V
cisceral leishmaniasis (VL), caused by the protozoan parasite
Leishmania donovani, results in 0.2 to 0.4 million cases re-
ported annually and poses a threat to about 200 million people
worldwide. With the advent of the human immunodeficiency vi-
ruses, VL has surged as an opportunistic infection among AIDS
patients (1). Pentavalent antimonials have been the first-line drugs
against all forms of leishmaniasis, although their use for the treat-
ment of VL has been discontinued in India due to the emergence
of resistance. Extended treatment regimens, parenteral adminis-
tration, and toxic side effects limit patient compliance with treat-
ment. Frequent therapeutic failures due to resistance to the anti-
monial drug sodium stibogluconate (SSG) necessitate the use of
second-line drugs such as amphotericin B (AMB), which are more
toxic. Alternate therapies for VL have resulted in the development
of several lipid- and liposome-based formulations of conventional
drugs, with enhanced efficacies and reduced toxicities (2–7). The
advantages of the use of liposomes as drug delivery vehicles for
treating macrophage-resident parasites such as Leishmania in-
volve their ease of preparation, low toxicity, and biodegradability
and the natural liposome homing sites in macrophages of the re-
ticuloendothelium system (8). Their efficacies, however, depend
on several physical parameters, including size, phospholipid com-
position, vesicle stability, and most importantly, surface charge
(9–13). Various studies with differentially charged liposomes in-
dicate increased uptake and thus enhanced homing of cationic
liposomes, in comparison to neutral or anionic liposomes, to den-
dritic cells and macrophages (14–17). In addition, we reported
earlier that cationic liposomes bearing egg phosphatidylcholine
(PC) and stearylamine (SA) (i.e., PC-SA) had direct killing activity
against various strains of Leishmania, an activity not observed
with anionic or neutral liposomes (18, 19). Encapsulation of a
suboptimal dose of SSG within these cytolitic vesicles (i.e., PC-
SA-SSG) further enhanced the therapeutic potential of either of
the monotherapies, as single-dose treatments against both SSG-
responsive and non-SSG-responsive established murine VL (20,
21). The aim of antileishmanial therapy is to achieve maximum
efficacy with minimum doses of the drug and carrier for a success-
ful cure. Therefore, we investigated the potential leishmanicidal
activity of cationic liposomes, composed of various cationic lipids,
against both SSG-sensitive and SSG-resistant L. donovani para-
sites, in the search for an optimal formulation of SSG to treat a
nonhealing murine model of VL.
TABLE 1 Chemical formulae of cationic lipids

<table>
<thead>
<tr>
<th>Lipid</th>
<th>IUPAC name</th>
<th>Chemical formula</th>
<th>Molecular wt</th>
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<tbody>
<tr>
<td>CTAB</td>
<td>Hexadecytrimethylammonium bromide</td>
<td>C_{16}H_{34}NBBr</td>
<td>364.46</td>
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<tr>
<td>DTAB</td>
<td>Dodecytrimethylammonium bromide</td>
<td>C_{12}H_{25}NBBr</td>
<td>308.35</td>
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<tr>
<td>DDAB</td>
<td>Didodecyldimethylammonium bromide</td>
<td>C_{20}H_{41}NBBr</td>
<td>631</td>
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<tr>
<td>SA</td>
<td>Octadecan-1-amine</td>
<td>C_{18}H_{37}N</td>
<td>269.5</td>
</tr>
<tr>
<td>DOTAP</td>
<td>N-[1-(3,3-Dioleoyloxy)propyl]-N,N,N-trimethylammonium chloride</td>
<td>C_{38}H_{65}NOCl</td>
<td>698.54</td>
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MATERIALS AND METHODS

Materials. Cetyltrimethylammonium bromide (CTAB), dodecyltrimethylammonium bromide (DTAB), dimethyldioctadecylammonium bromide (DDAB), dioleoyltrimethylammonium propane (DOTAP), and egg PC were purchased from Sigma-Aldrich (St. Louis, MO). SA was purchased from Fluka (Buchs, Switzerland) (Table 1). SSG was purchased from Glucanate Health Ltd. (Kolkata, India). Minimal (M199) and RPMI 1640 medium used in the experiments were purchased from Sigma-Aldrich. Both fetal bovine serum (FBS) and antibiotics (penicillin-streptomycin) were Gibco products from Invitrogen Corp. Assay kits for performing toxicity tests (serum urea, creatinine, serum glutamate pyruvate transaminase [SGPT], and serum alkaline phosphatase [ALP] assays) were from Dr. Reddy’s Laboratories Ltd. (Hyderabad, India). 3-(4,5-Dimethyloxazol-2)-2,5-diphenyterrazolium bromide (MTT) was from MP Biomedicals, LLC (Solon, OH). All other chemicals were of analytical grade.

