2, 7.6	12			J	XII	8.2	12		40
B657	8.2, 7.6	12			K	XIII	8.2	12		75
C106	8.2, 7.6	12			J	XIV	8.2	12		40
<b>Group II</b>										
O574	8.3, 7.6, 5.4	11	1		L	XV	8.3			95
R549	8.3, 7.6, 5.4	11	1		M	XVI	8.3			125
<b>Group III</b>										
W142	8.25, 7.6, 5.4	11	1	CMY-8	N	XVII	8.25		CMY-8	25
W580	8.25, 7.6, 5.4	11	1	CMY-8	N	XVII	8.25		CMY-8	25
T384	8.25, 7.6, 5.4	11	1	CMY-8	N	XVII	8.25		CMY-8	25
<b>Group IV</b>										
O787	8.2, 7.6, 5.4	12	1	UD <sup>b</sup>	O	XVIII	8.2	12	UD	150

<sup>a</sup> Conjugation experiments with strains T923, B262, and T255 were not successful.

<sup>b</sup> UD, undetermined.

<sup>c</sup> All transconjugates lacked TEM.

of which expressed a pI 8.25 β-lactamase. DNA sequencing of the amplicons revealed the same nucleotide sequence among these 3 isolates. Sequence comparison with other known β-lactamases deposited in GenBank showed a close relationship to two AmpC β-lactamases, CMY-1 (accession no. X92508) and MOX-1 (accession no. D13304). The enzyme, now designated CMY-8, contains the β-lactamase active site S-V-S-K, the conserved triad K-T-G, and the class C typical motif Y-X-N on the basis of its deduced amino acid sequence (Fig. 1). Comparisons among CMY-8, CMY-1 (4) and MOX-1 (9) by using the GAP program of the Genetics Computer Group software package showed that CMY-8 shares 98.6 and 98.1% nucleotide identities, and 98.6 and 95.2% amino acid identities with CMY-1 and MOX-1, respectively. Multiple sequence alignments of the amino acid sequences of these three enzymes were performed with the PILEUP program of the Genetics Computer Group package and the result is shown in Fig. 2.

**Plasmid analysis and PFGE.** The results of plasmid and PFGE analyses are summarized in Table 2 and shown in Fig. 3. Overall, 20 clinical isolates of ESBL producers gave 15 and 18 different types of plasmid and PFGE profiles, respectively. All 14 isolates in group I showed different PFGE patterns, while all 3 isolates in group III had the same plasmid and PFGE profiles.

**DISCUSSION**

The prevalence rate of ESBL producers among clinical isolates of *K. pneumoniae* in this study was 8.5%, which is similar to those reported in many other parts of the world (13, 16, 21, 24). Among 20 ESBL producers, 15 (75%) carried SHV-de-

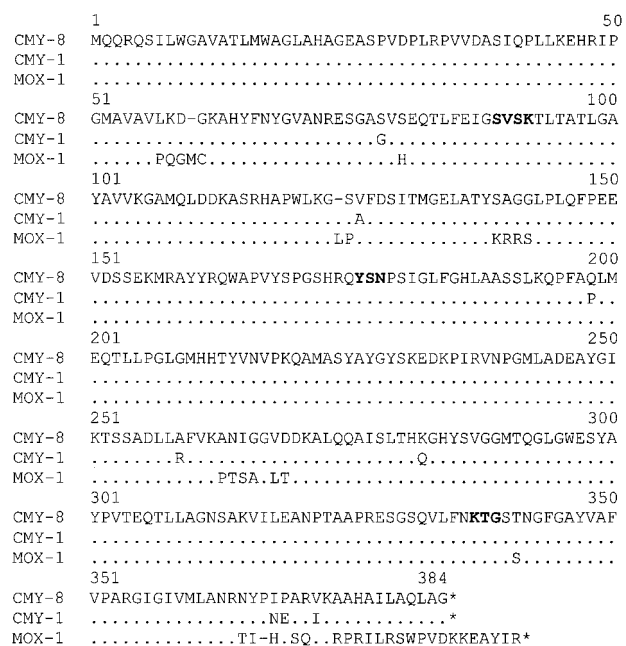


FIG. 2. Alignment of deduced amino acid sequences of CMY-8 with those of CMY-1 and MOX-1. Identical amino acids are marked with dots. The stop codon is indicated with an asterisk. The β-lactamase active site S-V-S-K, the conserved triad K-T-G, and the class C typical motif Y-X-N are shown in boldface type.



