

Tigecycline Resistance Can Occur Independently of the *ramA* Gene in *Klebsiella pneumoniae*

Mark Veleba and Thamarai Schneiders

Centre for Infection and Immunity, Queen's University Belfast, Medical Biology Centre, Belfast, United Kingdom

Tigecycline resistance in *Klebsiella pneumoniae* results from *ramA* upregulation that causes the overexpression of the efflux pump, AcrAB-TolC. Tigecycline mutants, derived from *Ecl8ΔramA*, can exhibit a multidrug resistance phenotype due to increased transcription of the *marA*, *rara*, *acrAB*, and *oqxAB* genes. These findings support the idea that tigecycline or multidrug resistance in *K. pneumoniae*, first, is not solely dependent on the *ramA* gene, and second, can arise via alternative regulatory pathways in *K. pneumoniae*.

Tigecycline resistance in *Klebsiella pneumoniae* is on the increase, with reported cases in Greece (9), India (3), and Saudi Arabia (1), and its effectiveness as a therapeutic agent is uncertain, with patients treated with tigecycline showing persistent bacteraemia caused by *Escherichia coli*, *Acinetobacter baumannii*, and *K. pneumoniae* (2).

The AcrAB-TolC pump complex is a clinically relevant efflux system activated by several AraC-type transcriptional regulators such as MarA, SoxS, and RamA (7). Genetic studies have shown that resistance to tigecycline is mediated by the increased expression of the *ramA* gene, which subsequently results in the upregulation of the efflux pump *acrAB* in *K. pneumoniae* (10, 11) and *Enterobacter cloacae* (6). It has been shown that the increased expression of the *ramA* gene is linked to mutations within the cognate repressor, *ramR*, that is divergently transcribed from the *romA-ramA* genes (10). However, in a recent study (10), it was shown that high-level tigecycline resistance is exhibited in clinical strains of *K. pneumoniae* that do not overexpress *ramA* or *acrAB*; hence, we hypothesized that alternative pathways to tigecycline resistance must exist in *K. pneumoniae*. Accordingly, we searched the *Klebsiella* genome for AraC-type transcriptional regulators that had a size similar to those of the other multidrug resistance regulators such as *marA*, *soxS*, and *ramA*. The bioinformatic analyses of the genome of *Klebsiella* MGH78578 (NC_09648) located an additional regulator, which we have termed *rara* (Fig. 1). In an accompanying paper, we report the characterization of this novel multidrug-resistant (MDR) regulator (13), which, when overexpressed, exhibits a MDR phenotype independently of *marA-soxS-rob* and *ramA*. In *K. pneumoniae* and *Enterobacter* spp., the chromosomally encoded *rara* regulator lies downstream of the efflux pump, *oqxAB* (Fig. 1), which has been previously linked to decreased susceptibility to olaquinox, ciprofloxacin, and chloramphenicol (5). Interestingly, in *E. coli*, the gene locus encoding the efflux pump has been shown to be located on the pOLA52 plasmid in strains isolated from pig manure (12), where the locus is flanked

by two IS26 insertion sequences, highly suggestive of its genetic mobility.

Our research issue was whether alternative pathways to tigecycline resistance exist in *K. pneumoniae*. Accordingly, we used a genetically modified *Klebsiella pneumoniae* *Ecl8ΔramA* strain, with a markerless deletion of the *ramA* gene, in a tigecycline selection experiment. The markerless deletion of the *ramA* gene was constructed as described previously (8). Briefly, *Ecl8ΔramA* from an overnight culture was grown overnight at 37°C on agar plates containing tigecycline concentrations of 0.5 μg/ml, 1 μg/ml, 2 μg/ml, 4 μg/ml, and 8 μg/ml. The overnight culture was also diluted in phosphate-buffered saline (PBS) and grown on LB plates without antibiotic to establish baseline growth levels. After overnight incubation, we picked three mutants (TGC1-3) from the plate with 4 μg/ml for further analyses. Accordingly, we found that the mutational frequency of *Ecl8ΔramA* with respect to tigecycline was 0.466×10^{-6} . MIC testing (performed in triplicate) of tigecycline, ciprofloxacin, norfloxacin, tetracycline, and olaquinox was undertaken as described in the British Society for Antimicrobial Chemotherapy (BSAC) guidelines (4). Additionally, quantitative real-time PCR analyses were carried out to assess expression levels of genes *rara*, *oqxB*, *acrA*, *marA*, and *soxS* in the 3 mutant strains by the use of cDNA (generated by AffinityScript [Agilent]) and a Brilliant III kit (Agilent) for amplification. Experiments were conducted using a Stratagene Mx3005P system (Agi-

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Address correspondence to Thamarai Schneiders, t.schneiders@qub.ac.uk.

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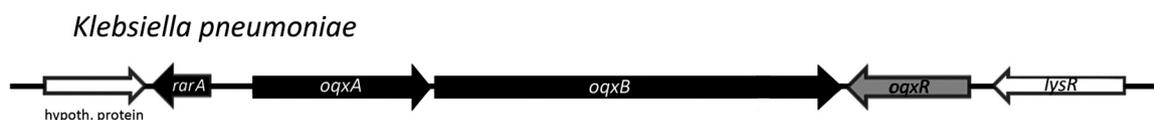


FIG 1 Organization of *rara* locus in *K. pneumoniae*. This genomic organization is conserved in *Enterobacter* sp. 638, *Serratia proteamaculans* 568, and *Enterobacter cloacae*.

