

## Resistance Mechanisms in Clinical Isolates of *Candida albicans*

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Resistance to azole antifungals continues to be a significant problem in the common fungal pathogen *Candida albicans*. Many of the molecular mechanisms of resistance have been defined with matched sets of susceptible and resistant clinical isolates from the same strain. Mechanisms that have been identified include alterations in the gene encoding the target enzyme *ERG11* or overexpression of efflux pump genes including *CDR1*, *CDR2*, and *MDR1*. In the present study, a collection of unmatched clinical isolates of *C. albicans* was analyzed for the known molecular mechanisms of resistance by standard methods. The collection was assembled so that approximately half of the isolates were resistant to azole drugs. Extensive cross-resistance was observed for fluconazole, clotrimazole, itraconazole, and ketoconazole. Northern blotting analyses indicated that overexpression of *CDR1* and *CDR2* correlates with resistance, suggesting that the two genes may be coregulated. *MDR1* overexpression was observed infrequently in some resistant isolates. Overexpression of *FLU1*, an efflux pump gene related to *MDR1*, did not correlate with resistance, nor did overexpression of *ERG11*. Limited analysis of the *ERG11* gene sequence identified several point mutations in resistant isolates; these mutations have been described previously. Two of the most common point mutations in *ERG11* associated with resistance, D116E and E266D, were tested by restriction fragment length polymorphism analysis of the isolates from this collection. The results indicated that the two mutations occur frequently in different isolates of *C. albicans* and are not reliably associated with resistance. These analyses emphasize the diversity of mechanisms that result in a phenotype of azole resistance. They suggest that the resistance mechanisms identified in matched sets of susceptible and resistant isolates are not sufficient to explain resistance in a collection of unmatched clinical isolates and that additional mechanisms have yet to be discovered.

*Candida albicans* is a pathogenic yeast that causes oral, vaginal, and systemic infections (for a review, see reference 25). These infections are commonly associated with immune dysfunction, as they are frequently found in AIDS patients and bone marrow transplant patients. Antifungal drugs that are available for the treatment of these infections include the polyene amphotericin B and the azoles, such as fluconazole. In the 1990s, many human immunodeficiency virus (HIV)-infected patients received long-term, low-level azole antifungal therapy, which resulted in azole-resistant isolates of *C. albicans* (for a review, see reference 41). One study documented azole resistance in up to one-third of the oral *C. albicans* isolates from HIV-positive patients (12). In recent years, resistance to antifungal drugs has been documented in other patient populations such as bone marrow transplant recipients (9, 19, 20, 22, 24). In a clinical setting, there are many reasons why a fungal infection does not respond to antifungal drugs, including the immune status of the patient, the characteristics of the drug, and the susceptibility of the fungus to the drug (9, 41).

In the last 5 years, several molecular mechanisms by which *C. albicans* develops resistance to antifungal drugs have been elucidated (for a review, see reference 41). The azole drugs including fluconazole target lanosterol 14 $\alpha$ -demethylase, the product of the *ERG11* gene. Erg11p is one of the enzymes in the biosynthesis of ergosterol, the major sterol of fungal mem-

branes and an analogue of cholesterol in mammalian systems. Antifungal drug resistance has been associated with point mutations and increased levels of expression of the *ERG11* gene. Evidence is accumulating that changes in other enzymes in the ergosterol biosynthetic pathway can also contribute to resistance. Drug efflux from the cells is another component of resistance in *C. albicans*, as overexpression of two types of efflux pump has been correlated with antifungal resistance. The ABC transporter genes *CDR1* and *CDR2* encode ATP-dependent efflux pumps that are overexpressed in many azole-resistant isolates. Deletion of these genes results in hypersensitivity to azoles (1, 6, 36, 37). The major facilitator gene *MDR1* encodes a pump that uses the proton motive force at the membrane to transport drugs and other compounds across the plasma membrane. Overexpression of this pump is also associated with resistance, and deletion results in hypersensitivity to azole drugs (2, 5, 7). Recently, another major facilitator gene, *FLU1*, was identified in *C. albicans* (3). This gene increases the level of azole resistance when it is expressed in *Saccharomyces cerevisiae* and increases the level of susceptibility when it is deleted from *C. albicans*, but overexpression of the gene has not yet been correlated with azole resistance in clinical isolates of *C. albicans* (3).

The known molecular mechanisms of resistance are best illustrated by a series of 17 isolates from an HIV-infected patient (39–42). Resistance developed over time in this series of isolates. The alterations detected in these isolates included mutation and overexpression of the *ERG11* gene and overexpression of the efflux pumps encoded by *CDR1*, *CDR2*, and *MDR1*. Changes in the level of susceptibility are associated

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with each of these molecular alterations. The level of susceptibility for fungal cells is usually measured as the MIC, and the MIC determination method has recently been standardized for reproducibility and interlaboratory consistency (23).

To date, mechanisms of resistance have been determined in clinical isolates in which matched sets of resistant and susceptible isolates of the same strain were analyzed (14, 27, 37, 42). In the most recently published work, the molecular mechanisms of resistance in matched sets of susceptible and resistant isolates of the same strain from an HIV-infected patient population were investigated (27). The study found that 85% of isolates overexpressed efflux pumps, 65% of isolates had mutations in *ERG11*, and 35% of isolates overexpressed *ERG11*. Most of the point mutations identified in that study were previously described in a survey of point mutations in *ERG11* (18).

The purpose of the present study was to conduct an epidemiologic survey of resistance in an unmatched set of clinical isolates, half susceptible and half resistant, from a variety of clinical settings to determine if the standard molecular techniques and the current understanding of resistance mechanisms are sufficient to explain the resistance phenotypes of arbitrarily selected isolates, like those that might be found in isolate collections in a clinical microbiology laboratory.

#### MATERIALS AND METHODS

**Isolates and growth of cultures.** The *C. albicans* isolates used in this study represent a collection of 38 arbitrarily selected isolates from 12 hospitals in California. Approximately half of the isolates were susceptible to fluconazole and half were resistant to fluconazole. The isolates were from a variety of body sites. The isolates were initially identified as *C. albicans* at the hospital of origin and sent to one of the authors (D.A.S.) for MIC determination and further characterization. The isolates were plated on ChromAgar (DRG International, Mountainside, N.J.) to preliminarily characterize them as *C. albicans*. Two well-characterized isolates, a susceptible isolate (isolate C1 [isolate 2-76]) and a resistant isolate (isolate C17 [isolate 12-99]), from a series of 17 oral isolates from a single patient with AIDS (32) were used as controls.

Cultures were routinely inoculated from single colonies. The isolates were grown at 30°C in YEPD broth (10 g of yeast extract, 20 g of peptone, and 20 g of dextrose per liter) or on YEPD agar plates (10 g of yeast extract, 20 g of peptone, 20 g of dextrose, and 15 g of agar per liter). Isolates were stored at 4°C, subcultured as required, or stored at -80°C in YEPD broth containing 10% glycerol.

**Confirmation of isolate species.** Most of the clinical isolates grew as green colonies on ChromAgar plates, suggesting that they are *C. albicans*. However, *C. dubliniensis* also grows as dark green colonies on ChromAgar plates and forms hyphae similarly to *C. albicans*. To determine if any of the isolates were *C. dubliniensis*, a PCR was performed with two oligonucleotides (oligonucleotides CA-INT-L and CA-INT-R) directed to the transposable intron in the large-subunit (LSU) rRNA gene (rDNA), as described previously (21), and the PCR products were electrophoresed through an agarose gel. The electrophoresis pattern distinguishes three different subgroups of *C. albicans*, as well as *C. dubliniensis*.

**Susceptibility testing.** MICs were determined by the broth macrodilution method of NCCLS (23) by using laboratory reference resistant and susceptible isolates as internal controls. MIC breakpoints for clotrimazole have not been previously reported but were determined by the same protocols used for the other azole drugs. MICs were redetermined for many of the isolates in the collection; the MICs obtained by all repeat determinations fell within a fourfold range for all isolates tested, consistent with NCCLS standards.