Animals and parasites. BALB/c mice (approximately 25 g), 4 to 6 weeks of age, were used for in vivo studies, in accordance with standard procedures approved by the Animal Ethics Committee of the Indian Institute of Chemical Biology (protocol 147/1999/CPCSEA) under the Committee for the Purpose of Control and Supervision of Experiments on Animals (CPCSEA), Ministry of Social Justice and Empowerment, Government of India. The Leishmania donovani isolates AG83 (MHOM/IN/83/AG83), GE1F8R (derived from GE1; MHOM/IN/89/GE1), and CK1R (clinical isolate, isolated as MHOM/IN/1995/CK from a non-SSG-IN/83/AG83) parasites were isolated from infected spleens and allowed to transform in vivo for 3 weeks of age, were used for in vivo studies, in accordance with standard procedures approved by the Animal Ethics Committee of the Indian Institute of Chemical Biology (protocol 147/1999/CPCSEA) under the Committee for the Purpose of Control and Supervision of Experiments on Animals (CPCSEA), Ministry of Social Justice and Empowerment, Government of India. The Leishmania donovani isolates AG83 (MHOM/IN/83/AG83), GE1F8R (derived from GE1; MHOM/IN/89/GE1), and CK1R (clinical isolate, isolated as MHOM/IN/1995/CK from a non-SSG-responsive patient) were maintained by serial passage in hamsters. Amastigotes were isolated from infected spleens and allowed to transform to promastigotes by cultivation at 22°C in complete medium, i.e., M199 (pH 7.4) supplemented with 20% heat-inactivated FBS, 2 mM glutamine, 25 mM HEPES, 100 IU/ml penicillin, and 100 μg/ml streptomycin.

Preparation of liposomes. Cationic liposomes were prepared with 20 mg of egg PC with CTAB, DTAB, DDAB, DOTAP, or SA (Table 1) at a molar ratio of 7:2, as reported previously (21), and were desiccated over night. The thin dry films were dispersed in 1 ml of 20 mM phosphate-buffered saline (PBS). For drug entrapment, lipid films containing PC-SA or PC-DDAB were dispersed in 20 mM PBS containing 1 mg/ml of SSG.

Electrophoretic characterization of liposomes. The average diameters of liposomes were determined in appropriately diluted liposome suspensions using photon correlation spectroscopy with a Nano Zs Zeta-potential analyzer (Brookhaven Instruments Corp., Holtsville, NY). Small aliquots from the different cationic liposomal preparations, prepared as described above but in 20 mM phosphate buffer without saline, were diluted to an appropriate volume with the same buffer, and the zeta potentials and mobilities of the particles were measured at 25°C (23).

Effects on L. donovani promastigotes. To investigate the effects of cationic liposomes on promastigotes freshly transformed from amastigotes of L. donovani AG83, GE1F8R, or CK1R, 2 × 10^6 parasites/ml suspended in complete medium were incubated with graded concentrations (8.85 to 212 μM) of cationic liposomes for 1 h at 37°C (18). The control parasites were incubated with 0.01% Triton X-100. Following treatment, the tubes were washed with PBS. The pellets were resuspended in 100 μl of a solution of 2 mg/ml MTT in PBS, and the mixtures were incubated for 4 h at 22°C. The reduced formazan, which is a measure of cell viability, was dissolved in dimethyl sulfoxide. The absorbance at 550 nm was measured in a spectrophotometer (Hitachi Electronics, Japan) (24).

Effects on parasite-infected macrophages. Peritoneal macrophages from naive BALB/c mice were isolated and cultured at 37°C with 5% CO2 in RPMI 1640 medium supplemented with 10% FBS, penicillin (100 IU/ml), and streptomycin (100 μg/ml). Resident macrophages (10^6 cells) were infected for 3 h at 37°C with freshly transformed promastigotes, at a macrophage/promastigote ratio of 1:10. The unphagocytosed parasites were removed by three washes with medium. Infected macrophages were then incubated at 37°C with medium containing graded concentrations of free cationic liposomes. In another experiment, infected macrophages were incubated with SSG-loaded PC-SA and PC-DDAB liposomes. After 1 h, excess liposomes were removed and the cells were placed in fresh complete RPMI 1640 medium for an additional 72 h. Cells were then air dried, fixed in methanol, and stained with Giemsa stain. The numbers of amastigotes in 200 macrophages in drug-treated and control cultures were counted microscopically (20).

Isolation of splenic amastigotes. To isolate amastigotes, 25 spleens of infected hamsters were rinsed in ice-cold 20 mM PBS with glucose (1.8%) and lightly homogenized. Macroscopic particles were allowed to settle, and the turbid suspension was decanted. This suspension was centrifuged at 100 × g for 30 min at 4°C (20). The encapsulation efficiency values for SSG within PC-SA-SSG and PC-DDAB-SSG liposomes, as measured by atomic absorption spectroscopy (AAS), were 60% and 65%, respectively (22). The PC/SA/SSG and PC/DDAB/SSG molar ratios were 7:2:0.4 and 7:2:0.6, respectively.

Physicochemical characterization of liposomes. The average diameters of liposomes were determined in appropriately diluted liposome suspensions using photon correlation spectroscopy with a Nano Zs Zeta-Sizer system (Malvern Instruments, Worcestershire, United Kingdom), as described earlier (23). The electrophoretic mobilities and zeta potentials of the complexes were measured using a Zetaplus zeta potential analyzer (Brookhaven Instruments Corp., Holtsville, NY). Small aliquots from the different cationic liposomal preparations, prepared as described above
treated promastigotes (24). Briefly, cells were fixed with 3% glutaraldehyde in PBS, postfixed with 1% OsO4 for 1 to 2 h, gradually dehydrated in ethanol, and finally embedded in Spurr’s resin. Ultrathin sections cut with a DuPont diamond knife in an ultramicrotome were stained with uranyl acetate and lead acetate and were observed with a FEI Tecnai G2 Spirit Bio Twin FP 5018/40 transmission electron microscope (FEI Co., Hillsboro, OR).

