

## $\beta$ -Lactamases of *Kluyvera ascorbata*, Probable Progenitors of Some Plasmid-Encoded CTX-M Types

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Received 24 July 2001/Returned for modification 28 October 2001/Accepted 7 April 2002

***Kluyvera ascorbata* produces a  $\beta$ -lactamase that results in an atypical susceptibility pattern, including low-level resistance to penicillins, cephalothin, and cefuroxime, but this resistance is reversed by clavulanate. Ten nucleotide sequences of the corresponding gene, *bla*<sub>KLUA</sub>, were obtained and were found to have minor variations (96 to 100%). Otherwise, *bla*<sub>KLUA</sub> was found to be similar (95 to 100%) to some plasmid-encoded CTX-M-type  $\beta$ -lactamases. Finally, mobilization of *bla*<sub>KLUA</sub> on a plasmid was found to be mediated probably by a genetic mobile element like *ISEc1*.**

Resistance to oxyimino- $\beta$ -lactams such as cefotaxime, ceftazidime, or ceftriaxone in gram-negative rods has commonly been associated with the expression of acquired  $\beta$ -lactamases. Most of them are class A extended-spectrum  $\beta$ -lactamase (ESBL) derivatives of the TEM-1, TEM-2, or SHV-1 enzyme (4). Since the 1990s, a new step in resistance to  $\beta$ -lactams was discovered with the demonstration of the mobilization of a chromosomal  $\beta$ -lactamase on a plasmid, allowing the wide dissemination of  $\beta$ -lactamases and conferring the potential for epidemic problems. Many plasmid-mediated AmpC  $\beta$ -lactamases have been described, such as MIR-1 (*Enterobacter cloacae*), CMY-2 (*Citrobacter freundii*), DHA-1 (*Morganella morganii*), and ACC-1 (*Hafnia alvei*) (20). More recently, the discovery of SFO-1 suggested a probable mobilization from the chromosome of *Serratia fonticola* on a plasmid harbored in an *E. cloacae* isolate in Japan (16). To date, that was the only report of the mobilization of a chromosomally mediated class A  $\beta$ -lactamase.

The CTX-M-type  $\beta$ -lactamases (CTX-M-1 to CTX-M-15, Toho-1 and Toho-2, and UOE-1 and UOE-2), encoded by transferable plasmids, constitute a novel group of class A ESBLs whose origins are still unknown (2, 10, 11, 13, 15, 18, 21, 23). These CTX-M-type enzymes are not closely related to TEM or SHV ESBLs but share extensive similarities (70 to 75%) with the chromosomal *Klebsiella oxytoca*  $\beta$ -lactamases (23). Nevertheless, *Kluyvera ascorbata*, a species of the family *Enterobacteriaceae* that is rarely detected in medical practice (7, 24), was suggested as another probable progenitor because of the particular susceptibility patterns of some clinical isolates including resistance to cefotaxime and aztreonam but susceptibility to ceftazidime and the high potentiation effect of the reversal of resistance by clavulanate (D. Bertei et al., Abstr. 14th Réunion Interdisciplin. Chimiothér. Anti-Infect., abstr. 199, p. 10, 1994).

**Bacterial strains.** Ten nonduplicate strains of *K. ascorbata* including the type strain were supplied by the Institut Pasteur Collection (Paris, France), and two strains were isolated at Tenon and Saint-Louis Hospitals (Paris, France), respectively. Each strain was identified on the basis of its behavior in culture and its biochemical characteristics by using the API 20E system (bioMérieux, Marcy-l'Étoile, France) and by using Biotype-100 carbon source utilization strips (bioMérieux) and Recognizer software (Taxolab, Institut Pasteur, Paris, France).

**Antimicrobial agents and susceptibility testing.** The patterns of susceptibility to antibiotics usually active against members of the family *Enterobacteriaceae* were determined by the disk diffusion method, as described previously (8).