**DNA and RNA extractions and Northern and Southern blotting analyses.** Genomic DNAs from the clinical isolates were prepared as described previously (8). Preparation of total RNA, restriction enzyme digestions, gel electrophoresis, Northern blotting, oligonucleotide labeling with polynucleotide kinase, and random priming for radioactive probe preparation were performed by standard published methods (17, 34). Total RNA was prepared from isolates in the mid-logarithmic phase of growth, when the cells had reached an optical density (600 nm) of approximately 1.0 (optical density range, 0.8 to 1.2). All reagents

were purchased from Fisher Scientific Co. (Pittsburgh, Pa.) or Sigma Chemical Co. (St. Louis, Mo.), unless otherwise specified.

**DNA probes.** Gene fragments and oligonucleotides were used as probes for Northern blotting. For each gene described below, the numbers represent the nucleotide position in the published sequence. If the order of the numbers is reversed, the probe was designed as the reverse complement of the sequence. The GenBank accession number, followed by the reference number for the publication in which the sequence is described, is given in parentheses. Oligonucleotides were prepared to be complementary to the mRNAs for *ACT1*, *FLU1*, *CDR1*, and *CDR2*. The probe for *ACT1*, the gene for actin, was a 50mer, positions 2527 to 2478 (GenBank accession no. X16377 [15]); the probe for the *FLU1* gene was a 48mer, positions 950 to 902 (GenBank accession no. AF188621 [3]); and the oligonucleotides for the *CDR* genes were as follows: *CDR1*, positions 1260 to 1211 (GenBank accession no. X77589 [30]), and *CDR2*, positions 950 to 902 (GenBank accession no. U63812 [36]). The probes for *ERG11* and *MDR1* were gene fragments prepared by PCR of genomic DNA. The segments that were amplified were as follows: *ERG11*, positions 164 to 1589 (GenBank accession no. X13296 [10]), and *MDR1*, positions 2885 to 3754 (GenBank accession no. X53823 [5]).

After hybridization and washing of the membranes used for Northern blotting, the filters were exposed to a phosphorimaging screen and scanned with a phosphorimager (Storm 860; Molecular Dynamics, Sunnyvale, Calif.). Signal quantification was carried out with the Molecular Dynamics ImageQuant program.

**Sequence analysis.** PCRs for sequences spanning the *ERG11* gene were performed with genomic DNA. The oligonucleotides used for the reactions and the processing of the PCRs have been described previously (40). The purified PCR fragments were sequenced with an automated DNA sequencer with *Taq* dye-primers and dye-terminator chemistries (Applied Biosystems, Foster City, Calif.).

**Restriction fragment length polymorphism (RFLP) analysis.** To screen the isolates for the D116E mutation within the *ERG11* gene, two oligonucleotides were used to create a PCR fragment that spans the restriction enzyme site. The oligonucleotides were at positions 133 to 151 and 702 to 683 (GenBank accession no. X13296 [10]). The PCR product is 550 bp in length and is split into fragments of 350 and 200 bp by digestion with *HindIII* when the D116E mutation is present.

To screen the isolates for the E266D mutation, two different oligonucleotides, from positions 678 to 696 and positions 1255 to 1234 (GenBank accession no. X13296 [10]), were used. The resulting PCR product was 577 bp. It was digested with *HpyCH4 IV* (New England Biolabs, Beverly, Mass.), an isoschizomer of *MaeII*. This enzyme digested the PCR product into bands of 198, 64, and 280 bp when the wild-type sequence was present and bands of 198 and 344 bp when the mutant sequence was present.

#### RESULTS

**Collection of isolates.** An initial screen on ChromAgar of the arbitrarily selected collection of isolates used in the present study indicated that two of the isolates were not *C. albicans* and that four of the isolates were mixtures of *C. albicans* and *C. tropicalis* cells. The two non-*C. albicans* isolates were not analyzed further. The four isolates with mixtures of cells were plated to obtain single colonies, and the *C. albicans* colonies were isolated and further analyzed. The *C. tropicalis* cells from the mixed populations were not analyzed further.

Two *C. albicans* isolates appeared dark green on ChromAgar plates, an indication that the isolates might be *C. dubliniensis*. Oligonucleotides were used in PCR to obtain a fragment that spans the transposable intron in the LSU rDNA (21) to distinguish the 36 isolates into *C. dubliniensis* and three subgroups of *C. albicans* isolates (Table 1). All 36 isolates were *C. albicans*; none were *C. dubliniensis*. Twenty-nine of the 36 isolates were subgroup A, while 3 isolates were subgroup B and 4 isolates were subgroup C. There does not appear to be a correlation between the subgroup and the MIC or the mechanism of resistance for this series of isolates, consistent with previous findings (21).

**MICs.** The MICs of four different azoles, fluconazole, clotrimazole, itraconazole, and ketoconazole, were determined

TABLE 1. Identification, isolation, MIC, and resistance mechanism for isolate collection

