

# Identification of a Plasmid-Borne Chloramphenicol-Florfenicol Resistance Gene in *Staphylococcus sciuri*

STEFAN SCHWARZ,<sup>1\*</sup> CHRISTIANE WERCKENTHIN,<sup>1,2</sup>  
AND CORINNA KEHRENBURG<sup>1</sup>

Institut für Tierzucht und Tierverhalten der Bundesforschungsanstalt für Landwirtschaft Braunschweig (FAL),  
29223 Celle,<sup>1</sup> and Institut für Medizinische Mikrobiologie, Infektions- und Seuchenmedizin,  
Tierärztliche Fakultät der Ludwig-Maximilians-Universität München,  
80539 Munich,<sup>2</sup> Germany

Received 18 January 2000/Returned for modification 23 March 2000/Accepted 14 June 2000

**The 16.5-kbp plasmid pSCFS1 from *Staphylococcus sciuri* mediated combined resistance to chloramphenicol and florfenicol. The gene responsible for this resistance property, *cfr*, was cloned and sequenced. The amino acid sequence of the Cfr protein revealed no homology to known acetyltransferases or efflux proteins involved in chloramphenicol and/or florfenicol resistance or to other proteins whose functions are known.**

*Staphylococcus sciuri* is a common inhabitant of the physiological skin flora of most rodents, ungulates, carnivora, and marsupials. Although classified as rarely pathogenic (6), *S. sciuri* isolates have been obtained occasionally from cases of mastitis in goats (10) and bronchopneumonia in cattle (13). Antimicrobial resistance is common among *S. sciuri* isolates, and a number of plasmids carrying one or more resistance genes have been identified (11, 13, 14). Resistance to chloramphenicol (CM) in staphylococci has usually been associated with plasmid-borne *cat* genes (11, 13), whose gene products inactivate CM by diacetylation. CM acetyltransferases, however, are unable to inactivate florfenicol (FF), a fluorinated CM derivative which was licensed in Germany in 1995 as a therapeutic agent to control bacterial respiratory infections in cattle. Genes whose gene products mediate combined resistance to CM and FF by efflux of both drugs have been identified in gram-negative bacteria, such as *Salmonella enterica* serovar Typhimurium (2) and *Photobacterium damsela* subsp. *piscicida*, formerly known as *Pasteurella piscicidae* (5). In staphylococci and related organisms, FF resistance genes have not been described yet.

An *S. sciuri* isolate obtained from the nasal swab of a calf suffering from an infection of the respiratory tract proved to be resistant to tetracycline, erythromycin, kanamycin, CM, and FF. Plasmid analysis revealed the presence of six plasmids in the size range between 1.5 and 16.5 kbp. Experiments involving transformation into protoplasts of *Staphylococcus aureus* RN4220 (12) and subsequent selection of the transformants on regeneration media containing 20 µg of FF/ml (Essex, Munich, Germany) identified only the 16.5-kbp plasmid, designated pSCFS1, as the mediator of resistance to CM and FF. This plasmid also mediated resistance to erythromycin by an inducibly expressed *ermC* gene as confirmed by PCR analysis (7). Cloning experiments revealed that the *ermC* gene was located on a 2.5-kbp *PstI* fragment of pSCFS1 (data not shown). The original *S. sciuri* isolate and *S. aureus* RN4220:pSCFS1 showed FF MICs of 64 µg/ml and CM MICs of 32 µg/ml. Preincubation of these isolates in the presence of either 0.5 µg of FF or