The MICs of 12  $\beta$ -lactams (Table 1) including penicillins and cephalosporins in the presence or absence of  $\beta$ -lactamase inhibitors (clavulanate at 2  $\mu$ g/ml or tazobactam at 4  $\mu$ g/ml) were determined by a dilution method on Mueller-Hinton agar (Sanofi Diagnostics Pasteur, Marnes La Coquette, France) with an inoculum of  $10^5$  CFU/spot applied with a multi-inoculation device (Multipoint Inoculator A400; Denleytech, Woking, United Kingdom) (8).

All strains tested a low level of resistance to penicillins (amoxicillin, ticarcillin, piperacillin) but also displayed a low level of resistance to cephalothin and cefuroxime (Table 1). Significant synergy with clavulanate was observed for amoxicillin and ticarcillin (32- to 512-fold). A low level of synergy was also observed between piperacillin and tazobactam, which correlated with the high level of activity of the ureidopenicillin piperacillin. Finally, all strains tested were highly susceptible to extended-spectrum cephalosporins, moxalactam, cefoxitin, aztreonam, and imipenem.

**Kinetic and IEF analyses.** Subsequently, the resistance pattern was correlated with  $\beta$ -lactamase production. The  $\beta$ -lactamase contents were obtained from sonicated extracts of bacteria cultured overnight at 37°C in Trypticase soy broth (bioMérieux), prepared as described previously (8). Analytical isoelectric focusing (IEF) was performed in a polyacrylamide gel as described previously by the iodometric procedure with benzylpenicillin for detection (8). We detected  $\beta$ -lactamases

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TABLE 1. MICs of 12  $\beta$ -lactams for the 12 clinical *K. ascorbata* isolates, recipient strain *E. coli* XL-1, and the *E. coli* XL-1 transformant producing KLUA-1

$\beta$ -Lactam(s) <sup>a</sup>	No. of isolates for which MIC ( $\mu$ g/ml) was:													MIC ( $\mu$ g/ml) for <i>E. coli</i> XL-1 recipient	
	<0.03	0.06	0.125	0.25	0.5	1	2	4	8	16	32	64	128	R+	R-
Amoxicillin						1	1	4	4	2				>128	4
Amoxicillin + CA			3 <sup>b</sup>	3	4	2								1	1
Ticarcillin										2	3	4	3	>128	0.5
Ticarcillin + CA			5 <sup>b</sup>	2	4	1								4	0.125
Piperacillin					1	4	4	3						128	1
Piperacillin + TZ			4 <sup>b</sup>	3	3	1	1							2	0.12
Cephalothin								7	3					>128	0.5
Cefuroxime							1	6	5					64	1
Cefoxitin				1	4	4	3							2	1
Cefotaxime	3	7	2											128	<0.03
Ceftriaxone	2	6	4											14	<0.03
Ceftazidime	6	5	1											4	0.06
Cefepime	9	2	1											4	<0.03
Aztreonam	4	6	2											8	0.12
Imipenem		1	6	5										0.25	<0.03

<sup>a</sup> CA, 2  $\mu$ g of clavulanic acid per ml; TZ, 4  $\mu$ g of tazobactam per ml.

<sup>b</sup> MIC, <0.125  $\mu$ g/ml.

<sup>c</sup> R-, *E. coli* recipient; R+, *E. coli* with recombinant plasmid producing KLUA-1.

with different pIs, as follows: pI 6.9, two strains; pI 8.0, six strains; pI 8.4, four strains (see Table 3).

Finally, the substrate and inhibition profiles of a single crude extract (type strain CIP 82.95T) were determined. This extract was produced from 4 liters of culture grown in brain heart infusion (Difco, Detroit, Mich.) and incubated at 37°C for 5 h after initial inoculation of 300 ml of a starter culture. Cells were harvested by centrifugation at 5,800  $\times$  g for 30 min at 4°C. The pellets (about 20 g) were washed by resuspension in 40 ml of ice-cold 0.1 M NaCl plus 0.05 M sodium azide and centrifuged as described above. The washed pellet was resuspended in 40 ml of the same solution and lysed by ultrasound treatment (three times for 10 s each time per gram of pellets; power = 50 W and frequency = 20 kHz with type 20-200 equipment [Alcatel, Paris, France]). The crude extract was cleared by centrifugation at 48,000  $\times$  g for 30 min at 4°C. Nucleic acids were precipitated by adding 0.2 M spermine (Sigma, Saint-Quentin Fallavier, France) and centrifugation (48,000  $\times$  g for 30 min at 4°C). The crude extract was then subjected to chromatography on a Sephadex G100 column. The active fractions were pooled, subjected to chromatography on Bio-Rex 70 resin, and then concentrated by ultrafiltration.