**In vitro and in vivo cytotoxicity assays.** Hemolytic assays were carried out with different concentrations of cationic liposomes incubated with packed human erythrocytes, at a ratio of 1:10 (vol/vol), for 1 h at 37°C (26). The samples were then centrifuged at 800 x g for 10 min. By measuring the optical density at 540 nm in a spectrophotometer, the release of hemoglobin from lysed erythrocytes was monitored. Controls used for 0% hemolysis (blank) and 100% hemolysis consisted of erythrocytes suspended in PBS and 1% Triton X-100, respectively.

The cytotoxicity of different cationic liposomes for peritoneal macrophages was assessed with a lactate dehydrogenase (LDH) assay (27). Adherent macrophages (10⁶ cells) were incubated with different concentrations of cationic liposomes in RPMI 1640 medium for 3 h at 37°C. LDH activity was determined by measuring the decrease in the absorbance of NADH at 340 nm in the macrophage supernatants.

Specific enzyme levels related to normal kidney and liver functions were chosen to determine the in vivo toxic effects of cationic liposomal formulations. PC-DDAB and PC-DDAB-SSG were injected intravenously (i.v.) into healthy mice at 5, 8, and 10 mg of PC/mouse in 200 μl PBS. Analyses of serum levels of creatinine, urea, and enzymes such as serum glutamate pyruvate transaminase (SGPT) and serum alkaline phosphatase (ALP) were carried out 15 days after drug administration, using the respective assay kits.

**Infection and treatment.** For experimental infections, BALB/c mice were injected with 2.5 x 10⁷ hamster spleen-derived *L. donovani* amastigotes via the tail vein. Eight-week-infected mice (6 or 7 mice per group) received a single i.v. dose of either free SSG (3 mg/kg body weight), drug-free PC-DDAB liposomes (5 mg of PC/mouse), or PC-DDAB-SSG (75 μg SSG, equivalent to 3 mg/kg, associated with 5 mg of PC/mouse). For comparative evaluation of efficacy, groups of BALB/c mice were treated with a single injection of 75 μg of SSG in 5 mg of PC either as PC-DDAB-SSG or as PC-SA-SSG. At 90 days postinfection, animals were sacrificed, with a single injection of 75 μg of SSG in 5 mg of PC (i.v.) into healthy mice at 5, 8, and 10 mg of PC/mouse in 200 μl PBS. In mice treated with other cationic liposomes, composites, and those for GE1F8R strains of promastigote cultures. Of the five cationic liposome types evaluated, PC-CTAB and PC-DOTAP had no appreciable antiparasitic effects, even at concentrations as high as 213 μM, for both AG83 and GE1F8R strains (Fig. 1). In contrast, PC-CTAB and PC-DOTAP were found to have appreciable leishmanicidal activity. PC-SA and PC-DDAB were nearly equipotent at higher doses, tested against both strains of *L. donovani*. At concentrations of 213 μM, these liposomes induced >85% killing of *L. donovani* promastigotes, compared to PBS treatment. The IC₅₀ values of PC-SA and PC-DDAB for AG83 promastigotes were 20 and 24 μM and those for GE1F8R were 52 and 70 μM, respectively (Table 3). For CK1R, all of the liposomes had high IC₅₀ values (>80 μM), with PC-DDAB being most effective at 81.21 μM.

**Antileishmanial activity of cationic liposomes against *L. donovani* promastigotes.** We reported previously that SA-bearing liposomes (PC-SA) had potent antileishmanial activity (18, 19). To determine whether liposomes prepared with other cationic lipids also had antiparasitic effects, cationic liposomes at graded concentrations were incubated for 1 h with both SSG-sensitive AG83 and SSG-resistant GE1F8R and CK1R strains of promastigote cultures. Of the five cationic liposome types evaluated, PC-CTAB and PC-DOTAP had no appreciable antiparasitic effects, even at concentrations as high as 213 μM, for both AG83 and GE1F8R strains (Table 3). For CK1R, all of the liposomes had high IC₅₀ values (>80 μM), with PC-DDAB being most effective at 81.21 μM.

**Cationic liposomes elicit antileishmanial activity against intracellular amastigotes.** Treatment of infected macrophages with various cationic liposomes resulted in dose-dependent killing of *L. donovani* amastigotes of both SSG-sensitive and SSG-resistant strains by all of the liposomes (Fig. 2). PC-SA, PC-DDAB, and PC-DOTAP induced effective inhibition of parasite growth. Results

### Table 2. Physicochemical properties of cationic liposomes

<table>
<thead>
<tr>
<th>Formulation</th>
<th>Particle diameter (nm)</th>
<th>Zeta potential (mV)</th>
<th>Electrophoretic mobility (μV/cm)</th>
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<tbody>
<tr>
<td>PC-CTAB</td>
<td>252.0 ± 39.77</td>
<td>23.2 ± 3.9</td>
<td>5.5</td>
</tr>
<tr>
<td>PC-DTAB</td>
<td>299.7 ± 23.76</td>
<td>20.3 ± 4.4</td>
<td>4.5</td>
</tr>
<tr>
<td>PC-DDAB</td>
<td>197.6 ± 10.25</td>
<td>33.6 ± 8.4</td>
<td>16.7</td>
</tr>
<tr>
<td>PC-SA</td>
<td>303.8 ± 35.22</td>
<td>23.5 ± 6</td>
<td>15</td>
</tr>
<tr>
<td>PC-DOTAP</td>
<td>240.3 ± 79.25</td>
<td>18.5 ± 2.1</td>
<td>2.5</td>
</tr>
</tbody>
</table>