Reference isolate no.	Isolate designation	Strain type <sup>a</sup>	Hospital code	Site of isolation	MIC ( $\mu\text{g/ml}$ ) <sup>c</sup>				Gene(s) overexpressed	ERG11 sequence analysis <sup>d</sup>
					FLC	CLT	ITC	KTC		
1	95-142	A	A	Throat	<u><math>\geq 64</math></u>	<u>8</u>	<u><math>\geq 8</math></u>	<u>4</u>	<i>CDR1, CDR2</i>	
2	95-68	A	B	Unknown	<u><math>\geq 64</math></u>	<u>4</u>	<u><math>\geq 8</math></u>	<u>16</u>	<i>CDR1, CDR2</i>	
3	95-158	A	F	Oropharynx	<u><math>\geq 64</math></u>	<u>4</u>	<u>8</u>	<u>4</u>		Hot spot III
4	98-112	B	H	Blood	<u><math>\geq 64</math></u>	<u>2</u>	<u><math>\geq 8</math></u>	<u>16</u>		Complete gene
5	98-145 <sup>e</sup>	A	G	Urine	<u><math>\geq 64</math></u>	<u>2</u>	<u><math>\geq 8</math></u>	<u>16</u>		Hot spot III
6	95-111 <sup>e</sup>	A	F	Mouth	<u><math>\geq 64</math></u>	<u>2</u>	<u><math>\geq 8</math></u>	<u>4</u>	<i>CDR1, CDR2</i>	
7	95-16	A	F	Paracentesis fluid	<u><math>\geq 64</math></u>	<u>2</u>	<u><math>\geq 8</math></u>	<u>4</u>		Complete gene
8	96-23	A	A	Throat	<u><math>\geq 64</math></u>	<u>2</u>	<u>1</u>	<u>8</u>		Hot spot III
9	98-136	C	B	Peritoneal dialysate	<u><math>\geq 64</math></u>	<u>1</u>	<u><math>\geq 8</math></u>	<u>16</u>		Hot spot III
10	95-188	A	J	Wrist tissue	<u>64</u>	<u>2</u>	<u><math>\geq 8</math></u>	<u>16</u>		
11	95-165	A	F	Vaginal	<u><math>\geq 64</math></u>	<u>4</u>	<u><math>\geq 8</math></u>	<u><math>\leq 0.5</math></u>		Complete gene
12	95-157	A	I	Tongue	<u>64</u>	<u>4</u>	<u>2</u>	<u><math>\leq 0.5</math></u>		
13	95-190	A	K	Esophageal tissue	<u>32</u>	<u>2</u>	<u>4</u>	<u>8</u>	<i>CDR1, CDR2</i>	
14	95-120	A	A	Blood	<u>32</u>	<u>2</u>	<u>1</u>	<u>8</u>		
15	95-133	A	B	Mouth	<u>64</u>	<u>1</u>	<u><math>\leq 0.5</math></u>	<u><math>\leq 0.5</math></u>	<i>CDR1, CDR2</i>	
16	98-24	C	L	Urine	<u>8</u>	<u>0.5</u>	<u>1</u>	<u>1</u>		
17	97-363	A	L	Abdominal fluid	<u>2</u>	<u>2</u>	<u><math>\leq 0.5</math></u>	<u>16</u>		
18	98-144	A	G	Blood	<u>2</u>	<u>0.5</u>	<u><math>\geq 8</math></u>	<u>4</u>		
19	97-224	A	F	Blood	<u>2</u>	<u>0.5</u>	<u>2</u>	<u>1</u>		
20	98-155	A	G	Blood	<u><math>\leq 0.5</math></u>	<u>0.5</u>	<u>1</u>	<u>2</u>		
21	98-143	A	A	Urine	<u><math>\leq 0.5</math></u>	<u><math>\leq 0.25</math></u>	<u><math>\geq 8</math></u>	<u>4</u>		
22	96-25 <sup>f</sup>	C	A	Abdominal fluid	<u>32</u>	<u>0.5</u>	<u>1</u>	<u><math>\leq 0.5</math></u>	<i>ERG11, MDR1</i>	Hot spot III
23	98-141	C	B	Indwelling catheter	<u>8</u>	<u>2</u>	<u><math>\leq 0.5</math></u>	<u><math>\leq 0.5</math></u>		
24	98-7	A	A	Blood	<u>4</u>	<u><math>\leq 0.25</math></u>	<u><math>\geq 8</math></u>	<u><math>\leq 0.5</math></u>		
25	97-119 <sup>f</sup>	A	C	Abdomen	<u>1</u>	<u>1</u>	<u><math>\leq 0.5</math></u>	<u><math>\leq 0.5</math></u>		
26	98-216	A	A	Urine/catheter	<u>1</u>	<u><math>\leq 0.25</math></u>	<u><math>\geq 8</math></u>	<u><math>\leq 0.5</math></u>		
27	98-85 <sup>f</sup>	A	A	Peritoneal fluid	<u><math>\leq 0.5</math></u>	<u>0.5</u>	<u>1</u>	<u><math>\leq 0.5</math></u>	<i>CDR1, (CDR2)</i>	
28	98-125	A	A	Blood	<u><math>\leq 0.5</math></u>	<u><math>\leq 0.25</math></u>	<u>2</u>	<u><math>\leq 0.5</math></u>	<i>CDR2</i>	
29	97-87	B	A	Blood	<u><math>\leq 0.5</math></u>	<u><math>\leq 0.25</math></u>	<u><math>\leq 0.5</math></u>	<u>8</u>		
30	98-13	A	E	Paracentesis fluid	<u><math>\leq 0.5</math></u>	<u><math>\leq 0.25</math></u>	<u><math>\leq 0.5</math></u>	<u>1</u>		
31	95-175	A	A	Endotracheal aspirate	<u>32</u>	<u><math>\leq 0.25</math></u>	<u><math>\leq 0.5</math></u>	<u><math>\leq 0.5</math></u>		
32	97-197 <sup>f</sup>	A	B	Blood	<u>2</u>	<u>0.5</u>	<u><math>\leq 0.5</math></u>	<u><math>\leq 0.5</math></u>		
33	98-8	A	B	Oral	<u><math>\leq 0.5</math></u>	<u>0.5</u>	<u><math>\leq 0.5</math></u>	<u><math>\leq 0.5</math></u>	<i>ERG11</i>	
34	98-113	A	D	Peritoneal fluid	<u><math>\leq 0.5</math></u>	<u><math>\leq 0.25</math></u>	<u><math>\leq 0.5</math></u>	<u><math>\leq 0.5</math></u>		
35	98-126	A	A	Chest tube	<u><math>\leq 0.5</math></u>	<u><math>\leq 0.25</math></u>	<u><math>\leq 0.5</math></u>	<u><math>\leq 0.5</math></u>		
36	98-146	B	A	Pleural tissue	<u><math>\leq 0.5</math></u>	<u><math>\leq 0.25</math></u>	<u><math>\leq 0.5</math></u>	<u><math>\leq 0.5</math></u>		
C1 <sup>g</sup>	2-76	A		Oral	<u>0.25</u>	<u>NA<sup>h</sup></u>	<u><math>\leq 0.5</math></u>	<u><math>\leq 0.5</math></u>		
C17 <sup>g</sup>	12-99	A		Oral	<u><math>\geq 64</math></u>	<u>NA</u>	<u><math>\leq 0.5</math></u>	<u>2</u>	<i>ERG11, CDR1, CDR2, MDR1</i>	

<sup>a</sup> Strain type, as determined by PCR and restriction enzyme digestion (19).

<sup>b</sup> Isolates were obtained from 12 hospitals throughout California, coded by letter.

<sup>c</sup> FLC, fluconazole; CLT, clotrimazole; ITC, itraconazole; KTC, ketoconazole. Results that indicate that the isolate was resistant according to the breakpoints discussed in the text are underlined.

<sup>d</sup> The *ERG11* genes of some of the isolates were sequenced. Either the entire gene or hot spot III, which includes amino acids 405 to 488, was sequenced.

<sup>e</sup> Colonies were dark green on ChromAgar but were confirmed to be *C. albicans*.

<sup>f</sup> The isolates were originally parts of mixed infections.

<sup>g</sup> C1 and C17, control isolates 1 and 17, respectively, from a well-characterized matched set.

<sup>h</sup> NA, not available.

(Table 1). For some isolates the MICs of all azoles were relatively high, for other isolates the MICs of one or two azoles were relatively high but the MICs of the other azoles tested were low, and some isolates were susceptible to all azoles tested.

MIC breakpoints that correlate with clinical treatment success have been determined for fluconazole and itraconazole (33). For fluconazole, a resistant isolate is defined as an isolate for which the MIC is  $\geq 64 \mu\text{g/ml}$ ; a susceptible isolate is an isolate for which the MIC is  $\leq 8 \mu\text{g/ml}$ . Isolates for which MICs are 16 and  $32 \mu\text{g/ml}$  are defined as susceptible, dose dependent

(SDD), as they appear to respond to higher doses of drug. For itraconazole, the breakpoints have been defined as follows: susceptible, MIC  $\leq 0.125 \mu\text{g/ml}$ ; SDD, MIC = 0.25 to  $0.5 \mu\text{g/ml}$ ; and resistant, MIC  $\geq 1 \mu\text{g/ml}$ . The 36 *C. albicans* isolates included 13 that were resistant, 4 that were SDD, and 19 that were susceptible to fluconazole and 24 that were resistant and 12 that were susceptible or SDD to itraconazole. Clinical breakpoints for ketoconazole have not been proposed, but they are likely to be close to the breakpoints for itraconazole, as the drug concentrations achievable in blood are similar for both drugs when similar doses are administered, and

the distributions of resistant and sensitive isolates are similar for both itraconazole and ketoconazole when  $\geq 1 \mu\text{g/ml}$  is used to define resistance to both drugs. By using an MIC of  $\geq 1 \mu\text{g/ml}$  to define resistance in this study, the 36 isolates included 20 isolates that were resistant to ketoconazole and 16 isolates that were susceptible or SDD to ketoconazole. The MICs of clotrimazole are not routinely determined, although clotrimazole is commonly used to treat mucosal candidiasis in children with HIV infection. Recently, Pelletier et al. (26) described a collection of isolates from children for which the clotrimazole MIC was  $\geq 1 \mu\text{g/ml}$ ; these isolates represented 10% of the total collection and were cross resistant to other azoles. Amphotericin B was required for successful treatment. By using an MIC of  $\geq 1 \mu\text{g/ml}$  to define clotrimazole resistance, the 36 isolates included 18 that were resistant and 18 that were susceptible or SDD to clotrimazole. The distribution of isolates in Table 1 shows that 10 isolates (isolates 1 to 10) were resistant to all azoles tested, 4 isolates (isolates 11 to 14) were resistant to three azoles, 7 isolates (isolates 15 to 21) were resistant to two azoles, 9 isolates (isolates 21 to 30) were resistant to one azole, and 6 isolates (isolates 31 to 36) were susceptible to all azoles tested. This distribution of resistance to azoles was broad and appropriate for the testing of the molecular mechanisms of resistance in a collection of clinical isolates.