The kinetic constants  $V_{\max}$ , which was expressed relative to the  $V_{\max}$  for benzylpenicillin (which was set at 100%), and  $K_m$  were determined by computerized microacidimetric assay in 0.1 M NaCl at pH 7.0 and 37°C (14). One unit of  $\beta$ -lactamase was defined as the amount of enzyme that hydrolyzed 1  $\mu$ mol of benzylpenicillin per min at pH 7.0 and 37°C. Kinetic analysis showed that the activity of the enzyme was highest with cephalothin, but that cefuroxime and cefotaxime were also good substrates (Table 2). Conversely, the enzyme displayed a low level of activity with ceftazidime, aztreonam, and cephamycin. Tazobactam was the most efficient inhibitor (50% inhibitory concentration [IC<sub>50</sub>], 10 nM), closely followed by clavulanic acid (IC<sub>50</sub>, 20 nM). Sulbactam was the least potent inhibitor (IC<sub>50</sub>, 100 nM). Finally, chloride ions at a concentration of 0.5 M had no inhibitory effect.

**Genetic characterization of *bla*<sub>KLUA</sub>.** The identification of only one susceptibility pattern among the 12 strains of *K. ascorbata* tested suggested that this species possesses a naturally occurring, chromosomally mediated  $\beta$ -lactamase. Furthermore, this assumption was confirmed by the failure of both conjugation experiments and plasmid extraction with the High Pure plasmid isolation kit (Roche Biochemicals, Neuilly-sur-Seine, France).

On the basis of the observed pattern of susceptibility and enzymatic properties, we designed degenerate oligonucleotide primers from the sequences of genes encoding CTX-M-type enzymes (primer MA1-forward [5'-SCSATGTGCAGYACCA GTAA-3'] and primer MA2-reverse [5'-CCGCRATATGRTT GGTGGTG-3']) and used them to amplify an internal fragment of the *K. ascorbata bla* genes. In each PCR, the cycling conditions were as follows: 35 cycles of 60 s at 94°C for denaturation, 30 s at 60°C for annealing, and 30 s at 72°C for elongation with 2 U of *Taq* DNA polymerase (Perkin-Elmer, Foster City, Calif.). Amplifications were obtained for all *K. ascorbata* strains producing a  $\beta$ -lactamase of pI 8.0 or 8.4. Nevertheless, the two *bla* genes encoding  $\beta$ -lactamase of pI 6.9

TABLE 2. Kinetic constants ( $V_{\max}$ ,  $K_m$ ) of  $\beta$ -lactamase produced by *K. ascorbata* strain CIP 82.95T<sup>a</sup>

Substrate	$V_{\max}$ (relative)	$K_m$ ( $\mu$ M)	$V_{\max}/K_m$ (relative)
Benzylpenicillin	100	17.5	100
Ticarcillin	7.5	10	13
Cephalothin	1,100	141	136
Cefuroxime	80	35	40
Cefotaxime	110	375	5.1
Ceftazidime	<0.1	50	<0.04
Aztreonam	0.3	105	0.06
Cefoxitin	<0.1	110	<0.02

<sup>a</sup>  $V_{\max}$  was expressed relative to the  $V_{\max}$  for benzylpenicillin (which was set at 100%).