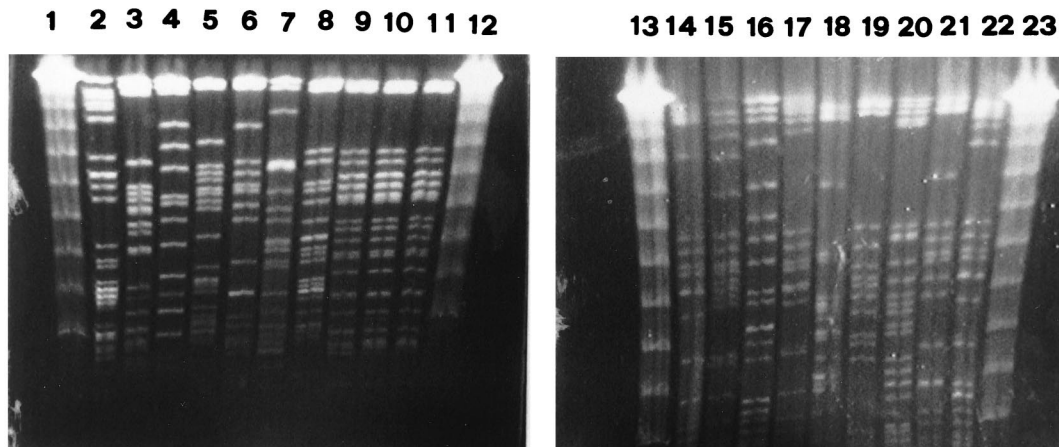


FIG. 3. PFGE of *Xba*I-digested genomic DNAs from 19 ESBL-producing *K. pneumoniae* isolates. Lanes 1, 12, 13, and 23, bacteriophage lambda DNA concatamers (GibcoBRL, Gaithersburg, Md.) which served as molecular size marker; lanes 2 to 11 and 14 to 22, isolates V293, D155, B657, C106, O574, R549, O787, W142, W580, T384, X386, C097, Q381, T923, T386, R436, T986, B262, and T255, respectively. Isolate S276 is not shown here. All ESBL producers except for three isolates carrying CMY-8 (W142, W580, and T384) had different PFGE patterns.

rived ESBLs, and 11 (55%) of them harbored SHV-12. SHV-12, identified in 1997 by Nüesch-Inderbinen et al. (23), has rarely been found in countries other than Korea (14). This report adds Taiwan to the list of countries where SHV-12 is prevalent. Another report from a district hospital in central Taiwan showed that the prevalence rate of ESBL-producing *K. pneumoniae* was 30% and that SHV-5 was the most common type of ESBL (18). The differences in the prevalence and types of ESBLs between different hospitals within a country may reflect variations in patient population, hospital care practices, and infection control activities.

The phenotype of group III involved high-level resistance to cefotaxime and ceftazidime (Table 1). The conjugation experiment and IEF analysis revealed a plasmid-mediated  $\beta$ -lactamase with a pI of 8.25 responsible for the resistance. The enzyme, designated CMY-8, could be inhibited by cloxacillin but not by clavulanic acid. Sequence analysis revealed the class C-specific amino acid motif Y-X-N and active-site serine at position 64 in the deduced amino acid sequence of the enzyme (Fig. 1). These data indicate that the enzyme is an AmpC  $\beta$ -lactamase. The alignments of the nucleotide and amino acid sequences showed that CMY-8 has a high degree of similarity with CMY-1 and MOX-1 (Fig. 1), two plasmid-mediated AmpC enzymes found in Korea (4) and Japan (9), respectively, suggesting that they might have a common origin. These three enzymes also have a similar resistance phenotype: high-level resistance to cefotaxime as well as ceftazidime.

