

Staphylococcus aureus Mutants Selected by BMS-284756

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BMS-284756, a novel des-fluoro(6)-quinolone, was used to select for in vitro mutants of Staphylococcus aureus ISP794. Step mutants were obtained, and the quinolone resistance-determining regions of four target genes, gyrA, gyrB, grlA, and grlB, were sequenced. The data suggest that DNA gyrase is the primary target for BMS-284756 in S. aureus.

BMS-284756 is a novel des-fluoro(6)-quinolone which has a fluorine incorporated through a C₈ difluoromethyl ether linkage instead of the classical C₆ fluorine of fluoroquinolones (1, 3). When compared to five fluoroquinolones (trovafloxacin, moxifloxacin, levofloxacin, ofloxacin, and ciprofloxacin), BMS-284756 was the most active against staphylococci, streptococci, pneumococci, and Enterococcus faecalis (1).

S. aureus ISP794, a group II wild-type strain of NCTC8325 (pig-131), was obtained from David C. Hooper (Massachusetts General Hospital, Boston, Mass.) and was used for the selection of spontaneous step mutants by both BMS-284756 and ciprofloxacin (6). Mutants were isolated from Mueller-Hinton agar plates containing two, four and eight times the MIC determined for the preceding step strain (4). The frequency of resistance emergence (FRE) was calculated by dividing the number of mutants obtained at the highest concentration of drug on which resistant colonies of bacteria emerged in each round of selection by the number of cells plated. Each FRE experiment was repeated twice. All MICs were determined according to the National Committee for Clinical Laboratory Standards microdilution standard protocol (5). The quinolone resistance-determining regions (QRDRs) of the parental and mutant strains were sequenced.

Two microliters of a culture with an optical density at 550 nm of 0.2 was directly lysed in a standard PCR, or isolated chromosomal DNA was used as the template to obtain PCR fragments for sequencing. The following primers were used to amplify the QRDR of each target gene: grlA, 5' primer, ACT TGAAGATGTTTTAGGTGAT; and 3' primer, TTAGGAA ATCTTGATGGCAA; grlB, 5' primer, AGACAAATTGCCA TTCTATTTAGAAG; and 3' primer, AAACCTTTGTAACG TTGTAACG; gyrA, 5' primer, TATTACCAGTGAAATGCG TRAATC; and 3' primer, ACGAGAACGCATTTGAATTG AACG; and gyrB, 5' primer, TAGACTTTCTGGTGAAGAT ACACG; and 3' primer, ATTTTGGTGTGGATTCAATT CAG. PCRs were purified for sequencing using the QIAquick PCR Purification Kit (Qiagen, Valencia, Calif.) according to the manufacturer's instructions. Internal primers as well as PCR primers were used to sequence the DNA templates on an ABI Prism 3700 capillary sequencing machine (PE Applied Biosystems, Norwalk, Conn.). Sequencing traces were

analyzed using the Lasergene software program. Any mutation was confirmed by sequencing an independently obtained PCR fragment. The mutant strain designation reflects the following information: mutant step, the antibiotic used for mutant selection, and the number of the mutant (Fig. 1).

In all mutants selected by BMS-284756, for which the MIC was ≥ 0.06 $\mu\text{g/ml}$, the GyrA 84 (Ser→Leu) mutation was present (Tables 1 and 2). In the case of fourth-step BMS-284756-selected mutants which had an amino acid substitution in both GyrA 84 (Ser→Leu) and GrlA 80 (Ser→Phe) (Table 1), the ciprofloxacin MICs increased four to eightfold when the GrlA 80 (Ser→Phe) mutation was present. In contrast, the BMS-284756 MIC remained the same (Tables 1 and 2). To explore whether mutations outside the QRDRs might be responsible for the increased MICs for BMS-284756 fourth-step mutants, the entire gyrA/gyrB region was sequenced for two mutants.