0.5 µg of CM increased the FF MICs to 512 µg/ml and the CM MICs to 64 µg/ml, suggesting that pSCFS1-mediated resistance to FF and CM in both staphylococcal hosts is inducible by FF as well as CM. Plasmid pSCFS1 was mapped (Fig. 1) and subjected to cloning experiments. Restriction fragments of pSCFS1 generated by the enzymes *EcoRI* and *BclI-BamHI* were cloned into pBluescript SKII<sup>+</sup>. The recombinant plasmids were transformed into the recipient strain *Escherichia coli* HB101 and plated on Luria-Bertani (LB) agar supplemented with 20 µg of FF/ml. Only *E. coli* HB101 clones which carried a 3.8-kbp *EcoRI* fragment of pSCFS1 (Fig. 1) grew on these selective plates. Subclones of this *EcoRI* fragment were produced and tested for their ability to grow on LB agar supplemented with 20 µg of FF/ml (Fig. 1). Subclones which carried a 3-kbp *ClaI-EcoRI* fragment, a 2.9-kbp *EcoRI-XbaI* fragment, or a 2-kbp *ClaI-XbaI* fragment grew on this selective medium and also on LB agar supplemented with 15 µg of CM/ml. The MICs of FF and CM for these subclones were 32 µg/ml; preincubation in the presence of subinhibitory concentrations of FF or CM increased the FF MICs to 64 µg/ml but had no effect on the CM MICs. A lack of increase in CM MICs has also been observed when inducible *cat* genes from *Staphylococcus* spp. were expressed in *E. coli* hosts (15). All subclones generated by *BamHI* digestion, e.g., those carrying 1.1-kbp *ClaI-BamHI* and 0.95-kbp *BamHI-XbaI* fragments (Fig. 1), failed to exhibit resistance to FF and CM.

The sequence of the smallest restriction fragment that conferred resistance to FF and CM, the 2,037-bp *ClaI-XbaI* fragment, was determined on both strands. Three open reading frames (ORFs) were detected. The *BamHI* site was located within an ORF for a peptide of 349 amino acids (aa) (positions 570 to 1619). This reading frame, designated *cfr* (CM and FF resistance) was followed by a pair of inverted repeated sequences of 13 bp, which may represent the transcriptional terminator. The *cfr* reading frame was preceded by a potential promoter structure (–35: TTTACA, positions 168 to 173; –10: TTACAG, positions 190 to 195; A, position 204) and two overlapping reading frames, ORF1 (positions 237 to 416) and ORF2 (positions 371 to 505), coding for putative peptides of 59 and 44 aa, respectively. The amino acid sequences encoded by both small ORFs did not exhibit significant homology to protein sequences deposited in the databases. Deletion of the *cfr* gene upstream region as shown in the *HpaI-XbaI* and the *MspI-XbaI* subclones (Fig. 1) resulted in sensitivity to FF and

\* Corresponding author. Mailing address: Institut für Tierzucht und Tierverhalten der Bundesforschungsanstalt für Landwirtschaft Braunschweig (FAL), Dörnbergstr. 25-27, 29223 Celle, Germany. Phone: 49-5141-384673 or -384675. Fax: 49-5141-381849. E-mail: schwarz@ktf.fal.de.