Both isolates in group II produced  $\beta$ -lactamases with pIs of 5.4, 7.6, and 8.3 (Table 2). PCR and sequence analysis revealed nucleotide sequences identical to those of TEM-1 and SHV-11, which were consistent with the pI 5.4 and 7.6 enzymes, respectively. The resistance phenotype was transferable and their transconjugants showed no amplification of the *bla*<sub>TEM</sub> and *bla*<sub>SHV</sub> genes. Since SHV-11 and TEM-1 are not ESBLs, the pI 8.3 lactamase must be responsible for resistance to extended-spectrum  $\beta$ -lactams in both isolates and should be a non-TEM and non-SHV enzyme.

The isolate in group IV had three bands reflecting pI values of 5.4, 7.6, and 8.2 on an IEF gel. The transconjugant acquiring the pI 8.2 enzyme expressed a resistance phenotype similar to that of the isolate (Table 1). The activity of the pI 8.2  $\beta$ -lactamase could not be inhibited by cloxacillin or clavulanic acid. SHV-12 was detected in the isolate and its transconjugant by

PCR and DNA sequencing. Since SHV-12 does not confer resistance to ceftazidime (15, 23), our findings suggest that an AmpC  $\beta$ -lactamase with a pI equal to or very close to 8.2 is coexistent with SHV-12 on the same plasmid in this isolate. More studies such as molecular cloning and DNA sequencing are needed to clarify the paradoxical findings.

The *bla*<sub>TEM</sub> genes were amplified from 80% of ESBL-producing *K. pneumoniae* isolates. IEF and sequence analyses showed that all TEM enzymes in these isolates were TEM-1. Our data indicate that TEM-derived  $\beta$ -lactamases do not play an important role in resistance to extended-spectrum  $\beta$ -lactams among *K. pneumoniae* in this area despite the prevalence of TEM-1.

All 15 isolates harboring an SHV-derived ESBL revealed different PFGE profiles (Table 2), indicating that dissemination of resistance plasmids was responsible for the spreading of SHV-derived enzymes at this hospital. All three isolates carrying CMY-8 had identical PFGE and plasmid profiles (Table 2 and Fig. 3), suggesting the presence of an epidemic strain harboring CMY-8 at this hospital. Surveillance of dissemination of this epidemic strain is important because of its high-level resistance to cephalosporins as well as extended-spectrum  $\beta$ -lactams.

#### ACKNOWLEDGMENTS

We kindly thank J. Kim, Dankook University, Korea, for provision of a *K. pneumoniae* strain carrying CMY-1.

This work was partially supported by grants NCKUH89-054 from National Cheng Kung University Hospital and NSC89-2314-B-006-031 from the National Science Council, Republic of China.

#### REFERENCES

- Arlet, G., M. Rouveau, I. Casin, P. J. M. Bouvet, P. H. Lagrange, and A. Philippon. 1994. Molecular epidemiology of *Klebsiella pneumoniae* strains that produce SHV-4  $\beta$ -lactamase and which were isolated in 14 French hospitals. *J. Clin. Microbiol.* **32**:2553-2558.
- Bachmann, B. J., and K. B. Low. 1980. Linkage map of *Escherichia coli* K-12, edition 6. *Microbiol. Rev.* **44**:1451-1456.
- Bauernfeind, A., I. Stemplinger, R. Jungwirth, and H. Giamarellou. 1996. Characterization of the plasmidic  $\beta$ -lactamase CMY-2, which is responsible for cephalosporin resistance. *Antimicrob. Agents Chemother.* **40**:221-224.
- Bauernfeind, A., I. Stemplinger, R. Jungwirth, R. Wilhelm, and Y. Chong. 1996. Comparative characterization of the cephalosporinase *bla*<sub>CMY-1</sub> gene and its relationship with other  $\beta$ -lactamase genes. *Antimicrob. Agents Chemother.* **40**:1926-1930.
- Bauernfeind, A., H. Grimm, and S. Schweighart. 1990. A new plasmidic