TABLE 1 MIC and QPCR measurements of tigecycline mutants (TGC1, TGC3, and TGC5)

Strain	MIC ^a					Fold expression increase vs Ecl8ΔramA			
	NOR	CIP	TET	OQX	TIG	rara	oqxA	marA	acrB
Ecl8	0.250	0.031	4	16	2				
Ecl8ΔramA	0.063	0.016	2	8	1				
TGC1	8	0.25	16	32	16	4.59	6.06	6.06	55.72
TGC3	8	0.25	8	32	8	17.15	13.93	12.13	103.97
TGC5	8	0.25	8	16	8	2.14	3.03	5.28	27.86

^a NOR, norfloxacin; CIP, ciprofloxacin; TET, tetracycline; OQX, olaquinox; TIG, tigecycline.

lent) and analyzed using MxPro software (Agilent). MIC susceptibility testing demonstrated that all 3 clones exhibited reduced susceptibility to the antibiotics tested in comparison to the parental strain, Ecl8ΔramA (see Table 1). We also found that the transcriptional levels of *rara* (4.59-fold, 17.15-fold, and 2.14-fold, respectively) and *marA* (6.06-fold, 12.13-fold, and 5.28-fold) as well as those of the efflux operons *oqxAB* (6.06-fold, 13.93-fold, and 3.03-fold) and *acrAB* (55.72-fold, 103.97-fold, and 27.86-fold) were higher than the expression levels seen with the parental strain (Ecl8ΔramA) (see Table 1). Of note, *soxS* levels were found to be unaltered in comparison to those seen with the parental strain (Ecl8ΔramA).

Our work shows that tigecycline exposure to Ecl8ΔramA can generate mutants that exhibit low-level multidrug resistance. However, in order to pinpoint the individual contributions of *rara*, *marA*, and *oqxAB*, individual gene deletions are essential. We surmise that both *rara* and *marA* provide alternative pathways for the emergence of multidrug resistance in *K. pneumoniae* in the absence of the *ramA* gene. Additionally, we demonstrate that both the *acrAB* and the newly described *oqxAB* efflux pump can contribute to the MDR phenotype. From our findings, it is evident that a functioning *ramA* gene is not always needed to confer tigecycline resistance in *K. pneumoniae*.

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REFERENCES

- Al-Qadheeb NS, Althawadi S, Alkhalaf A, Hosaini S, Alrajhi AA. 2010. Evolution of tigecycline resistance in *Klebsiella pneumoniae* in a single patient. *Ann. Saudi Med.* 30:404–407.
- Anthony KB, et al. 2008. Clinical and microbiological outcomes of serious infections with multidrug-resistant gram-negative organisms treated with tigecycline. *Clin. Infect. Dis.* 46:567–570.
- Arya SC, Agarwal N. 2010. Emergence of tigecycline resistance amongst multi-drug resistant gram negative isolates in a multi-disciplinary hospital. *J. Infect.* 61:358–359.
- Hamilton-Miller JMT. 1999. New BSAC sensitivity testing guidelines. *J. Antimicrob. Chemother.* 44:850–851.
- Hansen LH, Jensen LB, Sorensen HI, Sorensen SJ. 2007. Substrate specificity of the OqxAB multidrug resistance pump in *Escherichia coli* and selected enteric bacteria. *J. Antimicrob. Chemother.* 60:145–147.
- Hornsey M, et al. 2010. Emergence of AcrAB-mediated tigecycline resistance in a clinical isolate of *Enterobacter cloacae* during ciprofloxacin treatment. *Int. J. Antimicrob. Agents* 35:478–481.
- Koutsolioutsou A, Martins EA, White DG, Levy SB, Demple B. 2001. A *soxRS*-constitutive mutation contributing to antibiotic resistance in a clinical isolate of *Salmonella enterica* (serovar Typhimurium). *Antimicrob. Agents Chemother.* 45:38–43.
- Merlin C, McAteer S, Masters M. 2002. Tools for characterization of *Escherichia coli* genes of unknown function. *J. Bacteriol.* 184:4573–4581.
- Neonakis IK, Stylianou K, Daphnis E, Maraki S. 2011. First case of resistance to tigecycline by *Klebsiella pneumoniae* in a European University Hospital. *Indian J. Med. Microbiol.* 29:78–79.
- Rosenblum R, Khan E, Gonzalez G, Hasan R, Schneiders T. 2011. Genetic regulation of the *ramA* locus and its expression in clinical isolates of *Klebsiella pneumoniae*. *Int. J. Antimicrob. Agents* 38:39–45.
- Ruzin A, Visalli MA, Keeney D, Bradford PA. 2005. Influence of transcriptional activator RamA on expression of multidrug efflux pump AcrAB and tigecycline susceptibility in *Klebsiella pneumoniae*. *Antimicrob. Agents Chemother.* 49:1017–1022.
- Sorensen AH, Hansen LH, Johannesen E, Sorensen SJ. 2003. Conjugative plasmid conferring resistance to olaquinox. *Antimicrob. Agents Chemother.* 47:798–799.
- Veleba M, Higgins PG, Gonzalez G, Seifert H, Schneiders T. 2012. Characterization of Rara, a novel AraC family multidrug resistance regulator in *Klebsiella pneumoniae*. *Antimicrob. Agents Chemother.* 56:4450–4458.