**Molecular mechanisms of resistance.** Previous work on the analysis of resistance mechanisms has routinely used matched sets of susceptible and resistant clinical isolates of the same strain (14, 27, 37, 42). This is necessary, as *C. albicans* is mostly clonal in nature (31) and isolates might differ considerably in their levels of expression of different genes. Matched sets of isolates were not available for the collection of isolates described above, nor will matched sets be available in most clinical settings. The purpose of the experiments described below is to use standard molecular techniques to screen for known resistance mechanisms without the aid of matching the isolates with pretreatment isolates and to determine if these methods are sufficient to determine mechanisms of resistance in unmatched isolates. Controls for these experiments included previously characterized matched susceptible and resistant isolates from the series of 17 isolates discussed above and hybridization with *ACT1* as a control to normalize the amount of RNA from each isolate. Since the specific activities of the probes vary, the relative intensities of the gene-specific hybridization signals compared to that for *ACT1* are presented, but the absolute values or a numerical value for the relative intensities are not significant, as they will vary with the specific activities of the probes. The relative levels of expression of the hybridization signals by the isolates are presented in Fig. 1 for each probe. The isolates are numbered on the basis of their levels of resistance (Table 1), so the first 10 isolates on the left side of Fig. 1 are resistant to all four azoles, and the last 6 isolates on the right side of the figure are susceptible to all four azoles.

**CDR efflux pump overexpression.** The *CDR* genes are commonly overexpressed in resistant isolates (14, 27, 37). In general, the *CDR* genes are not normally expressed at high levels in susceptible isolates (16). Total RNA was prepared from the 36 isolates while they were in the mid-logarithmic phase of growth, which has been standard procedure in previous studies. The RNA was electrophoresed through an agarose gel,

blotted onto nitrocellulose, and hybridized with oligonucleotides specific for *CDR1* and *CDR2*.

Six isolates from the collection expressed *CDR1* at higher levels than susceptible control isolate C1 did (Fig. 1A). The isolates overexpressing *CDR1* included isolates 1, 2, 6, 13, 15, and 27. These isolates came from four different hospitals and were isolated from the mouth, throat, esophagus, and peritoneal fluid (the site of isolation of isolate 2 was unknown) (Table 1). Of the six isolates that overexpressed *CDR1*, three isolates were resistant to all four azoles, one isolate was resistant to three azoles and SDD to the other azole, one isolate was resistant to two azoles, and one isolate was resistant to one azole (itraconazole).

Six isolates expressed *CDR2* at higher levels than susceptible control isolate C1 did. The isolates overexpressing *CDR2* included isolates 1, 2, 6, 13, 15, and 28 (Fig. 1B). Five of the six isolates overexpressed both *CDR1* and *CDR2*, and these five isolates were resistant to most, if not all, azoles tested (Table 1). The level of expression of *CDR2* in isolate 27, which overexpresses *CDR1*, is approximately the same as that in isolate C1, suggesting that *CDR1* and *CDR2* are jointly overexpressed in six different isolates from this collection. When all isolates that overexpressed *CDR1* and *CDR2* are considered, there is no preference for resistance to one azole over that to any of the other azoles tested.

**MDR1 efflux pump overexpression.** The *MDR1* gene for the major facilitator efflux pump has been shown to be expressed in several resistant clinical isolates (14, 27, 37, 39). The *MDR1* gene is not normally expressed in susceptible isolates under normal conditions, as determined by Northern blotting analysis (16). Northern blotting analysis was performed as described above with an *MDR1*-specific probe (Fig. 1C). Only one isolate, isolate 22, showed significant levels of expression of the *MDR1* gene by Northern blotting analysis, although its level of expression was still lower than the level of *MDR1* expression in isolate C1. Isolate 22, originally taken from abdominal fluid, was mildly resistant to itraconazole, but it was susceptible to the other azoles tested (Table 1). Clearly, *MDR1* overexpression during the mid-logarithmic phase of growth is not closely correlated with resistance in this collection. Two other isolates, isolates 9 and 12, had low levels of expression of *MDR1* (Fig. 1C). Isolate 9 was resistant to all four azoles tested; isolate 12 was resistant to all the azoles tested but ketoconazole (Table 1).

**FLU1 efflux pump overexpression.** When the recently described *FLU1* gene is deleted from *Saccharomyces* or *C. albicans*, the resulting cells are hypersusceptible to azole drugs (3). However, overexpression of the gene has not been linked to resistance in clinical isolates. Northern blotting analysis was performed, and *FLU1* gene expression was quantitated as described above using an *FLU1*-specific probe (Fig. 1D). In this Northern blotting analysis, isolate C1 expressed more *FLU1* than isolate C17 did, consistent with previous data from our laboratory (data not shown). Seven isolates (isolates 4, 13, 14, 25, 27, 31, and 33) expressed more *FLU1* than the other isolates did. There does not appear to be a correlation between expression of high levels of *FLU1* and resistance in these isolates.

**ERG11 target enzyme overexpression.** The gene for the azole target enzyme, *ERG11*, is expressed during growth of

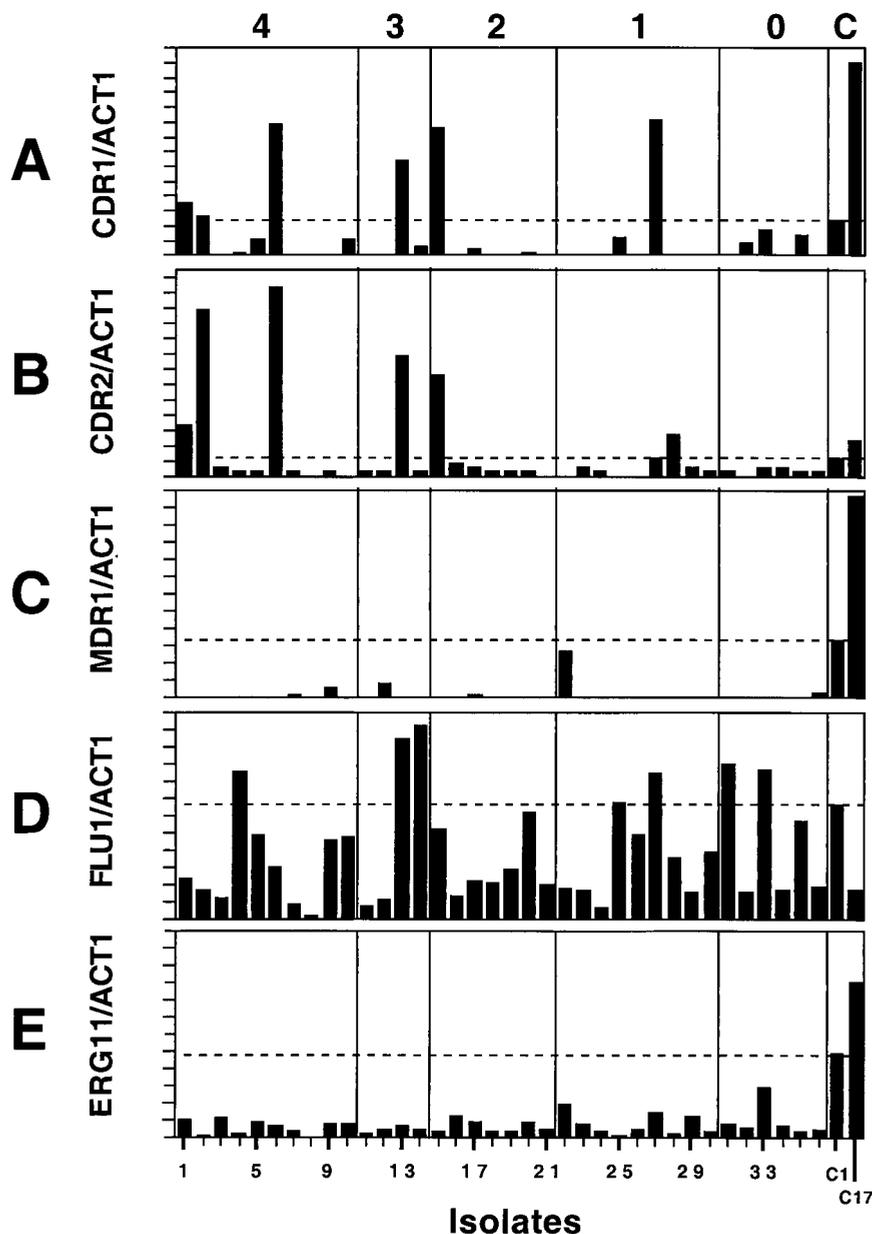


FIG. 1. Levels of gene expression relative to *ACT1* gene expression, as determined by Northern blotting analysis. Total RNA from the 36 isolates and 2 controls was hybridized with gene-specific probes on Northern blots. The mRNA signals for the two probes were quantitated with a Storm phosphorimager, and the ratio of the two intensities was computed and graphed. The x axis represents the 36 isolates, arranged in numerical order. The last two isolates on the right (isolates 37 and 38) are the control isolates (isolates C1 and C17, respectively) from the well-characterized matched series. The dotted lines indicate the level of expression of each of the genes by susceptible control isolate C1. As indicated at the top, isolates 1 to 10 are resistant to four azoles, isolates 11 to 14 are resistant to three azoles, isolates 15 to 21 are resistant to two azoles, isolates 22 to 30 are resistant to one azole, and isolates 31 to 36 are susceptible to all azoles tested. C, control.