TABLE 3. Genotypic characterization of β-lactamase contents of *K. ascorbata* strains and β-lactamase pIs

Strain	pI	Result of PCR with primer(s):			Gene	Genbank accession no.
		MA1-MA2 (208–751 bp; 543 bp)	MA3 (1–430 bp; 429 bp)	MA4 (461–883 bp; 422 bp)		
CIP 82.95T	8.0	+	+	+	<i>bla</i> <sub>KLUA-1</sub>	AJ272538
IP 15.79	8.4	+	+	+	<i>bla</i> <sub>KLUA-2</sub>	AJ251722
IP 5.79	8.0	+	+	+	<i>bla</i> <sub>KLUA-3</sub>	AJ427461
IP 8.79	8.0	+	+	+	<i>bla</i> <sub>KLUA-4</sub>	AJ427462
IP 12.79	8.0	+	+	+	<i>bla</i> <sub>KLUA-5</sub>	AJ427463
SL Ka99	6.9	–	–	–		
IP 2.89	6.9	–	–	–		
IP 3.89	8.0	+	+	+	<i>bla</i> <sub>KLUA-8</sub>	AJ427465
TN Ka01	8.4	+	+	+	<i>bla</i> <sub>KLUA-9</sub>	AJ427466
IP 43.50	8.0	+	+	+	<i>bla</i> <sub>KLUA-10</sub>	AJ427467
IP 4450.94	8.4	+	+	+	<i>bla</i> <sub>KLUA-11</sub>	AJ427468
IP 684.74	8.4	+	+	+	<i>bla</i> <sub>KLUA-12</sub>	AJ427469

were not amplified with these CTX-M-specific primers (Table 3).

The PCR products were then purified with Qiagen (Courtaubeuf, France) columns, and the DNA sequences were determined twice on both strands by the procedure of Sanger et al. (22) by using fluorescent dye-labeled dideoxynucleotides, thermal cycling with *Taq* polymerase, and an ABI 373A DNA sequencer (Applied Biosystems, Foster City, Calif.). All PCR products consisted of a 540-bp fragment, which was found to be highly similar (>97% identity) to the cluster that included the *bla*<sub>CTX-M-2</sub>, *bla*<sub>CTX-M-4</sub>, *bla*<sub>CTX-M-5</sub>, *bla*<sub>CTX-M-6</sub>, *bla*<sub>CTX-M-7</sub>, and *bla*<sub>Toho-1</sub> genes.

Finally, other primers more specific for the *bla*<sub>CTX-M</sub> gene of this cluster were used to amplify the entire *Kluyvera bla* gene: primers M3-forward (5'-ATGATGACTCAGAGCATTCGC-3') and M3-reverse (5'-GGGCAATCAGCTTATTCATGG-3') and primers M4-forward (5'-TTGCTCGCTCGTTGGGTGAT-3') and M4-reverse (5'-TATTGCATCAGAAACCGTGGG-3') (Table 3). Amplifications with primer M3 or M4 were performed by using the same cycling conditions described above. The nucleotide sequences of *K. ascorbata* genes encoding a β-lactamase with a pI of 8.0 or 8.4 (*bla*<sub>KLUA-1</sub> to *bla*<sub>KLUA-5</sub> and *bla*<sub>KLUA-8</sub> to *bla*<sub>KLUA-12</sub>) were determined (Table 3). High degrees of homology (>95%) were observed among all these *bla*<sub>KLUA</sub> genes coding for class A β-lactamases. Moreover, *bla*<sub>KLUA</sub> genes were also found to share extensive similarities (95 to 100%) with the genes producing the CTX-M-2, CTX-M-4, CTX-M-5, CTX-M-6, CTX-M-7,

and Toho-1 β-lactamases; but they displayed lower levels of identity with the genes for the other CTX-M types (68 to 84%) (Table 4). However, this high level of similarity observed suggested that the chromosome-encoded β-lactamase of *K. ascorbata* is a more probable progenitor of at least some plasmid-encoded CTX-M enzymes than *K. oxytoca* (23). Finally, the highest degrees of homology between *bla*<sub>KLUA</sub> and chromosome-encoded *bla* genes were observed for class A enzymes produced by *Kluyvera cryocrescens* (≈77%), *S. fonticola* (≈74%), *K. oxytoca* (≈73%), and *Citrobacter sedlakii* (≈73%) (6, 9, 19; C. Humeniuk, unpublished data).