### Results

**Liposome formulations and characterization.** Size measurements of the diluted liposomes formed with various cationic lipids, performed on the day of preparation, indicated average diameters of 252.0 ± 39.77 nm, 299.7 ± 23.76 nm, 197.6 ± 10.25 nm, 303.8 ± 35.22 nm, and 240.3 ± 79.25 nm for PC-CTAB, PC-DTAB, PC-DDAB, PC-SA, and PC-DOTAP, respectively, with no significant differences between the vesicles (P > 0.05) (Table 2). All liposomes were of heterogeneous size, and their polydispersity indices varied between 0.2 and 0.4. The sizes remained consistent for 4 weeks for all liposomes except for PC-DTAB, which showed an increase, probably due to aggregation. The zeta potentials of PC-CTAB, PC-DTAB, PC-DDAB, PC-SA, and PC-DOTAP liposomes were 23.2 ± 3.7 mV, 20.3 ± 4.4 mV, 33.6 ± 8.4 mV, 23.5 ± 6 mV, and 18.5 ± 2.1 mV, respectively (Table 2). The greatest electrophoretic mobility of PC-DDAB correlated with its highest zeta potential.

**Antileishmanial activity of cationic liposomes against *L. donovani* promastigotes.** We reported previously that SA-bearing liposomes (PC-SA) had potent antileishmanial activity (18, 19). To determine whether liposomes prepared with other cationic lipids also had antiparasitic effects, cationic liposomes at graded concentrations were incubated for 1 h with both SSG-sensitive AG83 and SSG-resistant GE1F8R and CK1R strains of promastigote cultures. Of the five cationic liposome types evaluated, PC-CTAB and PC-DOTAP had no appreciable antiparasitic effects, even at concentrations as high as 213 μM, for both AG83 and GE1F8R strains (Fig. 1). In contrast, PC-CTAB and PC-DOTAP were found to have appreciable leishmanicidal activity. PC-SA and PC-DDAB were nearly equipotent at higher doses, tested against both strains of *L. donovani*. At concentrations of 213 μM, these liposomes induced >85% killing of *L. donovani* promastigotes, compared to PBS treatment. The IC₅₀ values of PC-SA and PC-DDAB for AG83 promastigotes were 20 and 24 μM and those for GE1F8R were 52 and 70 μM, respectively (Table 3). For CK1R, all of the liposomes had high IC₅₀ values (>80 μM), with PC-DDAB being most effective at 81.21 μM.

**Cationic liposomes elicit antileishmanial activity against intracellular amastigotes.** Treatment of infected macrophages with various cationic liposomes resulted in dose-dependent killing of *L. donovani* amastigotes of both SSG-sensitive and SSG-resistant strains by all of the liposomes (Fig. 2). PC-SA, PC-DDAB, and PC-DOTAP induced effective inhibition of parasite growth. Results...
demonstrated that infected macrophages treated with PC-DDAB showed remarkable reductions in parasite burdens (96% and 91% for AG83 and GE1F8R, respectively), compared to macrophages treated with PC-SA (89% and 81% for AG83 and GE1F8R, respectively), after 72 h of incubation at a dose of 213 μM. Amastigotes of the clinical isolate CK1R were much more susceptible to cationic liposomes, with PC-DDAB, PC-DTAB, and PC-CTAB being more effective than PC-SA and PC-DOTAP. PC-DDAB showed an IC50 of 16.10 μM, a value 5-fold greater than the IC50 for PC-SA. In an MTT-based viability study using freshly isolated splenic amastigotes (AG83), we showed that the parasites were more susceptible to PC-DDAB than PC-SA and the activities were reversed by annexin V treatment to almost the same extents (see Fig. S1 in the supplemental material).

**Uptake of liposomes into infected macrophages.** Fig. 2C shows PC-SA (Fig. 2C, left) and PC-DDAB (Fig. 2C, right) liposomes inside infected macrophages. We microscopically visualized the intracellular fates of PC-SA and PC-DDAB liposomes by labeling the membrane with rhodamine B (red) and encapsulating the green dye 5-carboxyfluorescein (green). Intact liposomes (yellow) were seen inside the infected macrophages (nuclei stained blue), close to the amastigotes (small dots stained blue). Although we did not observe any significant differences in the uptake of liposomes at the initial time points (data not shown), liposomes bearing DDAB showed a more diffuse appearance at 2 h after treatment, whereas PC-SA appeared more punctate (Fig. 2C). It appears from the images that PC-DDAB liposomes interact better with the amastigotes, resulting in faster disintegration and release of their contents, as we could see the diffused green dye in the cytoplasm. In the case of PC-SA, most of the liposomes appeared intact, with the dye entrapped, at the same time point. Another interesting observation was that liposomes were preferentially taken up by infected macrophages versus uninfected macrophages (data not shown), probably due to changes in phagocytic behavior after infection (29). From these observations, it can be inferred that PC-DDAB could serve as an effective carrier of SSG and might deliver the drug to intracellular amastigotes better than PC-SA liposomes.

