

Decreasing *pfmdr1* Copy Number Suggests that *Plasmodium falciparum* in Western Cambodia Is Regaining *In Vitro* Susceptibility to Mefloquine

Pharath Lim,^{a,b} Dalin Dek,^b Vorleak Try,^b Sokunthea Sreng,^b Seila Suon,^b Rick M. Fairhurst^a

Laboratory of Malaria and Vector Research, National Institute of Allergy and Infectious Diseases, National Institutes of Health, Rockville, Maryland, USA^a; National Center for Parasitology, Entomology, and Malaria Control, Phnom Penh, Cambodia^b

Dihydroartemisinin-piperaquine is the current frontline artemisinin combination therapy (ACT) for *Plasmodium falciparum* malaria in Cambodia but is now failing in several western provinces. To investigate artesunate plus mefloquine (AS+MQ) as a replacement ACT, we measured the prevalence of multiple *pfmdr1* copies—a molecular marker for MQ resistance—in 844 *P. falciparum* clinical isolates collected in 2008 to 2013. The *pfmdr1* copy number is decreasing in Western Cambodia, suggesting that *P. falciparum* is regaining *in vitro* susceptibility to MQ.

Artemisinin-based combination therapy (ACT) is used worldwide to treat uncomplicated *Plasmodium falciparum* malaria. Artesunate plus mefloquine (AS+MQ) was adopted as Cambodia's first-line treatment in 2000. AS+MQ treatment failures were first observed in 10 to 20% of patients in Pailin and Battambang Provinces, Western Cambodia, in 2003 to 2004 (1, 2). The presence of multiple (i.e., ≥ 2) *pfmdr1* copies—a genetic marker of MQ resistance—was associated with AS+MQ failures and reduced parasite *in vitro* susceptibility to MQ not only in Pailin and Battambang but in neighboring Pursat and Kampot Provinces as well (3–5). This development, along with emerging evidence of reduced *in vitro* susceptibility to MQ in other provinces (6), prompted Cambodia's National Malaria Control Program to adopt dihydroartemisinin-piperaquine (DHA-PPQ) as the first-line ACT in Western Cambodia in 2008 and in the entire country in 2010. Unfortunately, recent clinical studies suggest that the efficacy of DHA-PPQ is rapidly declining in five Western Cambodian provinces: Pursat, Battambang, Pailin, Oddar Meanchey, and Preah Vihear. For example, DHA-PPQ treatment failures were observed in 25% and 11% of patients in Pailin and Pursat in 2010 (7) and in 36% of patients in Oddar Meanchey in 2013 (8). The rapidly increasing prevalence of DHA-PPQ failures in these provinces, likely due to entrenched artemisinin resistance (9, 10) and suspected PPQ resistance (7, 8), demands additional evaluations of newer antimalarial drugs (11, 12), as well as reevaluation of previously used ACTs.

Only a few ACTs, including AS+MQ and artemether-lumefantrine (AL), are presently available for widespread use in Cambodia to replace DHA-PPQ as it fails. Although AS+MQ and AL treatment failures have previously been reported from Battambang and Pursat (2, 13), the clinical efficacy of these regimens in the setting of DHA-PPQ resistance is not known. However, studies have suggested a lack of cross-resistance between PPQ and MQ *in vitro*; for example, *pfmdr1* amplification is associated with decreased susceptibility to MQ but increased susceptibility to PPQ (14). Given these and related findings (15), and the fact that earlier use of DHA-PPQ was associated with decreasing prevalence of multiple *pfmdr1* copies in Pailin (from 33% in 2005 to 5% in 2007) (16), we hypothesized that the recent substantially reduced use of AS+MQ in Pursat and Preah Vihear would select for parasites

that have regained sensitivity to MQ. To explore this hypothesis, we used real-time PCR methods to quantify the *pfmdr1* copy number in 844 *P. falciparum* clinical isolates collected in 2008 to 2013 from Pursat and Preah Vihear, as well as Ratanakiri Province in Eastern Cambodia, where DHA-PPQ treatment failures have not been documented. *pfmdr1* copy numbers for 360 of these isolates were previously reported (17). Parasitized whole-blood samples were collected from patients enrolled in completed parasite clearance rate (10, 18) and ongoing DHA-PPQ efficacy (ClinicalTrials.gov Identifier, NCT01736319) studies. All patients or their parents provided written informed consent. The *pfmdr1* copy number in extracted DNA samples was quantified as previously described (17), and the *P. falciparum* lines 3D7 (copy number normalized to 1) and Dd2 (mean copy number \pm SD, 2.05 ± 0.19 ; $n = 29$) were used as controls.