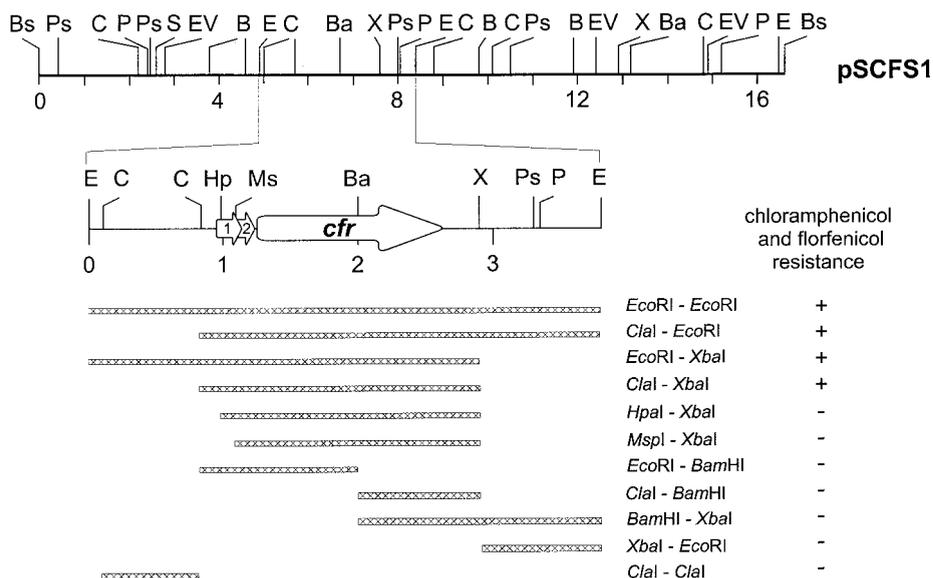


FIG. 1. Restriction map of plasmid pSCFS1 from *S. sciuri* and subcloning strategy for obtaining restriction fragments which mediate (or do not mediate) resistance to CM and FF. Restriction enzyme abbreviations: B, *Bcl*I; Ba, *Bam*HI; Bs, *Bst*EII; C, *Clal*; E, *Eco*RI; EV, *Eco*RV; Hp, *Hpa*I; Ms, *Msp*I; P, *Pvu*II; Ps, *Pst*I; S, *Sac*I; X, *Xba*I. A distance scale in kilobase pairs is given below each map. Arrows, locations of the *cfr* reading frames, ORF1 and ORF2, and their directions of transcription.

CM, suggesting that this region is essential for the expression of combined resistance to FF and CM. Further analysis of the upstream region revealed similarities to the upstream regions of inducible *cat* genes from *Staphylococcus* and *Bacillus* spp. (8, 16). The region between the stop codon of ORF2 and the start codon of *cfr* comprised a pair of inverted repeated sequences (IR1: positions 515 to 527; IR2: positions 549 to 563) which might be able to form a stable mRNA secondary structure ( $\Delta G = -60.3$  kJ/mol). The *cfr*-associated ribosome binding site was located within the IR2 sequence. Moreover, the terminal part of ORF2 (5'-GTGCAAAAAGAAATTGATTCT-3') showed considerable homology to previously identified ribosome stall sequences in the reading frames of the regulatory peptides involved in inducible CM resistance (8, 16). A ribosome stalled in the terminal part of ORF2 will overlap the IR1 sequence and abolish mRNA secondary structure formation, thus rendering the *cfr*-associated ribosome binding site accessible to ribosomes and allowing translation of the *cfr* transcripts. Assuming that inducible expression of *cfr* occurs via a translational attenuation-like process (8, 16), deletion of the upstream region which comprises relevant elements for such a regulatory system may explain the loss of resistance to FF and CM.

Comparison of the Cfr amino acid sequence as deduced from the nucleotide sequence revealed no homology to acetyltransferases or efflux proteins (2, 5, 9) so far known to be associated with resistance to FF and/or CM. However, homology to a number of proteins from a wide variety of bacteria, including *Mycobacterium tuberculosis* H37RV (accession no. Q10806), *Treponema pallidum* (accession no. AAC65061), *Haemophilus influenzae* Rd (accession no. P44665), *Pseudomonas aeruginosa* PAO1 (accession no. Q51385), *E. coli* K12 (accession no. P36979), *Bacillus subtilis* 16 (accession no. CAA74265), the soil bacterium *Streptomyces coelicolor* A3(2) (accession no. CAA19907), the cyanobacterium *Synechocystis* sp. strain PCC6803 (accession no. Q55880), and the archaeobacterium *Thermotoga maritima* MSB8 (accession no. AAD36781) was detected (Fig. 2). The reading frames encoding most of these proteins were identified during whole-

genome sequencing of the respective organisms. These proteins have some properties in common: they exhibit similar sizes of 340 to 390 aa, have no known functions, and do not exhibit any specific features such as ATP binding domains which might point to their possible functions. Recently, the terminal 133 aa of a protein from *S. aureus* (accession no. CAB60749) which shows 53% homology to the Cfr protein have been reported (3). This protein was assumed to be an auxiliary protein which might play a role in the expression of methicillin resistance (3). Analysis of the Cfr protein sequence confirmed the lack of ATP binding domains (1). Use of the TMpred program ([http://www.ch.embnet.org/software/TMPRED\\_form.html](http://www.ch.embnet.org/software/TMPRED_form.html)) did not result in the detection of any topology typical for transmembrane proteins. This observation suggested that the Cfr protein is unlikely to be secreted or anchored to the membrane (4). Moreover, the negative results of a CM acetyltransferase assay and a bioassay to demonstrate the enzymatic inactivation of FF and CM (12) confirmed that neither the original *S. sciuri* nor the *S. aureus* RN4220:pSCFS1 transformant was resistant to FF and CM by enzymatic inactivation of the drugs. Even though the mechanism of Cfr-mediated FF and CM resistance remains to be elucidated, these observations indicate that the *cfr* gene represents a novel type of transferable CM-FF resistance gene, the product of which confers resistance to both drugs not only in staphylococci but also in *E. coli* and obviously is not associated with any of the so far known mechanisms of FF and CM resistance.