**Ultrastructural alterations induced by PC-DDAB-SSG.** Electron microscopic analysis demonstrated many alterations in the ultrastructure of promastigotes treated with PC-DDAB and PC-DDAB-SSG formulations. In contrast to untreated parasites (Fig. 3A), promastigotes treated with a 75% inhibitory concentration dose of PC-DDAB appeared with increased intracellular vacuolization (Fig. 3B). However, parasites treated with an equivalent dose of PC-DDAB with incorporated SSG became completely permeated, as manifested by a loss of electron-dense intracellular space and a loss of the shapes of cytoplasmic organelles (Fig. 3C). This effect could be caused by alterations in the physicochemical properties of the membrane system of the parasites, making them more susceptible to disruption. The ultrastructural alterations did respectively (Table 3). Amastigotes of the clinical isolate CK1R were much more susceptible to cationic liposomes, with PC-DDAB, PC-DTAB, and PC-CTAB being more effective than PC-SA and PC-DOTAP. PC-DDAB showed an IC50 of 16.10 μM, a value 5-fold greater than the IC50 for PC-SA. In an MTT-based viability study using freshly isolated splenic amastigotes (AG83), we showed that the parasites were more susceptible to PC-DDAB than PC-SA and the activities were reversed by annexin V treatment to almost the same extents (see Fig. S1 in the supplemental material).

**TABLE 3 In vitro antileishmanial activity of cationic liposomes**

<table>
<thead>
<tr>
<th>Formulation</th>
<th>AG83</th>
<th>GE1F8R</th>
<th>AG83</th>
<th>GE1F8R</th>
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<tr>
<td>PC-CTAB</td>
<td>510 ± 4</td>
<td>220 ± 9</td>
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<tr>
<td>PC-DTAB</td>
<td>53 ± 12.8</td>
<td>83 ± 8.2</td>
<td>92 ± 3.2</td>
<td>43 ± 1.3</td>
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<tr>
<td>PC-DDAB</td>
<td>24 ± 17.4</td>
<td>70 ± 1.6</td>
<td>27 ± 0.5</td>
<td>40 ± 0.8a</td>
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<tr>
<td>PC-SA</td>
<td>20 ± 33.9</td>
<td>52 ± 5</td>
<td>53 ± 2.4</td>
<td>70 ± 1.6</td>
</tr>
<tr>
<td>PC-DOTAP</td>
<td>180 ± 2.2</td>
<td>456 ± 6</td>
<td>216 ± 0.1</td>
<td>436 ± 0.7</td>
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</tbody>
</table>

a P < 0.05, compared to PC-SA.
not pinpoint any specific primary site of action of PC-DDAB-SSG on promastigotes, although it is likely that the parasite membranes are affected.

**Toxicity assays.** Considering the profound antileishmanial activity of PC-DDAB, which was even better than that of PC-SA, we proceeded to perform toxicity screening tests that are routinely used in drug development programs to determine the safety of the drug. The cytotoxic effects toward murine macrophages and human erythrocytes were measured with LDH and red blood cell (RBC) lysis assays. All of these liposomes were nontoxic in the therapeutic range of doses (see Table S1 in the supplemental material). Further *in vivo* toxicity studies were carried out with the PC-DDAB liposomes by estimating serum ALP and SGPT levels (specific enzymes related to liver dysfunction) and serum urea and creatinine levels (related to kidney dysfunction) at 15 days after injection of therapeutic as well as higher doses of liposomes, which demonstrated levels within the normal ranges (data not shown), indicating no toxicity under the given conditions.

**Combination therapy with suboptimal SSG doses in PC-DDAB liposomes showed enhanced antileishmanial efficacy.** A single suboptimal dose of SSG (3 mg/kg), both in free form and entrapped in PC-DDAB liposomes (5 mg PC/mouse), and empty PC-DDAB (5 mg PC/mouse) were administered intravenously to 60-day-infected BALB/c mice. The animals were sacrificed 30 days posttreatment to analyze the therapeutic potentiality against both SSG-sensitive AG83 and SSG-resistant GE1F8R *L. donovani* infections (Fig. 4) (20). Free SSG could suppress AG83 infection by only 26% in the liver and 10% in the spleen, with almost no activity against parasites in the bone marrow. Lower levels of parasites were observed in the group treated with only PC-DDAB vesicles (85%, 82%, and 68.4% suppression of liver, spleen, and bone marrow parasite burdens, respectively; *P* < 0.05), in comparison to untreated infected controls. Of the three therapies, combined therapy with SSG within PC-DDAB resulted in maximal inhibition, with 96.4%, 99.6%, and 86% suppression of AG83 parasite burdens in liver, spleen, and bone marrow, respectively (Fig. 4A to C). More importantly, PC-DDAB-SSG formulations could also mount successful activity against SSG-resistant GE1F8R *L. donovani* parasites, suppressing parasite burdens in liver, spleen, and bone marrow by 97.9%, 96.5%, and 84.4%, respectively, whereas free SSG was completely unsuccessful (Fig. 4D to F). The decreases in spleen and bone marrow parasite burdens induced by the combination therapy were significantly greater (*P* < 0.05) than those seen with only vesicles against both AG83 and GE1F8R strains. In addition, PC-DDAB-SSG-cured mice exhibited significant decreases (*P* < 0.05) in the liver and spleen weights, compared with untreated infected mice (Fig. 4A to D). These data indicate that therapy with SSG in PC-DDAB vesicles could overcome SSG resistance and also the natural failure of BALB/c mice to contain parasites in the visceral organs, particularly in the spleen and bone marrow, organs that are generally refractory to a broad range of chemotherapeutic interventions.

**Comparative antileishmanial activities of liposomal SSG formulations.** Recently we showed that PC-SA-SSG could cure established VL irrespective of SSG sensitivity (21). To assess better therapeutic options against established VL, a comparative study between PC-DDAB-SSG and previously reported PC-SA-SSG liposomal formulations was performed with an SSG-sensitive strain. The *in vitro* antileishmanial activities of graded doses of both cationic liposomal formulations were analyzed. We observed...
that PC-DDAB-SSG therapy was significantly more efficient than PC-SA-SSG treatment, with 8.8 μM lipids in PC-DDAB-SSG formulations suppressing 70% of the parasite burden, in contrast to 33% observed with PC-SA-SSG (P < 0.001) (Fig. 5A). The IC₅₀ calculated for PC-DDAB-SSG was 6.3 μM lipid. A 2-fold higher dose of PC-SA-SSG could induce an equivalent effect.