Only two *bla* genes encoding a β-lactamase of pI 6.9 were not amplified with the CTX-M-specific primers (Table 3). Characterization of a β-lactamase(s) produced by both of these strains is in progress. In fact, it appears that *K. ascorbata* could produce at least two different kinds of β-lactamase genes, as has been observed for *K. oxytoca*, in which two main groups of β-lactamase genes have been distinguished (*bla*<sub>OXY-1</sub> and *bla*<sub>OXY-2</sub>; identity, 90%) (9). This heterogeneity of the *K. ascorbata* β-lactamases could be correlated with the diversity observed among CTX-M-type enzymes, which are divided into five clusters (Table 4).

**Genetic environment of *bla*<sub>KLUA-1</sub>.** As the chromosomal β-lactamase genes of *K. ascorbata* are the probable progenitors of some of the plasmid-encoded extended-spectrum CTX-M-type enzymes, we investigated the genetic organization of *bla*<sub>KLUA-1</sub> by cloning and sequencing the surrounding regions of this gene. Chromosomal DNA was prepared from *K. ascor-*

TABLE 4. Identities of amino acid sequences of *K. ascorbata* β-lactamases and CTX-M, Toho, UOE β-lactamases

Enzyme(s)	Range % identity (reference) with clusters including:					
	KLUA-1, -2, -3, -4, -5, -8, -9, -10, -11, and -12 (this work)	CTX-M-1, -3, -10, -11, and -12 and UOE-1 (13, 18, 23 <sup>a</sup> )	CTX-M-2, -4, -5, -6, and -7 and Toho-1 (3, 10, 11, 23)	CTX-M-9, -13, -14, and -15 and UOE-2 (21 <sup>b</sup> )	Toho-2 (15)	CTX-M-8 (2)
KLUA-1, -2, -3, -4, -5, -8, -9, -10, -11, and -12	96–100	77–82	95–100	78–81	68–71	82–84
CTX-M-1, -3, -10, -11, and -12 and UOE-1		98–100	77–81	79–80	70–71	80–82
CTX-M-2, -4, -5, -6, and -7 and Toho-1			97–100	78–81	68–71	81–84
CTX-M-9, -13, -14, and -15 and UOE-2				99–100	87–88	85–86
Toho-2					100	75
CTX-M-8						100

<sup>a</sup> Genbank references numbers AY005110 for CTX-M-11 and AY013478 for UOE-1.

<sup>b</sup> Genbank reference numbers AF252621 for CTX-M-13, AF252622 for CTX-M-14, AF252623 for CTX-M-15, and AF311345 for UOE-1.