**Nucleotide sequence accession number.** The nucleotide sequence of the *cfr* gene and its adjacent regions has been submitted to the EMBL database and was assigned accession no. AJ249217.

C.K. received a scholarship from the Gesellschaft der Freunde der FAL (GdF). This study was supported by a grant from the Deutsche Forschungsgemeinschaft (SCHW 382/6-1).

We thank Georg Wolf for providing the *S. sciuri* isolate, Keith G. H. Dyke for helpful discussions, and B. Otto for help with sequence analysis.

M. tuberculosis	MVPELMFDEPRPGRPPRLADLDAAGRASAVAEGLPAPFAKOLA	45
S. coelicolor	MPKPGELTFVAPRGVKKPPRLADLTPAERKEAVAAGKPKFRAKOLS	48
E. coli	MSEQLVTPENVTTKDGKINLLDLNRQOMREFFKDLGKPKFRADQVM	46
H. influenzae	MCNNEAKMSSELLSVQSDAPAKKINLMDLTRQOMREFFKELGKPKFRADQV	51
P. aeruginosa	MTTITAGKVNLLGLTQPLEQFFESI GKRFRAFQVM	37
Synechocystis	MAPSPAPLLSLSLPELTDVWVQTGQPAYRKGKTH	34
T. maritima	MKNLLDLSEYELVTEITNLGLERYRADOIL	30
B. subtilis	MAELNKTVKRKLRTERPSTIYSEELDEIKQWLLDNGEKPKFRADQIF	46
S. sciuri	MNFNNTKYGKIQEELRSNNEPDYRIKQIT	30
T. pallidum	MEWCCALSGLLPEEIQKVCFAERREGVVVF	31
M. tuberculosis	HQYVGRLIADPRQMTDLPAAVRRDIAGAMFPNLLTASADITCDAGQTRKTL	96
S. coelicolor	QHYFARYAHAPEQWTDIPAGSREGLREALLPELMTVVRHLSTDQGTTRKTL	99
E. coli	KWMYHYCCDNFDEMTDINKVLRGKLEKVAEIRAPEV.VEEQRS SDGTI KWA	96
H. influenzae	KWIYHFGEDNPDNMTNINKKLRKLEKAVAEIKAPEVAVE. QRSADGTI KWA	101
P. aeruginosa	KWIHHFGVDDFDAMTNVGKALREKLEKASAEIRGPEI.VSQDISADGTRKVV	87
Synechocystis	QWLYQKGARSLTAMTDLPKVVRE. .KNVHYPIGRSVIDHCAPADHTRKYL	83
T. maritima	DWVFDKKNVNFDEMTNLSKKHRRALLKHEHFSISFLKLLDKKVSRI DGTTRKFL	81
B. subtilis	EWLYEKRVSSFDMTNLSKDLREKLNTRFVLTTLKTAVK. QTSQDGTTRKFL	96
S. sciuri	NAIFKQRISRFEDMKVLPKLLREDDLNNFGETVLNKKLAEQNSEQVTRKVL	81
T. pallidum	RWIAAGCTD.FHAMSDLSSETRARLARACVISTRV.YTTLRDVDGTLKELG	80
M. tuberculosis	WRAVDGTMFESVLMRYPRRN.TVCISSOASCGMACFFCATGGQGLTRNIST	146
S. coelicolor	WKLFDGTLVSVLMRYPDRV.TMCISSOASCGMNCPFCATGGQAGLDRNIST	149