To determine which cationic vesicles synergized better with SSG against in vivo infections with *L. donovani*, 60-day-infected BALB/c mice were subjected to treatment with PC-SA-SSG and PC-DDAB-SSG liposomal formulations and the levels of protection were evaluated 30 days posttreatment (Fig. 5B). Seventy-five micrograms of SSG within PC-DDAB vesicles at 5 mg PC/mouse significantly induced 10⁵-fold and 10⁶-fold greater suppression of liver and spleen parasitic burdens, respectively, compared to treatment with an equivalent dose of PC-SA-SSG (P < 0.0001), demonstrating that SSG encapsulated in PC-DDAB vesicles is more efficacious than PC-SA liposomes in suppressing liver and spleen infections with AG83 parasites.

**Immunomodulatory activity of PC-DDAB-SSG.** We reported previously that, in addition to direct killing of the parasites, PC-SA-SSG at a dose of 20 mg PC/mouse could effectively downregulate proparasitic IL-10 levels and induce IFN-γ production from splenocytes, providing a host immune status favorable for cure (21). To evaluate whether PC-DDAB-SSG at 5 mg PC/mouse could be as effective as PC-SA-SSG at a dose of 20 mg PC/mouse to bring about positive immunomodulation, we injected healthy BALB/c mice with SSG entrapped in PC-SA or PC-DDAB (5 mg PC/mouse) and compared the results with those obtained with our previously reported therapeutic dose, as mentioned above. All of the liposomal treatments showed significant decreases in IL-10 levels and upregulation of IFN-γ levels (P < 0.001, one-way ANOVA) (Fig. 6A and B). Interestingly, healthy mice treated with PC-DDAB-SSG at 5 mg PC/mouse demonstrated effective downregulation of IL-10 levels and concomitant upregulation of IFN-γ levels, which were comparable to the responses observed with a 4 times higher dose of PC-SA-SSG (P > 0.05). The IFN-γ response observed with PC-SA-SSG at 5 mg PC/mouse was comparatively weaker, and although IL-10 production was suppressed to an appreciable extent, the IFN-γ upsurge was significantly greater in the PC-DDAB-SSG-treated animals (P < 0.0001). Flow cytometric analysis revealed that IFN-γ production from CD4⁺ and CD8⁺ cells increased significantly (P < 0.05, one-way ANOVA) in all treated groups, with comparable IFN-γ production with PC-SA-SSG at 20 mg PC/mouse and PC-DDAB-SSG at 5 mg PC/mouse (Fig. 6C and D). Interleukin 10 production from CD4⁺ and CD8⁺ T cells and CD19⁺ B cells decreased in all treated mice, compared to untreated controls (Fig. 6E to G), with significant decreases in CD4⁺ and CD19⁺ cells (P < 0.05, one-way ANOVA). The decrease in IL-10 levels from CD4⁺ cells observed with PC-DDAB-SSG at 5 mg PC/mouse was again comparable to that observed with PC-SA-SSG at 20 mg PC/mouse. Additionally, the decreases in intracellular IL-10 levels from CD4⁺ T cells were significant for both PC-DDAB-SSG-treated and high-dose PC-SA-SSG-treated healthy mice, compared with low-dose PC-SA-SSG-treated mice (P < 0.05) (Fig. 6E). We also checked the ability of our liposomal therapy to induce delayed-type hypersensitivity (DTH) reactions in infected hosts posttreatment. PC-DDAB and PC-DDAB-SSG showed strong DTH reactions in response to leishmanial antigens, with PC-DDAB being the major contributor. The difference in footpad swelling between PC-DDAB and PC-DDAB-SSG was insignificant, however (data not shown).

![FIG 3](http://aac.asm.org/) Electron microscopic assessment of *L. donovani* promastigotes treated with free and SSG-encapsulated PC-DDAB. (A) Control promastigotes demonstrate an elongated body and a normal aspect of the intracellular organelles. (B) The 75% effective dose (ED₇₅) of PC-DDAB induced vacuolization and alterations at the plasma membrane. (C) Parasites treated with the ED₇₅ of PC-DDAB incorporating SSG were spherical, with a complete absence of electron-dense materials. Bars, 2 μm (magnification, ×30,000) (A and B) or 1 μm (magnification, ×60,000) (C). Nu, nucleus; Mt, mitochondria; K, kinetoplast; FP, flagellar pocket; DMt, dilated mitochondria; FN, fragmented nucleus; V, vacuoles; DM, dilated matrix of mitochondria; G, granules; MB, membrane blebbing.
DISCUSSION

We first compared the physicochemical properties of the liposomes. The tissue distributions and immune-potentiating properties of liposomes depend on their size (9, 30–32). Since all of the liposomes were of similar sizes, we could not draw any correlations between their sizes and efficacies. Therefore, we next checked whether any major differences existed in the charges of the liposomes, which may account for differences in antiparasitic activities. Inhibition of parasite growth was previously shown to be associated with the positive charge of the liposomes (18). Our attempt to correlate physical properties of the cationic liposomes with potent antileishmanial activity indicated that the presence of positive charge paralleled the optimal leishmanicidal efficiency. PC-DDAB liposomes showed the highest zeta potential and electrophoretic mobility, which correlated best with its antiparasitic activity, followed by PC-SA. We had previously studied the antileishmanial activity of PC-SA liposomes (21). Therefore, all of the liposomes were compared with respect to PC-SA for equivalent or better activity.