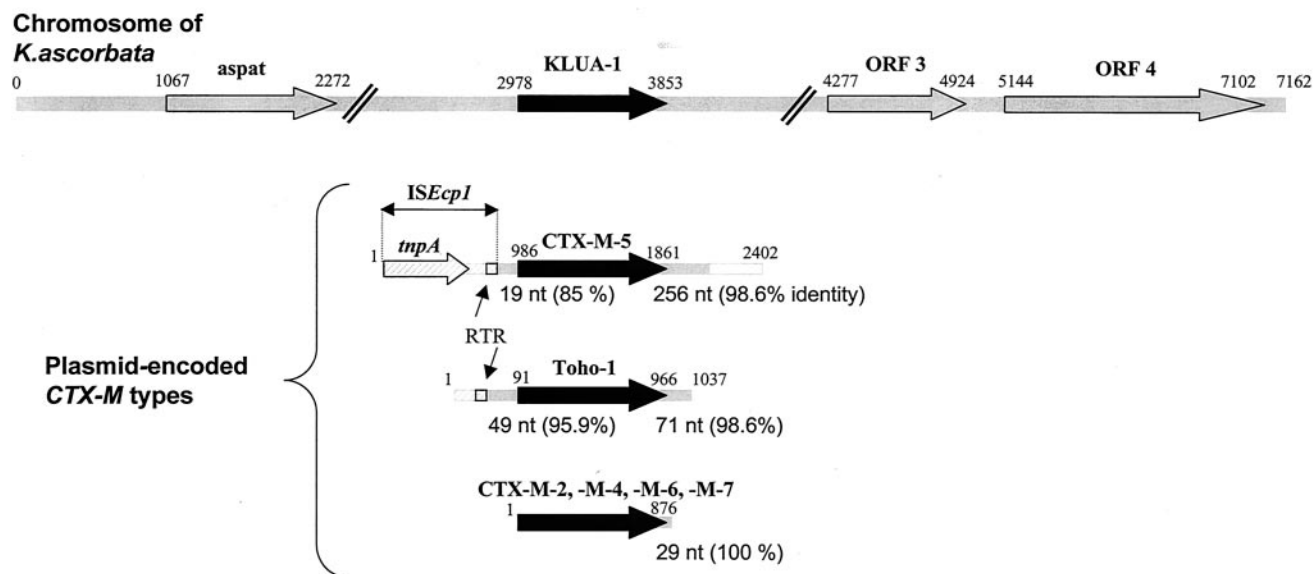


FIG. 1. Genetic environment of *bla*<sub>KLUA-1</sub> gene encoding the  $\beta$ -lactamase produced by *K. ascorbata* strain CIP 82.95T. The regions surrounding the chromosomal *bla*<sub>KLUA-1</sub> gene and the sequences adjacent to the plasmidic genes encoding  $\beta$ -lactamases CTX-M-2, CTX-M-4, CTX-M-5, CTX-M-6, CTX-M-7, and Toho-1 (2, 10, 11, 23) were compared. The RTR sequence and *tnpA* correspond to the right terminal repeat and transposase A, respectively, which are typical of *ISEcp1*.

*bata* CIP 82.95T, partially digested with *Sau3A*, and ligated into the *Bam*HI site of pBK-CMV (pBK Phagemid Vectors; Stratagene, La Jolla, Calif.), as described previously (17). The recombinant plasmid was introduced into *Escherichia coli* XL-1 by the standard CaCl<sub>2</sub> procedure. Several transformants were selected on Mueller-Hinton agar (Sanofi Diagnostics Pasteur) supplemented with amoxicillin (40  $\mu\text{g} \cdot \text{ml}^{-1}$ ) and kanamycin (25  $\mu\text{g} \cdot \text{ml}^{-1}$ ) and were further characterized by determination of their antibiotic susceptibility profiles and pIs. The molecular sizes of the inserts were estimated from the results of restriction digestion and electrophoresis in 1 to 3% agarose gels, as described previously (17). Finally, both strands of the DNA sequence of a 7-kb insert were determined. The BLASTN program on the National Center for Biotechnology Information website (revision date, 20 July 2001; <http://www.ncbi.nlm.nih.gov>) was used for database searches (1).

Three open reading frames (ORFs) were found immediately upstream (2,980 bp) and downstream (3,300 bp) of the *bla*<sub>KLUA</sub> gene (ORF 2) (Fig. 1). The protein corresponding to ORF 1 was 86% identical to an aspartate aminotransferase from *E. coli*. The deduced amino acid sequence of the protein encoded by ORF 3 was 43% identical to that of a putative protein from *E. coli*. The protein encoded by the fourth ORF showed various degrees of similarity (28 to 56%) to adhesive proteins (MisL, AidaI, VirG) known to contribute to the pathogenicities of several enterobacteria. Recently, the nucleotide sequence of *bla*<sub>KLUC</sub>, which encodes the chromosomal  $\beta$ -lactamase of *K. cryocrescens*, and its surrounding regions has been reported (6). Genes corresponding to an aspartate aminotransferase and a putative protein showing 94 and 66% identities with the proteins encoding by *K. ascorbata* ORF 1 and ORF 3, respectively, were found upstream and downstream of the *bla*<sub>KLUC</sub> gene. In fact, this particular genetic organization could be a characteristic of the genus *Kluyvera*.