- cefotaximase in a clinical isolate of *Escherichia coli*. *Infection* **18**:294–298.
6. **Chu, G., D. Vollrath, and R. W. Davis.** 1986. Separation of large DNA molecules by contour-clamped homogeneous electric fields. *Science* **234**:1582–1585.
  7. **Danel, F., L. M. C. Hall, D. Gur, and D. M. Livermore.** 1997. OXA-15, an extended-spectrum variant of OXA-2  $\beta$ -lactamase, isolated from a *Pseudomonas aeruginosa* strain. *Antimicrob. Agents Chemother.* **41**:785–790.
  8. **Farmer, J. J., III.** 1995. *Enterobacteriaceae*: introduction and identification, p. 438–449. *In* P. R. Murray, E. J. Baron, M. A. Tenover, and R. H. Tenover (ed.), *Manual of clinical microbiology*. American Society for Microbiology, Washington, D.C.
  9. **Horii, T., Y. Arakawa, M. Ohta, T. Sugiyama, R. Wacharotayankun, H. Ito, and N. Kato.** 1994. Characterization of a plasmid-borne and constitutively expressed *bla*<sub>MOX-1</sub> gene encoding AmpC-type  $\beta$ -lactamase. *Gene* **139**:93–98.
  10. **Jacoby, G. A., and P. Han.** 1996. Detection of extended-spectrum  $\beta$ -lactamases in clinical isolates of *Klebsiella pneumoniae* and *Escherichia coli*. *J. Clin. Microbiol.* **34**:908–911.
  11. **Jacoby, G. A., and J. Tran.** 1999. Sequence of the MIR-1  $\beta$ -lactamase gene. *Antimicrob. Agents Chemother.* **43**:1759–1760.
  12. **Jan, I. S., P. R. Hsueh, L. J. Teng, S. W. Ho, and K. T. Luh.** 1998. Antimicrobial susceptibility testing for *Klebsiella pneumoniae* isolates resistant to extended-spectrum  $\beta$ -lactam antibiotics. *J. Formos. Med. Assoc.* **97**:661–666.
  13. **Jarlier, V., M.-H. Nicolas, G. Fournier, and A. Philippon.** 1988. Extended broad-spectrum  $\beta$ -lactamases conferring transferable resistance to newer  $\beta$ -lactam agents in *Enterobacteriaceae*: hospital prevalence and susceptibility patterns. *Rev. Infect. Dis.* **10**:867–878.
  14. **Kim, J., Y. Kwon, H. Pai, J. W. Kim, and D. T. Cho.** 1998. Survey of *Klebsiella pneumoniae* strains producing extended-spectrum  $\beta$ -lactamase: prevalence of SHV-12 and SHV-2a in Korea. *J. Clin. Microbiol.* **36**:1446–1449.
  15. **Liu, P. Y. F., J. C. Tung, S. C. Ke, and S. L. Chen.** 1998. Molecular epidemiology of extended-spectrum  $\beta$ -lactamase-producing *Klebsiella pneumoniae* isolates in a district hospital in Taiwan. *J. Clin. Microbiol.* **36**:2759–2762.
  16. **Livermore, D. M., and M. Yuan.** 1996. Antibiotic resistance and production of extended-spectrum  $\beta$ -lactamases amongst *Klebsiella* spp. from intensive care units in Europe. *J. Antimicrob. Chemother.* **38**:409–424.
  17. **Mabilat, C., and S. Goussard.** 1993. PCR detection and identification of genes for extended-spectrum  $\beta$ -lactamases, p. 553–559. *In* D. H. Persing, T. F. Smith, F. C. Tenover, and T. J. White (ed.), *Diagnostic molecular microbiology: principles and applications*. American Society for Microbiology, Washington, D.C.