Among all of the vesicles tested, liposomes composed of double alkyl chains, i.e., DTAB and DDAB, showed significant antileishmanial activity like PC-SA against both promastigotes and intracellular amastigotes of *L. donovani* strains AG83 and GE1F8R. Promastigotes of the clinically isolated strain CK1R were almost equally susceptible, but interestingly the intracellular amastigotes displayed much higher levels of susceptibility to the liposomes, compared to GE1F8R. All bromide-containing cationic liposomes demonstrated potent activity against intracellular amastigotes, with PC-DDAB being the most effective. Unsaturated lipid DOTAP-containing liposomes failed to show effective killing of *Leishmania* promastigotes as well as amastigotes. Earlier reports showed CTAB to be a better antibacterial agent than DDAB,
which may be due to the more bulky nature of DDAB (33). As *Leishmania* parasites are larger than bacteria and have a membrane structure that exposes negatively charged lipids on their surfaces, DDAB may be better suited for interactions with their membranes for subsequent killing. Cationic liposomes probably interact with macrophage surface receptors (34) and subsequently are endocytosed by them (35). Inside the macrophages, these cationic liposomes probably exert dual effects by interacting directly with the intracellular parasites and also activating macrophage microbicidal activity. Both PC-SA and PC-DDAB localized close to the amastigotes when observed microscopically, but PC-DDAB could more efficiently interact and elicit responses. The bulk of cationic charge on DDAB may contribute to this activity. *Leishmania* membranes consist of multiple polar and nonpolar lipids (36), which contribute to the surface charges. The difference in the susceptibility of promastigotes and amastigotes may be due to differences in the compositions of their plasma membranes (37). Freshly isolated splenic amastigotes were more susceptible to PC-DDAB treatment than PC-SA treatment, and treatment with annexin V reversed this activity, implicating a role of negatively charged membrane phosphatidylserine (PS), although contributions of other negatively charged components cannot be ruled out.

PS is expressed on the surface of infective *Leishmania*, and PS exposure has been reported to correlate with infectivity (38, 39). Thus, anti-PS killing activity through cationic liposomes can provide an effective strategy to combat the disease.

In our earlier studies, both *L. donovani* strain GE1F8R, selected in the laboratory for SSG resistance, and strain CK1R, a resistant clinical isolate (IC₅₀ values of >100 μg/ml for both strains), showed equivalent rates of death in response to PC-SA-SSG treatment (21). The strain-independent activity was due to increased retention of the drug inside the parasites. Although multiple mechanisms are involved in rendering the parasites unresponsive to pentavalent antimonials both *in vitro* and in clinical settings (40–43), the ultimate reason for nonaction of drugs is decreased uptake or poor retention in the parasite. Although we observed some differences in susceptibility of the two strains to cationic liposomes, PC-DDAB was found to be consistently efficacious toward the isolates. The resistance mechanism in the laboratory-developed GE1F8R strain was studied by Mookerjee Basu et al. (42). Infection with GE1F8R induces the upregulation of multidrug-resistance protein 1 (MRP1) and permeability glycoprotein in host cells, resulting in nonaccumulation of intracellular SSG following treatment with SSG, favoring parasite replication (42). A role of the aquaglyceroporin (AQP) family of genes in the uptake of antimonials has also been reported for some *L. donovani* isolates (43). The studies indicate that, while downregulation of AQP1 may be one of the mechanisms of antimony resistance, it is not an invariable feature of such resistance, particularly in the case of GE1F8R (44). Although the resistance mechanisms of CK1R have not been studied in detail, the differences in expression levels of proteins responsible for influx and/or efflux of the drug may account for the difference in susceptibility observed by us (44).

Interestingly, our liposomal formulation bypasses the strain-to-strain variations in susceptibility to SSG. The cationic liposomes cause depletion of cytosolic ATP (45), rendering the efflux pumps ineffective. Moreover, liposomes containing DDAB form a more rigid structure than SA in association with helper lipid PC, which allows longer retention and slow release of the drug for sustained activity (46).

Cationic liposomes have often been associated with cytotoxicity. The liposomes used here were potentially nontoxic at therapeutic doses, suggesting their suitability for further studies. Therefore, we proceeded to perform *in vivo* efficacy studies with free and drug-entrapped liposomes. The *in vivo* data indicated that therapy with 3 mg/kg body weight SSG in PC-DDAB vesicles could overcome the natural failure of BALB/c mice to contain parasites not only in the liver but also in the spleen and bone marrow, organs generally refractory to a broad range of chemotherapeutic interventions (4, 5). The antileishmanial activity results suggested synergistic activity of SSG in combination with PC-DDAB. To our knowledge, there has been no report of such profound antileishmanial activity with such a low dose of the drug encapsulated in liposomes, against SSG-sensitive and -resistant parasites. Successful therapy of VL is accompanied by antiparasitic immunity and prevention of relapse and reinfection. We report here that 75 μg of SSG in merely 5 mg PC/mouse of PC-DDAB could effectively downregulate parasitic IL-10 levels, providing a favorable immune status in the host. Recent studies by Mukherjee et al. (47) show increased IL-10 secretion and resultant MDRI upregulation in SSG-resistant clinical isolates. In light of this finding, we may propose that liposomal SSG reverses this situation in favor of the
ELISA (A and B) and fluorescence-activated cell sorting analysis (C to G) of splenocytes from normal BALB/c mice treated with PC-SA-SSG or PC-DDAB-SSG. (A and B) Splenocytes were isolated from mice treated with PC-SA-SSG or PC-DDAB-SSG liposomes and were cultured for determination of IFN-γ (A) and IL-10 (B) levels. (C to G) Splenocytes treated with PC-SA-SSG or PC-DDAB-SSG were gated for CD4⁺ (C and D), CD8⁺ (D and F), and CD19⁺ (G) cells and permeabilized, and intracellular IFN-γ (C and D) and IL-10 (E, F, and G) levels were determined. Data represent means ± standard errors ($n = 4$). ***, $P < 0.0001$; *, $P < 0.05$. 