TABLE 1. QRDR Sequence results of BMS-284756 mutant protein changes from parental strain S. aureus ISP794^{a,b}

Strain	Location of mutation for indicated gene			
	GrIA	GrIB	GyrA	GyrB
Third-step mutant strains				
3(756)6-1	116 (Ala→Glu)	— ^c	84 (Ser→Leu)	—
3(756)6-2	—	—	84 (Ser→Leu)	—
3(756)6-3	157 (Pro→Leu)	—	—	—
3(756)10-1	—	—	84 (Ser→Leu)	—
3(756)10-2	—	—	84 (Ser→Leu)	—
3(756)10-3	—	—	84 (Ser→Leu)	—
Fourth-step mutant strains				
4(756)6-1	—	—	84 (Ser→Leu)	—
4(756)6-2	80 (Ser→Phe)	—	84 (Ser→Leu)	—
4(756)10-1	—	—	84 (Ser→Leu)	—
4(756)10-3	80 (Ser→Phe)	—	84 (Ser→Leu)	—
Fifth-step mutant strains				
5(756)6-1	—	472 (Glu→Lys) ^d	84 (Ser→Leu)	—
5(756)6-2	—	472 (Glu→Lys)	84 (Ser→Leu)	—
5(756)10-1	—	—	84 (Ser→Leu)	—
5(756)10-3	—	—	84 (Ser→Leu)	—

^a First- and second-step BMS-284756-selected mutants had no changes in the QRDRs of the four genes sequenced.

^b Four ciprofloxacin-selected mutants, 1C6(1)-1, 2C6(1)-1, 3C6(1)-1, and 4C6(1)-1, were sequenced. They all had only one QRDR mutation, grlA (A116E).

^c The designation — indicates no change.

^d Mutations not previously reported.

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At multiples of the MIC, the FRE of ciprofloxacin in first-step mutants was compared to that of BMS-284756 (Table 3). The FREs of ciprofloxacin for the parental *S. aureus* strain ISP794, FREs were 1.5×10^{-7} to 6.8×10^{-7} at four times the MIC of ciprofloxacin and higher at two times the MIC. There were no mutants at eight times the MIC.

In contrast, the FRE for BMS-284756 was 8.0×10^{-10} at four times the MIC and higher at two times the MIC, and no resistant mutants were selected at eight times the MIC, representing approximately a 3-log difference in FRE between BMS-284756 and ciprofloxacin (Table 3). In all other steps, BMS-284756 showed a 1- to 2-log-lower FRE than did ciprofloxacin (Table 3).

Differences in the susceptibility of these resistant mutants to the four quinolones were observed. MICs of ciprofloxacin for the parental strain and the first-, second-, third-, and fourth-step ciprofloxacin-selected mutants were 0.125, 1, 4, 4 to 8, and 4 to 8 $\mu\text{g/ml}$, respectively. In contrast, levofloxacin MICs for the same mutants were 0.125, 0.5, 1, 1, and 1 to 2 $\mu\text{g/ml}$, respectively, and moxifloxacin MICs for the same mutants were 0.06, 0.06, 0.25 to 0.5, 0.25, and 0.25 $\mu\text{g/ml}$, respectively. However, BMS-284756 MICs were 0.015, 0.03, 0.03 to 0.06, 0.06, and 0.06 $\mu\text{g/ml}$, respectively. The emergence of resistant mutants selected by BMS-284756 resulted in cross-resistance to the other three quinolones (Table 2). In addition, one step was required using ciprofloxacin selection for the MICs for the *S. aureus* mutants to reach the ciprofloxacin breakpoint of 1 $\mu\text{g/ml}$, just as with *Streptococcus pneumoniae* (2), while four steps were required by BMS-284756 for MICs to increase to 1 $\mu\text{g/ml}$ (still within the anticipated breakpoint of 2 to 4 $\mu\text{g/ml}$ [J. Barrett, Bristol-Myers Squibb, unpublished]). These data agree with

results for *S. pneumoniae* which showed that BMS-284756-selected mutants were isolated at a relatively low rate and that mutations were initially found in the GyrA protein within the QRDR (2). BMS-284756 is primarily selecting for the mutations present in DNA gyrase in gram-positive bacteria. The low FRE for BMS-284756 in *S. aureus* ISP794 and the lack of effect of GrlA 80 (Ser \rightarrow Leu) mutations on its MIC indicate the potential of this drug to have reduced resistance development and to cover quinolone-resistant pathogens in the clinic.

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