both susceptible and resistant isolates (16). Northern blotting analysis was performed and *ERG11* gene expression was quantitated with total RNA from cells in the mid-logarithmic phase of growth using an *ERG11*-specific probe (Fig. 1E). There was considerable variation in the levels of *ERG11* expression within this collection of isolates, but the level of expression did not appear to be related to the azole resistance of the isolates. All of the isolates expressed *ERG11* at levels below the level at which isolate C1 expressed it. The highest relative level of

*ERG11* expression was found by isolate 33, which is an isolate that is susceptible to all azoles tested, suggesting that absolute levels of *ERG11* expression are not a priori linked to azole resistance. One isolate (isolate 22) is SDD to fluconazole and mildly resistant to itraconazole, and it had a relatively high level of *ERG11* expression. It is noteworthy that this isolate is the one isolate in the collection with significant levels of expression of *MDR1*. The distribution of *ERG11* expression suggests that high levels of *ERG11* mRNA are not necessarily

TABLE 2. Protein sequence alterations associated with resistance<sup>a</sup>

Mutation	Result for isolates whose complete gene was sequenced			Result for isolates for which the hot spot III region was sequenced				
	4	7	11	3	5	8	9	22
D116E	Heterozyg.	Heterozyg.	Homozyg.					
K128T	WT	Heterozyg.	Homozyg.					
K143R	WT	WT	Homozyg.					
E266D	Homozyg.	WT	WT					
V437I	Heterozyg.	WT	WT	WT	Homozyg.	WT	WT	Homozyg.
G448R	WT	WT	WT	WT	WT	Homozyg.	WT	WT

<sup>a</sup> Point mutations that resulted in amino acid changes identified by sequencing of *ERG11* from the indicated isolates. WT, the amino acid at that position was wild type (no change occurred in either allele); Heterozyg., the amino acid at that position was mutated as indicated in one allele; Homozyg., the amino acid at that position was mutated as indicated in both alleles.

associated with increased azole resistance. There is no obvious link between the site of isolation in the human body and the level of *ERG11* expression.

***ERG11* sequence analysis.** Point mutations within the *ERG11* gene have been identified in several matched isolates (18, 27, 35, 40). In a recent compilation of mutations, Marichal et al. (18) noted that there were three diffuse hot spots within the amino acid sequence of the *ERG11* gene, including amino acid regions 105 to 165, 266 to 287, and 405 to 488. Sequence analysis of the entire *ERG11* genes of three resistant isolates (isolates 4, 7, and 11) was performed. These isolates did not overexpress any of the genes tested, including *ERG11*, *CDR1*, *CDR2*, and *MDR1*. The genomic DNAs were amplified by PCR, and the sequences of the PCR products were determined as described previously (40) and compared to the published sequence (10). As expected from unrelated clinical isolates, frequent silent mutations that do not change the protein sequence were identified (data not shown). In addition, since *C. albicans* is a diploid organism, allelic differences were observed upon sequencing of DNA from single isolates (data not shown). The sequence alterations that resulted in changes in the protein sequence are listed in Table 2. Amino acid substitutions are usually denoted by the amino acid position, preceded by the single-letter amino acid designation for the wild type and followed by the single-letter amino acid designation for the mutant (e.g., D116E). Five mutations, located in the three hot spots, were observed in the three complete sequences; these mutations included D116E, K128T, K143R, E266D, and V437I (Table 2). Careful inspection of the sequence data distinguished between point mutations in both alleles (the isolate is homozygous for the mutation) and mutations in only one allele (the isolate is heterozygous for the mutation). Isolate 11 contained three homozygous mutations (D116E, K128T, and K143R), all within the first hot spot. Isolate 7 contained two heterozygous mutations (D116E/D and K128T/K). Both of these mutations were also found in isolate 11 and were present in the first hot spot. Isolate 4 was heterozygous for D116E/D in the first hot spot, homozygous for E266D in the second hot spot, and heterozygous for V437I/V in the third hot spot.

As the mutations identified in these three isolates had previously been identified in other collections of clinical isolates, limited sequence analysis of the third hot spot region was also performed for five other resistant isolates. The V437I mutation was homozygous in isolates 5 and 22 (Table 2). An additional homozygous mutation, G448R, was identified in isolate 8. Of

these six mutations, the D116E, K128T, K143R, E266D, and V437I mutations have been described in isolates from other studies (18). The sixth mutation, G448R, has not been described previously, but a similar mutation, G448E, has been identified previously (13). D116E, K128T, and E266D were three of the most common mutations identified in a recent survey (18). Sequence analysis of the eight isolates from the present collection which had limited pump expression failed to identify additional mutations within the *ERG11* gene. Therefore, sequencing of additional isolates with unknown mechanisms of resistance was not performed.

***ERG11* RFLP analysis.** The D116E mutation in the first hot spot of *ERG11* was identified in each of the three isolates whose complete sequences were determined, and it was the most common mutation identified in a recent survey of mutations (18). The D116E mutation can be identified by *Hind*III restriction enzyme digestion of PCR products that span the mutation, since *Hind*III will recognize the sequence of the mutant allele but not the sequence of the wild-type allele. All 36 isolates were screened for the mutation (Table 3). Four isolates were homozygous for the mutation at amino acid position 116, and 19 isolates were heterozygous for the mutation at amino acid position 116. The four isolates that were homozygous were resistant to at least two azoles, although the numbers of isolates are insufficient to be significant.

The E266D mutation in the second hot spot of *ERG11* is also a frequent mutation in the *ERG11* gene (18). This mutation can be screened for by restriction enzyme digestion of PCR fragments that span this region. *Mae*II and its isoschizomers digest the wild-type sequence but do not digest the mutant sequence. PCR products that include the sequence were digested and analyzed by agarose gel electrophoresis. The results (Table 3) indicate that there is no correlation between the E266D mutation and azole resistance.

## DISCUSSION

The goal of the series of experiments described here was to use standard techniques to identify molecular mechanisms of resistance in a collection of arbitrarily selected susceptible and resistant isolates. The results of the study do not define any specific correlations. Six strains overexpressed *CDR1* and *CDR2*. This is much lower than the frequencies observed in oral isolates from patients with AIDS. Five of the six strains overexpressing *CDR1* and *CDR2* were resistant to one or more of the four azoles tested (Table 1), although the results are not

TABLE 3. RFLP analysis of the sequences at positions 116 and 266 in *ERG11*<sup>a</sup>

Amino acid present	Isolate(s) resistant to the following no. of azoles:				
	Four	Three	Two	One	None
116D/D	2, 5, 8, 9		15, 18, 20, 21	22, 23, 30	31, 32
116D/E	1, 3, 4, 7, 10	13, 14	17, 19	24, 25, 26, 27, 28, 29	33, 34, 35, 36
116E/E	6	11, 12	16		
266E/E	1, 2, 3, 5, 6, 7, 8, 9, 10	11, 13, 14	15, 17, 18, 19, 21	22, 24, 25, 26, 27, 28, 30	31, 33, 34
266E/D	4		16, 20	29	36
266D/D		12		23	32, 35

<sup>a</sup> RFLP analysis was performed with the 36 isolates to assess the sequences at two locations, corresponding to amino acids 116 and 266, as described in Materials and Methods. The possible amino acids at those positions can be distinguished by RFLP analysis.

statistically significant for any drug. The one strain that overexpressed *MDR1* was resistant to only one azole, while low-level expression of *MDR1* occurred in two strains resistant to three or four azoles (Fig. 1). Similarly, overexpression of *FLU1*, overexpression of *ERG11*, or frequent mutations within the *ERG11* genes were not correlated with resistance in this collection. Overall, this suggests that standard molecular analyses of unmatched isolates is not sufficient to identify resistance mechanisms.