  18. **Matthew, M., M. Harris, M. J. Marshall, and G. W. Rose.** 1975. The use of analytical isoelectric focusing for detection and identification of  $\beta$ -lactamases. *J. Gen. Microbiol.* **88**:169–178.
  19. **Mercier, J., and R. C. Levesque.** 1990. Cloning of SHV-2, OHIO-1, and OXA-6  $\beta$ -lactamase. *Antimicrob. Agents Chemother.* **34**:1577–1583.
  20. **National Committee for Clinical Laboratory Standards.** 1999. Performance standards for antimicrobial susceptibility testing: ninth informational supplement. M100-S9. National Committee for Clinical Laboratory Standards, Wayne, Pa.
  21. **Nordmann, P.** 1998. Trends in  $\beta$ -lactam resistance among *Enterobacteriaceae*. *Clin. Infect. Dis.* **27**(Suppl. 1):S100–S106.
  22. **Nüesch-Inderbinen, M. T., H. Hächler, and F. H. Kayser.** 1996. Detection of genes coding for extended-spectrum SHV beta-lactamases in clinical isolates by a molecular genetic method, and comparison with the E test. *Eur. J. Clin. Microbiol. Infect. Dis.* **15**:398–402.
  23. **Nüesch-Inderbinen, M. T., F. H. Kayser, and H. Hächler.** 1997. Survey and molecular genetics of SHV  $\beta$ -lactamase in *Enterobacteriaceae* in Switzerland: two novel enzymes, SHV-11 and SHV-12. *Antimicrob. Agents Chemother.* **41**:943–949.
  24. **Philippon, A., G. Arlet, and P. H. Lagrange.** 1994. Origin and impact of plasmid-mediated extended-spectrum beta-lactamases. *Eur. J. Clin. Microbiol. Infect. Dis.* **13**(Suppl. 1):S17–S29.
  25. **Piggot, P. J., M. Amjad, J. J. Wu, H. Sandoval, and J. Castro.** 1990. Genetic and physical maps of *Bacillus subtilis* 168, p. 493–543. *In* C. R. Harwood and S. M. Cutting (ed.), *Molecular biology methods for bacillus*. John Wiley & Sons Ltd., West Sussex, England.
  26. **Provence, D. L., and R. Curtiss III.** 1994. Gene transfer in gram-negative bacteria, p. 319–347. *In* P. Gerhardt, R. G. E. Murray, W. A. Wood, and N. R. Krieg (ed.), *Methods for general and molecular bacteriology*. American Society for Microbiology, Washington, D.C.
  27. **Rasheed, J. K., B. Metchock, F. Berkowitz, L. Weigel, J. Crellin, C. Steward, B. Hill, A. A. Medeiros, and F. C. Tenover.** 1997. Evolution of extended-spectrum  $\beta$ -lactam resistance (SHV-8) in a strain of *Escherichia coli* during multiple episodes of bacteremia. *Antimicrob. Agents Chemother.* **41**:647–653.
  28. **Sutcliffe, J. G.** 1978. Nucleotide sequence of the ampicillin resistance gene of *Escherichia coli* plasmid pBR322. *Proc. Natl. Acad. Sci. USA* **75**:3737–3741.
  29. **Takahashi, S., and Y. Nagano.** 1984. Rapid procedure for isolation of plasmid DNA and application to epidemiological analysis. *J. Clin. Microbiol.* **20**:608–613.
  30. **Tzouveleki, L. S., E. Tzelepi, and A. F. Mentis.** 1994. Nucleotide sequence of a plasmid-mediated cephalosporinase gene (*bla*<sub>LAT-1</sub>) found in *Klebsiella pneumoniae*. *Antimicrob. Agents Chemother.* **38**:2207–2209.