E. coli	IAVGQDR.VETVYIPEADRA.TLQVSSOVGCALCEKFCSTAAQQGFNRNRIV	145
H. influenzae	MQVGEQQ.VETVYIPEADRA.TLQVSSOVGCALACTFCSTAAQQGFNRNRITV	150
P. aeruginosa	VRVASGSCVEVYIPEQGGG.TLQVSSOAGCALDCSFCSTGKGFNSDITATA	137
Synechocystis	LRLADGLIIEVYIPESSKRL.TVQVSSOVGCAMDCNFCATGKGGFIRNIES	133
T. maritima	WELEDGNTIESVMLFHPDRI.TAGISTOVGCEPVKICFCATGMSGFVRNIST	131
B. subtilis	FELHDGYTIEVLMRHEYGN.SVCVTTQVGCRIQCTFCASTLGLKRNIEV	146
S. sciuri	FELVKNERVEVNMKYKAGWESFCISSOCNFGCKFCATGDI GELKNTIEV	132
T. pallidum	IELKDKRRVBAVLLVDQVSRKTALSCOVGCPMACAFCCTGGQLSFARNISA	131
M. tuberculosis	AEILBOVRAGAALRDDFDG...RLSNVFMGMGEBELANAYARVLAAVQRI	193
S. coelicolor	AEIVHOIVDGMRALRDGEVPGGPARLSNI VFMGMGEBELANYNRVVGAIRRL	200
E. coli	SEIIGOVWRRAKIVGAAKVVTQQR.PI TNVVMGMGEBELNLNNVV PAMEIM	195
H. influenzae	SEIIGOVWRASKIIGNFVGTQVR.PI TNVVMGMGEBELNINVANVVPAMEIM	200
P. aeruginosa	AEVIQOVWIANKSPGTVPKIDR.AT TNVVMGMGEBELNFDNVVAAMNIM	187
Synechocystis	HEIVDOVLTQVEEF.....HERVSNVFMGMGEBELNLPQVVKAVRECL	176
T. maritima	GEIVAOIILSMEKEKK.....KIGNVVMGMGEBELNINVENTKSRIRL	174
B. subtilis	GEIVAOVVKVQKAL.....DETERVSSVVMGIEPFDNFEMMLAFLKII	192
S. sciuri	DEITDOVLYFHLLGHQ.....IDSI SFMGMGEBELANRQVFLDSDFT	174
T. pallidum	SEIVBQFLHLERCVT.....LDNVVFMGMGEBELNLDVACRAIETL	173
M. tuberculosis	TARPPSGFGISARAVTVSTVGLAPATRNLADARLGVTLALSLHAPDDGLRD	244
S. coelicolor	TDPEPDGLGLQRGITVSTVGLVPAHRFTGEGFKCLRAISLHAPDDELRD	251
E. coli	LDDF..GFGLSKRRVTLSTSGVVPALDKL.GDMIDVALAISLHAPNDEIRD	243
H. influenzae	LDDF..AYGLSKRRVTLSTSGVVPALDNL.SKMIDVALAISLHAPNDEIRD	248
P. aeruginosa	MDDL..GYGISKRRVTLSTSGVVMIDDKL.GEVIDVSLAISLHAPNDELRN	235
Synechocystis	NQVV...GIGQRALTISTVGLPGKIRQLADRHLQVTFAVSLHAPNQTLRQ	223
T. maritima	NHKKM..GNIGIRRTISTVGI PDRIQLAEGLDKLALSLHAPNTRKRD	223
B. subtilis	NHDK..GLNIGARHITVSTSGIIPKIYEFADQOMQINFALISLHAPNTEIRS	241
S. sciuri	DPNL...FALSRRLSHSTIGIIPSTKKTQEYQVNLTFSLHSPYSEBRS	222
T. pallidum	SHPO..GRDLSKRITISTSGHCRGITYSLADRALQVRLAVSHTTANAPIRA	222