An important initial analysis of the collection of isolates used in the present study was confirmation that the isolates were *C. albicans*. Misidentification is not an uncommon problem, and experience indicates that isolates should always be rechecked for species identification and for the possibility that mixtures of isolates are present (data not shown). Rechecking of the hospital isolates indicated that two of the isolates were not *C. albicans*, and ChromAgar plate cultures of four of the isolates contained mixtures of *C. albicans* and *C. tropicalis*. Failure to detect these non-*C. albicans* isolates would have had important effects on the molecular analyses, as the oligonucleotides used as probes and primers for PCR would not react with the nucleic acids from the other species. Mixed infections would also have had consequences for MIC determinations and molecular characterizations. The possibility that mixtures of *C. albicans* strains were present was not addressed, although single colonies were used for the molecular analyses.

Recent identification and characterization of *C. dubliniensis* indicate that the species is easily confused with *C. albicans*. Limited techniques are available to quickly differentiate between *C. albicans* and *C. dubliniensis*. PCR with oligonucleotides that span a transposable element in the LSU rDNA of these species is a simple and specific test (21). Use of this test with the isolate collection indicated that all of the isolates were *C. albicans*. Furthermore, most of the isolates were identified as members of subgroup A of *C. albicans*, as defined previously (21).

The isolates in this collection were obtained from 38 different patients in 12 different hospitals throughout California over a 4-year period, making it highly unlikely that these isolates were related strains of *C. albicans*. This was confirmed by the distribution of the RFLP types presented in Table 3. The isolates in this collection were taken from a variety of body sites, including oral and vaginal mucosae, blood, urine, and abdominal fluid. While the HIV infection status of these patients is not known, it is likely that a large number of these

patients infected with either resistant or susceptible organisms are not HIV-positive or AIDS patients.

The relationship between the MIC and clinical success or failure is an important issue. Breakpoints between susceptibility and resistance for isolates from patients with oropharyngeal candidiasis have been determined for itraconazole and fluconazole (33). However, these breakpoints may not apply to isolates from patients with systemic infections. Breakpoints for ketoconazole and clotrimazole have not been proposed. The MICs of four azoles, fluconazole, clotrimazole, itraconazole, and ketoconazole, were determined for all of the isolates (Table 1).

The clotrimazole MIC was greater than 1 µg/ml for all 13 fluconazole-resistant isolates; this MIC has been suggested to be associated with clotrimazole resistance (26). In addition, of four isolates that were SDD to fluconazole, the clotrimazole MIC was >1 µg/ml for two of them. Sixteen isolates were considered susceptible to both fluconazole and clotrimazole, and the fluconazole and clotrimazole susceptibilities were discordant for only 3 of 36 isolates (fluconazole susceptible, clotrimazole resistant), demonstrating a strong correlation between fluconazole and clotrimazole resistance. This has important consequences for AIDS patients being treated with clotrimazole, especially HIV-infected children.

The MICs of itraconazole and ketoconazole were consistent with those of fluconazole and clotrimazole. Only 2 of the 17 isolates resistant or SDD to fluconazole would have been considered itraconazole susceptible and only 5 of the 17 isolates would have been considered ketoconazole susceptible by using the definitions of resistance proposed in this study. Ten isolates were resistant to all azoles tested, and most isolates were resistant to at least one of the azoles tested.

The collection of resistant isolates used in the present study was screened for the currently characterized molecular mechanisms of azole resistance. The expression of *CDR1* and *CDR2* in this collection (Fig. 1A and B) shows that several resistant isolates overexpress both genes. However, it was also found that at least one isolate resistant to only one azole also expressed large amounts of *CDR1* and *CDR2* (isolate 27 for *CDR1*, isolates 27 and 28 for *CDR2*). The fact that *CDR1* and *CDR2* are overexpressed simultaneously in six isolates suggests that the genes are coregulated. It raises the possibility that other efflux pumps are also coregulated, but the contributions of these other pumps to resistance cannot be predicted. The oligonucleotide probes specific for *CDR1* and *CDR2* do not

cross-hybridize to each other, to any other *CDR* genes, or to other genomic DNA (data not shown). Therefore, cross-hybridization cannot explain the coregulation.

The gene for the *MDR1* efflux pump is not expressed at high levels in cells in the mid-logarithmic phase of growth (16). The overexpression of *MDR1* in the present collection of isolates was limited to three isolates, all of which were resistant to one or more azole drugs. It has previously been shown that the *MDR1* message is variable in its pattern of expression even within a single isolate, possibly due to message instability (16). This may contribute to the expression or lack of expression of *MDR1* observed in these isolates.

The patterns of expression of the resistance genes in the present panel of isolates have been characterized only during the mid-logarithmic phase of growth, characterized as an optical density of approximately 1.0 in a single medium in the absence of azole drugs (Fig. 1), which is the standard methodology for the study of resistance mechanisms. The timing of expression of most of these genes has been studied during cellular growth (from the early logarithmic to the late stationary phase) in the control series of isolates (16). However, study of the timing of expression of each of these isolates in the presence and absence of azole drugs would be an extensive analysis that was beyond the scope of the present study.

The gene for the *FLU1* pump has not previously been associated with resistance (3), but its expression in this collection was evaluated because of the phenotypes of *Saccharomyces* and *Candida* cells from which the gene was deleted. The levels of expression do not clearly correlate with the levels of resistance of the isolates, adding additional support to the hypothesis that the gene is not involved in resistance in clinical isolates.

The pattern of *ERG11* expression is also variable and appears to have no correlation with resistance. The one resistant isolate that overexpressed *MDR1* also overexpressed *ERG11*, which might provide a hint that there is a link between the two. It is noteworthy that the levels of expression of *ERG11* in isolate C1 are elevated compared to those in all of the other isolates. This suggests that *ERG11* expression in the control susceptible isolate may already have been altered slightly in response to azole drugs but had not resulted in resistance.

The levels of mRNA for the genes associated with resistance were determined during the mid-logarithmic phase of growth in the absence of azole drugs, as has been the case previously (41). Certainly, the azole drugs will have additional effects on the levels of expression of these genes, which will be dependent on the concentration of the drug and the MIC for the cell. These factors add considerable complexity to the analysis and are beyond the scope of the present study.

Sequence analysis of the isolates was not extensive, but sufficient sequences were obtained to indicate that many of the point mutations identified in the present collection have already been identified in other isolates that have been sequenced previously (18). Therefore, it was likely that additional sequencing of the present series of isolates would not be informative. The mutations identified in the three complete sequences are located within the three hot spots identified by Marichal et al. (18).

The RFLP analysis for D116E and E266D performed in the present study is important, as it is the first time that these point

mutations have been screened for in a large collection of isolates, both susceptible and resistant. The fortuitous presence of a restriction enzyme recognition site at both locations allows easy screening of these isolates. Table 3 shows that both mutations are present in both susceptible and resistant isolates, at least in a heterozygous state. The absence of a susceptible isolate homozygous for 116E may be due to random sampling. These data indicate that two of the most frequent mutations identified in the survey are the result of allelic polymorphisms and are not likely to be associated with resistance.

To date, analyses of resistance have focused on oral isolates from HIV-infected patients, from whom matched susceptible and resistant isolates are most easily obtained (14, 27, 37, 42). The present analysis used the same techniques with a collection of unmatched isolates that included many isolates that were isolated from blood, urine, and other body fluids and locations. As isolates from HIV patients are usually oral, the isolates in this collection are unlikely to be associated with HIV infection. In addition, this survey documents the levels of resistance to three other azoles. A collection of isolates selected to include approximately equal numbers of isolates resistant and susceptible to fluconazole were shown to comprise a spectrum of susceptibility to the four azoles.