Finally, no *ampR* gene was detected upstream of the *bla*<sub>KLUA-1</sub> gene (Fig. 1). Indeed, some class A  $\beta$ -lactamases closely related to *K. ascorbata* enzymes are regulated by an *ampR* gene: *cumR* (*Proteus vulgaris*) (5), *sedR* (*C. sedlakii*) (19) and *fonR* (*S. fonticola*) (C. Humeniuk, unpublished data).

**Arguments for mobilization of chromosomal *bla*<sub>KLUA</sub> gene.** The analysis of the sequences surrounding the *bla*<sub>CTX-M</sub> genes revealed the presence of sequences similar to those for CTX-M-5 on the *K. ascorbata* chromosome (5 nucleotides [nt] upstream and 179 nt downstream with 80 and 100% identities, respectively, with the chromosome of *K. ascorbata*), Toho-1 (49 nt upstream and 71 nt downstream with 95.9 and 98.6% identities, respectively), and CTX-M-2, -M-4, -M-6, and -M-7 (29 nt upstream with 100% identity) (Fig. 1). However, major differences between the adjacent sequences of the other *bla*<sub>CTX-M</sub> genes (CTX-M-1, -M-3, -M-8, -M-12, -M-13, -M-14, and -M-15; Toho-2; UOE-1; and UOE-2) and the chromosome of *K. ascorbata* were observed. This fact confirms that the  $\beta$ -lactamase of *K. ascorbata* is probably the progenitor of only some plasmid-encoded CTX-M-type enzymes. To date, this is the second report of the mobilization of a chromosomally mediated class A  $\beta$ -lactamase.

So far we have investigated the potential mechanism of mobilization of the *bla*<sub>KLUA</sub> gene by sequencing surrounding regions of the *bla*<sub>CTX-M-5</sub> gene located on plasmid pCLL3417 (3). The GenBank accession number for this nucleotide sequence is AJ286192. Sequencing confirmed the presence of sequences similar to that of the *K. ascorbata* chromosome (19 nt upstream and 256 nt downstream with 85 and 98.6% identities, respectively) and revealed the presence of the novel insertion sequence, *ISEcp1*, described by Stapleton (P. D. Stapleton, Abstr. 39th Intersci. Conf. Antimicrob. Agents Chemother., abstr. 1457, 1999) (Fig. 1). Subsequently, this element was reported for several plasmid-encoded AmpC enzymes includ-



ing CMY-5, LAT-1, and ACC-1, which were probably mobilized from chromosomal  $\beta$ -lactamases of *C. freundii* or *H. alvei* (20). It was also detected on plasmids carrying other *bla*<sub>CTX-M</sub> genes (CTX-M-3-like, CTX-M-14, CTX-M-15, Toho-1, Toho-2, and UOE-2) (11, 12, 15; GenBank accession numbers AF252622, AF252623, and AF311345).

Given the low level of clinical importance of *K. ascorbata* (7, 24), it is important to consider the possible transfer of the *bla* gene in veterinary practice or in the environment. Several preliminary reports have indeed described *Salmonella enterica* serovar Typhimurium as producing  $\beta$ -lactamases of the CTX-M2, CTX-M-4, CTX-M-5, and CTX-M-7 types in various countries (Argentina, Latvia, Russia, Greece) (3, 10, 23). If *K. ascorbata* acts as a reservoir for these plasmid-encoded CTX-M enzymes, the sequences of more chromosomal genes must be determined to confirm the existence of a single progenitor due to the heterogeneity observed in the CTX-M cluster (Table 4).

**Nucleotide sequence accession number.** The GenBank accession number for the nucleotide sequence of the insert reported in this paper is AJ272538.

This work was financed by a grant for "Réseau  $\beta$ -lactamases: de la clinique à la recherche" from the Ministère de l'Éducation Nationale, de la Recherche et de la Technologie, Paris, France.

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