M. tuberculosis	TLVPVNNRWRVSEALDAARYYANVTG...RRVSTIEVALIRDVNDQ PWRADL	292
S. coelicolor	TLVPVNRWKVREVLDAAGFEYAAKSG...RRLSIEYALIRDINDQAWRGDR	299
E. coli	EIVPENKKYNIETFLAARRYLEKSNANQGRVTEIYVMDHVNVDGVEHAHQ	294
H. influenzae	EIVPENKKYNIETFLSDVNRVNLVSNANHGKVTIEYVMDHVNVDGVEHAHQ	299
P. aeruginosa	KLVPENKKYPLGMLLDACRRYISRLGKVR.VLTVEYTLKLDVNDQVEHAHQ	285
Synechocystis	SLIENARHYPLEQLLADCRAYVETG...RRVTFEYVLLAGVNDQVPHAEQ	271
T. maritima	QLVPELNKKYSTIEILNAVKIYQRKTG...NRVTEYVLLRGINDEISDAKK	271
B. subtilis	RLMPEINRAYKLPDLMEAVKYYINKTG...RRISFEYGLFGGVNDQVEHAEE	289
S. sciuri	KLMPENDRYPIDEVNMLDDEHIRTLS...RKVYIAYIMLPGVNDSLEHANE	279
T. pallidum	RLMPEAAHDSLAKLKSARVYNEKSG...KRVTELEALMRGVNTERHAQE	270
M. tuberculosis	LGKRLHRVL..GPLAHVNLLEPLNPTPGSDWDASPKP..VEREYKRVRAKG	339
S. coelicolor	LGRLLR...GRPVHNLLEPLNPTPGSKWTASRPE..DEKAEVEAIAAHG	343
E. coli	LAELLK...DTPCKINLLEPWNFFPGAPYGRSSNS..RIDREYKVLMSYG	338
H. influenzae	LAEVLK...NTPCKINLLEPWNFFPEAPYAKSSNT..RIDREYKTLMEYD	343
P. aeruginosa	MIALLK...DTPCKINLLEPWNFFPHSGVERPSNN..AIRREYQDMLHKG	329
Synechocystis	LAQKLR...GFQTHVNLLEPYNFISEVDYQRETEA..QINQEAQVLSDHR	315
T. maritima	LAELLR...NMKVFNLEPVEVETVEGLR.REPSRE..RLLTEKRI LLENG	314
B. subtilis	LADLLE...GVKCHVNLLEPVNYPEDVYVTRPD..QIFAEKTLKSRG	333
S. sciuri	VVSLKSRYSKGLYHVNLEPYNPTISAPEMYGEANEGQVEAYKVLKSAG	321
T. pallidum	VIDFAH...GLNVHVNLEPWNPEVASIHFEETPREVE..VAHEEALLMRAR	314
M. tuberculosis	VSVTRDRTRGREISAACGQLAAVGG	364
S. coelicolor	VPVTRDRTRGQEDIDGACGQLAASER	368
E. coli	FTTIVRKTTRGDDIDAAAGDVIDRTRKTLRKRMQGEAIDIKAV	384
H. influenzae	FTVIIIRKTTRGDDIDAAAGDVIDRTRKTKTAMKROFGQNGIVTEVN	390
P. aeruginosa	ENVTVRTRTRGDDIDAAAGDVGQVMDRTRRSERYIAVRQLAESESAANRN	379
Synechocystis	IAVSVRYSRQVQDAACGQLRASRKBELAE LTPMA	350
T. maritima	IEAETTRREKQTDIEAACGQLRLKRIKRS	343
B. subtilis	VNVTTRREQHIDDAACGQLRAKERQDETR	363
S. sciuri	IHTVTRRSQFGIDDAACGQLYGNYSQ	349
T. pallidum	IFVTRRYSRQNGIGGACGQLSKTAGV	340