FIG 6
host. Antiparasitic IFN-γ secretion is the key to clearance of infection and establishment of host protection (48). Liposomes containing saturated lipids preferably induce Th1-type immune responses (49) and DDAB is a known IFN-γ inducer (50). High concentrations of this cytokine in mice treated with PC-DDAB-SSG strengthen our claim for its profound leishmanicidal activity. Previously we observed synergism between PC-SA and SSG for IL-10 suppression in vitro (21). In the present study, however, we compared the therapeutically active dose of PC-DDAB-SSG with an equivalent dose of PC-SA-SSG.

The antiparasitic activity of positively charged SA-bearing liposomes was well observed against Toxoplasma, Trypanosoma, and Leishmania parasites (18, 19, 51, 52), and this might be attributed to charge-based interactions of cationic liposomes with negatively charged membranes of parasites (45, 53, 54), followed by direct cytotoxic action. Vesicles formed of DDAB exhibited antibacterial activity against different species of bacteria (55, 56) and fungicidal activity against Candida albicans, which were attributed to surface charge interactions (56, 57). Charge-based interactions were reported for antimicrobial cationic peptides such as magainin 2, temporins, and CA(1-8)M(1-18) (58, 59), which interact with pathogen membranes, leading to degeneration of intracellular organelles, blurred definition of the boundaries of intracellular organelles, increases in electron-translucent intracellular space, and apparent expansion and disruption of membrane structures. Here, among other vesicles, we have screened DDAB vesicles for maximal efficacy against Leishmania promastigotes and amastigotes. Experiments with DDAB-containing liposomes have pointed toward nonionic and hydrophobic interactions with membrane lipids or the hydrophobic portions of membrane proteins (60), although some degree of ionic interactions with phosphate groups of phospholipids and other negatively charged components of the membrane cannot be ruled out. It is possible that the stable binding of DDAB liposomes to parasite membranes allows the drug to enter inside, adding to the cellular disruption observed in the electron microscopic experiments. Although there are reports that promastigotes are insensitive to pentavalent antimony, studies show that high concentrations of the drug make promastigotes sensitive (40, 61). Furthermore, DDAB has been established as a potent adjuvant, mounting strong cell-mediated immune responses and proinflammatory reactions (30, 50). The immunoadjuvant properties of DDAB have been attributed to longer availability of DDAB-bearing liposomes at the site of injection and recruitment of monocytes for the generation of effective immune responses (62). Herein, a strong Th1 immune response synergizes with the parasite killing to confer better therapeutic protection to the host. Although the immune reactions that follow PC-DDAB-SSG therapy remain to be studied, triggering a strong antiparasitic immune environment in healthy mice injected with the formulation resulted in favorable modulation of the host immune responses.

Various liposomal formulations of SSG have been reported to elicit significant leishmanicidal activity in vivo in chronically infected BALB/c mice. Although these formulations demonstrated spectacular success against liver parasites (4–6, 63), they failed to perform in spleen (6, 64). Our previous liposomal SSG formulation with cationic SA-bearing liposomes could successfully eliminate SSG-sensitive and SSG-resistant parasites from liver, spleen, and bone marrow. However, these activities were obtained with the dose of 20 mg PC/mouse. In the present study, we demonstrated that 75 μg of SSG in 5 mg PC/mouse of PC-DDAB was enough for eradication of parasites from the liver, spleen, and bone marrow of the animals. The dose of SSG that could elicit such potent curative responses in a single administration was only 3 mg/kg body weight, 1/100th of the dose of free drug that is optimal for treatment (64). Such reductions in liposome and drug dosages may be favorable for the host cells, preventing unnecessary toxicity and attenuating the chances of refractoriness to antimonial therapy, making PC-DDAB-SSG a prospective antileishmanial agent.

The results obtained strongly demonstrate the advantages of using PC-DDAB encapsulating a very low dose of SSG against L. donovani infections, irrespective of SSG sensitivity. The use of SSG has been discontinued in India due to the emergence of clinical resistance. Since our liposomal formulation works by bypassing the resistance-inducing mechanisms of the parasite, we propose that this kind of approach can be developed further to increase the clinical life of SSG and other drugs that are prone to resistance development.

The liposomal formulation of SSG prepared in PC-DDAB confers cures against SSG-sensitive and SSG-resistant L. donovani infections in BALB/c mice at a suboptimal dose of the drug not reported earlier. This therapeutic strategy involving efficient targeting and immunomodulation can be utilized for the development of safe, single-shot, therapeutic agents not only against Leishmania infections but also against other parasitic diseases and cancer, where drug resistance limits the use of various potent therapeutic agents.

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We declare no conflicts of interest.

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