The most recent analysis of matched sets of oral isolates from HIV-infected patients described pump overexpression by 85% of fluconazole-resistant isolates and *ERG11* overexpression by 35% of fluconazole-resistant isolates (27). Estimates from the present survey showed that pump overexpression was found in 31% of fluconazole-resistant isolates (35% if isolates SDD to fluconazole are included), but it was also found in 11% of fluconazole-susceptible isolates. Similar percentages were found for the other azoles. The levels of expression of *ERG11* represent a gradual continuum, so it is difficult to define overexpression of this gene without a matched control.

The methods used in this analysis included Northern blotting analysis, DNA sequencing, and RFLP analysis, all of which can be done relatively quickly with a collection of isolates. Additional techniques could be applied to these isolates, although such techniques require extensive effort for each isolate. For instance, biochemical analysis of ergosterol synthesis could be performed to determine the sensitivity of the ergosterol synthesis pathway to the azole drugs, as reported previously (40). Microsomal analysis of Erg11p could also be performed to determine the susceptibility of an isolate (11). Drug accumulation can also be monitored, as described previously (4), by using fluorescent dyes. However, previous work suggests that the *CDR* pumps are primarily responsible for the efflux of rhodamine 6G, so the data from studies with fluorescent dyes may be limited. Finally, chromosomal copy numbers can be altered in resistant isolates compared to the number in matched susceptible isolates (28). However, *Candida* chromosome patterns are frequently used to differentiate strains (29), so differences in chromosome size or copy number between strains in a collection may not be specifically associated with resistance. Finally, *ERG11* genes from resistant isolates can be expressed in *Saccharomyces*, in which they can be analyzed in greater detail (27, 35). Such an analysis is best controlled by use of a susceptible version of the enzyme, which was not available from the present collection of isolates, and the tech-

niques used to express the genes eliminate the allelic variation that was shown to occur in the collection of isolates.

The known mechanisms of azole resistance were previously defined in matched sets of susceptible and resistant isolates, each from a single strain (for a review, see reference 41). Alterations in the resistant isolates were identified because they differ from the susceptible isolates of the same strain. However, these matched sets are rarely available in a clinical setting. It was of interest to use the methods that have been applied to matched sets of isolates in an analysis of a random collection of isolates for which matched susceptible and resistant isolates were not available. In this collection, mRNA levels could not be compared between susceptible and resistant isolates; they could only be normalized to the levels of mRNA for actin. Therefore, we are only comparing absolute levels in this collection, not increases within matched sets. The lack of correlation between resistance and levels of mRNA for *CDR1*, *CDR2*, *MDR1*, and *ERG11* in this collection is a lack of correlation of absolute levels and not a result of differences in levels of mRNA expression between susceptible and resistant isolates. Similarly, point mutations in the isolates could be associated with resistance, or they could be the result of allelic variations between different strains. *C. albicans* is known to reproduce predominantly by asexual fission (31) and to contain allelic differences, including balanced lethals, within each strain (38). In addition, several other factors contribute to resistance. Multiple molecular mechanisms have been associated with resistance, strains can differ in their levels of resistance to four different azole drugs, and each *C. albicans* strain reproduces clonally, resulting in highly unrelated strains. Therefore, it is difficult to make statistical arguments about correlations between molecular mechanisms and resistance phenotypes. Because of the differences associated with the analysis of mechanisms of resistance in matched sets of isolates and random isolates and because of the complexity of resistance in *C. albicans* strains, mechanisms of resistance identified in matched sets of isolates are not easily identified in random isolates.

Molecular tests based on our understanding of resistance mechanisms are unlikely to be sufficient to define clinical resistance in *C. albicans*. It may be important to consider alternate molecular tests that would monitor the general health or status of the fungal cell in the presence of drug in a shorter time than the present MIC determinations allow. Such tests might be able to accurately identify a resistant fungal isolate quickly, without the need for detailed knowledge about the exact molecular mechanisms of resistance.

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#### REFERENCES

1. Balan, I., A. M. Alarco, and M. Raymond. 1997. The *Candida albicans* *CDR3* gene codes for an opaque-phase ABC transporter. *J. Bacteriol.* **179**:7210–7218.
2. Ben-Yaacov, R., S. Knoller, G. A. Caldwell, J. M. Becker, and Y. Koltin. 1994. *Candida albicans* gene encoding resistance to benomyl and methotrex-

- ate is a multidrug resistance gene. *Antimicrob. Agents Chemother.* **38**:648–652.
3. Calabrese, D., J. Bille, and D. Sanglard. 2000. A novel multidrug efflux transporter gene of the major facilitator superfamily from *Candida albicans* (FLU1) conferring resistance to fluconazole. *Microbiology (United Kingdom)* **11**:2743–2754.
4. Clark, F. S., T. Parkinson, C. A. Hitchcock, and N. A. R. Gow. 1996. Correlation between rhodamine 123 accumulation and azole sensitivity in *Candida* species: possible role for drug efflux in drug resistance. *Antimicrob. Agents Chemother.* **40**:419–425.
5. Fling, M. E., J. Kopf, A. Tamarkin, J. A. Gorman, H. A. Smith, and Y. Koltin. 1991. Analysis of a *Candida albicans* gene that encodes a novel mechanism for resistance to benomyl and methotrexate. *Mol. Gen. Genet.* **227**:318–329.
6. Franz, R., S. Michel, and J. Morschhauser. 1998. A fourth gene from the *Candida albicans* *CDR* family of ABC transporters. *Gene* **220**:1–2.
7. Goldway, M., D. Teff, R. Schmidt, A. B. Oppenheim, and Y. Koltin. 1995. Multidrug resistance in *Candida albicans*: disruption of the *BEN<sup>r</sup>* gene. *Antimicrob. Agents Chemother.* **39**:422–426.
8. Hoffman, C. S., and F. Winston. 1987. A ten-minute DNA preparation from yeast efficiently releases autonomous plasmids for transformation of *Escherichia coli*. *Gene* **57**:267–272.
9. Holmberg, K., and D. A. Stevens. 1999. Resistance to antifungal drugs: current status and clinical implications. *Curr. Opin. Anti-Infective Investig. Drugs* **1**:306–317.
10. Lai, M. H., and D. R. Kirsch. 1989. Nucleotide sequence of cytochrome P450 L1A1 (lanosterol 14 alpha-demethylase) from *Candida albicans*. *Nucleic Acids Res.* **17**:804.
11. Lamb, D. C., D. E. Kelly, T. C. White, and S. L. Kelly. 2000. The R467K amino acid substitution in *Candida albicans* sterol 14 $\alpha$ -demethylase causes drug resistance through reduced affinity. *Antimicrob. Agents Chemother.* **44**:63–67.
12. Law, D., C. B. Moore, H. M. Wardle, L. A. Ganguli, M. G. Keane, and D. W. Denning. 1994. High prevalence of antifungal resistance in *Candida* spp. from patients with AIDS. *J. Antimicrob. Chemother.* **34**:659–668.
13. Loffler, J., S. L. Kelly, H. Hebart, U. Schumacher, C. Lass Florl, and H. Einsele. 1997. Molecular analysis of *cyp51* from fluconazole-resistant *Candida albicans* strains. *FEMS Microbiol. Lett.* **151**:263–268.
14. Lopez Ribot, J. L., R. K. McAtee, L. N. Lee, W. R. Kirkpatrick, T. C. White, D. Sanglard, and T. F. Patterson. 1998. Distinct patterns of gene expression associated with development of fluconazole resistance in serial *Candida albicans* isolates from human immunodeficiency virus-infected patients with oropharyngeal candidiasis. *Antimicrob. Agents Chemother.* **42**:2932–2937.
15. Losberger, C., and J. F. Ernst. 1989. Sequence of the *Candida albicans* gene encoding actin. *Nucleic Acids Res.* **17**:9488.
16. Lyons, C. N., and T. C. White. 2000. Transcriptional analyses of antifungal drug resistance in *Candida albicans*. *Antimicrob. Agents Chemother.* **44**:2296–2303.
17. Maniatis, T., E. F. Fritsch, and J. Sambrook. 1982. Molecular cloning: a laboratory manual. Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y.
18. Marichal, P., L. Koymans, S. Willemsens, D. Bellens, P. Verhasselt, W. Luyten, M. Borgers, F. C. S. Ramaekers, F. C. Odds, and H. Vanden Bossche. 1999. Contribution of mutations in the cytochrome P450 14 alpha-demethylase (Erg11p, Cyp51p) to azole resistance in *Candida albicans*. *Microbiology (United Kingdom)* **10**:2701–2713.
19. Marr, K. A., C. N. Lyons, T. R. Rustad, R. A. Bowden, and T. C. White. 1998. Rapid, transient fluconazole resistance in *Candida albicans* is associated with increased mRNA levels of *CDR*. *Antimicrob. Agents Chemother.* **42**:2584–2589.
20. Marr, K. A., T. C. White, J. A. H. van Burik, and R. A. Bowden. 1997. Development of fluconazole resistance in *Candida albicans* causing disseminated infection in a patient undergoing marrow transplantation. *Clin. Infect. Dis.* **25**:908–910.
21. McCullough, M. J., K. V. Clemons, and D. A. Stevens. 1999. Molecular and phenotypic characterization of genotypic *Candida albicans* subgroups and comparison with *Candida dubliniensis* and *Candida stellatoidea*. *J. Clin. Microbiol.* **37**:417–421.
22. Mori, T., M. Matsumura, Y. Kanamaru, S. Miyano, T. Hishikawa, S. Irie, K. Oshimi, T. Saikawa, and T. Oguri. 1997. Myelofibrosis complicated by infection due to *Candida albicans*: emergence of resistance to antifungal agents during therapy. *Clin. Infect. Dis.* **25**:1470–1471.
23. National Committee for Clinical Laboratory Standards. 1997. Reference method for broth dilution antifungal susceptibility testing of yeasts. Approved standard. NCCLS document M27-A. National Committee for Clinical Laboratory Standards, Wayne, Pa.
24. Nolte, F. S., T. Parkinson, D. J. Falconer, S. Dix, J. Williams, C. Gilmore, R. Geller, and J. R. Wingard. 1997. Isolation and characterization of fluconazole- and amphotericin B-resistant *Candida albicans* from blood of two patients with leukemia. *Antimicrob. Agents Chemother.* **41**:196–199.
25. Odds, F. C. 1988. *Candida* and Candidosis: a review and bibliography. Bailliere Tindall, London, United Kingdom.
26. Pelletier, R., J. Peter, C. Antin, C. Gonzalez, L. Wood, and T. J. Walsh. 2000.