FIG. 2. Amino acid alignment of the Cfr protein from *S. sciuri* with similar proteins from *M. tuberculosis* H37RV, *S. coelicolor* A3(2), *E. coli* K12, *H. influenzae* Rd, *P. aeruginosa* PAO1, *Synechocystis* sp. strain PCC6803, *T. maritima* MSB8, *B. subtilis* 16, and *T. pallidum* produced with the DNAMAN sequence analysis software (Lynn BioSoft, Vaudreuil, Quebec, Canada). Black boxes, identical amino acids; gray boxes, homologous amino acids which are present in at least 40% of the aligned sequences.

## REFERENCES

1. Allignet, J., V. Loncle, and N. El Solh. 1992. Sequence of a staphylococcal plasmid gene, *vga*, encoding a putative ATP-binding protein involved in resistance to virginiamycin A-like antibiotics. *Gene* **117**:45–51.
2. Arcangioli, M. A., S. Leroy-Setrin, J. L. Martel, and E. Chaslus-Dancla. 1999. A new chloramphenicol and florfenicol resistance gene linked to an integron structure in *Salmonella typhimurium* DT104. *FEMS Microbiol. Lett.* **174**:327–332.
3. De Lencastre, H., S. W. Wu, M. G. Pinho, A. M. Ludovice, S. Filipe, S. Gardete, R. Sobral, S. Gill, M. Chung, and A. Tomasz. 1999. Antibiotic resistance as a stress response: complete sequencing of a large number of chromosomal loci in *Staphylococcus aureus* strain COL that impact on the expression of resistance to methicillin. *Microb. Drug Resist.* **5**:163–175.
4. Foulger, D., and J. Errington. 1998. A 28 kbp segment from the *spoVM* region of *Bacillus subtilis* 168 genome. *Microbiology* **144**:801–805.
5. Kim, E., and T. Aoki. 1996. Sequence analysis of the florfenicol resistance gene encoded in the transferable R plasmid from a fish pathogen, *Pasteurella piscicida*. *Microbiol. Immunol.* **40**:665–669.
6. Kloos, W. E., and K. H. Schleifer. 1986. Genus IV. *Staphylococcus*, p. 1013–1035. In P. H. A. Sneath, N. S. Mair, M. E. Sharpe, and J. G. Holt (ed.), *Bergey's manual of systematic bacteriology*, vol. 2. Williams & Wilkins, Baltimore, Md.
7. Lodder, G., C. Werckenthin, S. Schwarz, and K. G. H. Dyke. 1997. Molecular analysis of naturally occurring *ermC*-encoding plasmids in staphylococci isolated from animals with and without previous contact with macrolide/lincosamide antibiotics. *FEMS Immunol. Med. Microbiol.* **18**:7–15.
8. Lovett, P. S. 1990. Translational attenuation as the regulator of inducible *cat* genes. *J. Bacteriol.* **172**:1–6.
9. Murray, I. A., and W. V. Shaw. 1997. *O*-Acetyltransferases for chloramphenicol and other natural products. *Antimicrob. Agents Chemother.* **41**:1–6.
10. Poutrel, B. 1984. Udder infection of goats by coagulase-negative staphylococci. *Vet. Microbiol.* **9**:131–137.
11. Schwarz, S., M. Cardoso, and H. Blobel. 1990. Detection of a novel chloramphenicol resistance plasmid from “equine” *Staphylococcus sciuri*. *J. Vet. Med. B* **37**:674–679.
12. Schwarz, S., M. Cardoso, and H. Blobel. 1990. Plasmid-mediated chloramphenicol resistance in *Staphylococcus hyicus*. *J. Gen. Microbiol.* **135**:3329–3336.
13. Schwarz, S., and S. Grözl-Krug. 1991. A chloramphenicol/streptomycin-resistance plasmid from a clinical strain of *Staphylococcus sciuri* and its structural relationships to other staphylococcal resistance plasmids. *FEMS Microbiol. Lett.* **82**:319–322.
14. Schwarz, S., and W. C. Noble. 1994. Tetracycline resistance genes in staphylococci from the skin of pigs. *J. Appl. Bacteriol.* **76**:320–326.
15. Schwarz, S., U. Spies, and M. Cardoso. 1991. Cloning and sequence analysis of a plasmid-encoded chloramphenicol acetyltransferase gene from *Staphylococcus intermedius*. *J. Gen. Microbiol.* **137**:977–981.
16. Stokes, H. W., and R. M. Hall. 1991. Sequence analysis of the inducible chloramphenicol resistance determinant in the Tn1696 integron suggests regulation by translational attenuation. *Plasmid* **26**:10–19.