- Emergence of resistance of *Candida albicans* to clotrimazole in human immunodeficiency virus-infected children: in vitro and clinical correlations. *J. Clin. Microbiol.* **38**:1563–1568.
27. Perea, S., J. L. Lopez Ribot, W. R. Kirkpatrick, R. K. McAtee, R. A. Santillan, M. Martinez, D. Calabrese, D. Sanglard, and T. F. Patterson. 2001. Prevalence of molecular mechanisms of resistance to azole antifungal agents in *Candida albicans* strains displaying high-level fluconazole resistance isolated from human immunodeficiency virus-infected patients. *Antimicrob. Agents Chemother.* **45**:2676–2684.
  28. Perepnikhatka, V., F. J. Fischer, M. Niimi, R. A. Baker, R. D. Cannon, Y. K. Wang, F. Sherman, and E. Rustchenko. 1999. Specific chromosome alterations in fluconazole-resistant mutants of *Candida albicans*. *J. Bacteriol.* **181**:4041–4049.
  29. Pfaller, M. A., C. J. Rhine, S. W. Redding, J. Smith, G. Farinacci, A. W. Fothergill, and M. G. Rinaldi. 1994. Variations in fluconazole susceptibility and electrophoretic karyotype among oral isolates of *Candida albicans* from patients with AIDS and oral candidiasis. *J. Clin. Microbiol.* **32**:59–64.
  30. Prasad, R., W. P. De, A. Goffeau, and E. Balzi. 1995. Molecular cloning and characterization of a novel gene of *Candida albicans*, *CDR1*, conferring multiple resistance to drugs and antifungals. *Curr. Genet.* **27**:320–329.
  31. Pujol, C., J. Reynes, F. Renaud, M. Raymond, M. Tibayrenc, F. J. Ayala, F. Janbon, M. Malli'e, and J. M. Bastide. 1993. The yeast *Candida albicans* has a clonal mode of reproduction in a population of infected human immunodeficiency virus-positive patients. *Proc. Natl. Acad. Sci. USA* **90**:9456–9459.
  32. Redding, S., J. Smith, G. Farinacci, M. Rinaldi, A. Fothergill, C. J. Rhine, and M. Pfaller. 1994. Resistance of *Candida albicans* to fluconazole during treatment of oropharyngeal candidiasis in a patient with AIDS: documentation by *in vitro* susceptibility testing and DNA subtype analysis. *Clin. Infect. Dis.* **18**:240–242.
  33. Rex, J. H., M. A. Pfaller, J. N. Galgiani, M. S. Bartlett, A. Espinel-Ingroff, M. A. Ghannoum, M. Lancaster, F. C. Odds, M. G. Rinaldi, T. J. Walsh, and A. L. Barry. 1997. Development of interpretive breakpoints for antifungal susceptibility testing: conceptual framework and analysis of *in vitro/in vivo* correlation data for fluconazole, itraconazole, and *Candida* infections. *Clin. Infect. Dis.* **24**:235–247.
  34. Sambrook, J., E. F. Fritsch, and T. Maniatis. 1989. *Molecular cloning: a laboratory manual*, 2nd ed. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.
  35. Sanglard, D., F. Ischer, L. Koymans, and J. Bille. 1998. Amino acid substitutions in the cytochrome P-450 lanosterol 14 $\alpha$ -demethylase (CYP51A1) from azole-resistant *Candida albicans* clinical isolates contribute to resistance to azole antifungal agents. *Antimicrob. Agents Chemother.* **42**:241–253.
  36. Sanglard, D., F. Ischer, M. Monod, and J. Bille. 1997. Cloning of *Candida albicans* genes conferring resistance to azole antifungal agents: characterization of *CDR2*, a new multidrug ABC transporter gene. *Microbiology* **143**:405–416.
  37. Sanglard, D., K. Kuchler, F. Ischer, J. L. Pagani, M. Monod, and J. Bille. 1995. Mechanisms of resistance to azole antifungal agents in *Candida albicans* isolates from AIDS patients involve specific multidrug transporters. *Antimicrob. Agents Chemother.* **39**:2378–2386.
  38. Whelan, W. L., and D. R. Soll. 1982. Mitotic recombination in *Candida albicans*: recessive lethal alleles linked to a gene required for methionine biosynthesis. *Mol. Gen. Genet.* **187**:477–485.
  39. White, T. C. 1997. Increased mRNA levels of *ERG16*, *CDR*, and *MDR1* correlate with increases in azole resistance in *Candida albicans* isolates from a patient infected with human immunodeficiency virus. *Antimicrob. Agents Chemother.* **41**:1482–1487.
  40. White, T. C. 1997. The presence of an R467K amino acid substitution and loss of allelic variation correlate with an azole-resistant lanosterol 14 $\alpha$ -demethylase in *Candida albicans*. *Antimicrob. Agents Chemother.* **41**:1488–1494.
  41. White, T. C., K. A. Marr, and R. A. Bowden. 1998. Clinical, cellular, and molecular factors that contribute to antifungal drug resistance. *Clin. Microbiol. Rev.* **11**:382–402.
  42. White, T. C., M. A. Pfaller, R. G. Rinaldi, J. Smith, and S. W. Redding. 1997. Stable azole drug resistance associated with a substrain of *Candida albicans* from an HIV-infected patient. *Oral Dis.* **3**:S102–S109.