The distribution of 844 parasite isolates by province and year, in relation to Cambodia's recommended use of ACTs, is shown in Table 1. The prevalence of parasites with multiple *pfmdr1* copies decreased significantly in Pursat during the 2008-to-2013 period (Cochran-Armitage trend test, $P < 0.001$), decreased in Preah Vihear during the 2011-to-2013 period ($P = 0.065$), and did not change in Ratanakiri in the 2010-to-2013 period ($P = 0.869$). The latter finding may be explained by the fact that Cambodia's remaining stocks of AS+MQ were still being distributed to Ratanakiri until mid-2013, which may have maintained the low prevalence of multiple *pfmdr1* copies in this province. In 2013, the prevalence of parasites with multiple *pfmdr1* copies was estimated

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Address correspondence to Rick M. Fairhurst, rfairhurst@niaid.nih.gov.

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TABLE 1 *pfmdr1* copy number in 844 *P. falciparum* clinical isolates, stratified by Cambodian province and year of collection

Province	<i>pfmdr1</i> copy no.	No. (%) of isolates with indicated copy no. where the recommended ACT(s) ^a was as indicated in:						<i>P</i> value ^b
		2008	2009	2010	2011	2012	2013	
Pursat		AS+MQ, DHA-PPQ	DHA-PPQ	DHA-PPQ	DHA-PPQ	DHA-PPQ	DHA-PPQ	<0.001
	1	19 (50)	64 (70)	40 (67)	73 (66)	44 (90)	64 (90)	
	2	10 (26)	9 (10)	9 (15)	11 (10)	3 (6)	6 (9)	
	3	6 (16)	13 (14)	8 (13)	14 (13)	1 (2)	1 (1)	
	4	2 (5)	4 (5)	1 (2)	7 (6)	1 (2)	0 (0)	
5	1 (3)	1 (1)	2 (3)	5 (5)	0 (0)	0 (0)		
Preah Vihear		AS+MQ	AS+MQ	AS+MQ	AS+MQ	AS+MQ, DHA-PPQ	AS+MQ, DHA-PPQ	0.065
	1	NA ^c	NA	NA	63 (78)	67 (93)	28 (88)	
	2	NA	NA	NA	10 (12)	2 (3)	2 (6)	
	3	NA	NA	NA	2 (3)	3 (4)	2 (6)	
	4	NA	NA	NA	1 (1)	0 (0)	0 (0)	
5	NA	NA	NA	5 (6)	0 (0)	0 (0)		
Ratanakiri		AS+MQ	AS+MQ	AS+MQ	AS+MQ	AS+MQ, DHA-PPQ	AS+MQ, DHA-PPQ	0.869
	1	NA	NA	54 (100)	91 (91)	52 (100)	33 (97)	
	2	NA	NA	0 (0)	8 (8)	0 (0)	1 (3)	
	3	NA	NA	0 (0)	0 (0)	0 (0)	0 (0)	
	4	NA	NA	0 (0)	0 (0)	0 (0)	0 (0)	
5	NA	NA	0 (0)	1 (1)	0 (0)	0 (0)		

^a DHA-PPQ, combination therapy of dihydroartemisinin and piperazine; AS+MQ, combination therapy of artesunate and mefloquine.

^b *P* values were calculated using the Cochran-Armitage trend test (exact *P* values were estimated by Monte Carlo simulation using the coin R package; number of Monte Carlo replicates = 10⁶). Boldface indicates a statistically significant difference.

^c NA, not available.

TABLE 2 *In vitro* susceptibility of *P. falciparum* clinical isolates to eight antimalarial drugs in 2010 to 2013, stratified by Cambodian province and *pfmdr1* copy number

Province	Drug	Susceptibility of isolates with <i>pfmdr1</i> copy number of:				<i>P</i> value ^b
		One		Multiple		
		No. of isolates	IC ₅₀ (nM) [GM ^a (range)]	No. of isolates	IC ₅₀ (nM) [GM ^a (range)]	
Pursat	Chloroquine	133	474.4 (103.9–1,084.3)	52	294.1 (27.6–1,312.8)	<0.0001
	Mefloquine	172	14.2 (1.7–56.4)	53	28.2 (7.8–71.3)	<0.0001
	Quinine	174	273.2 (65.5–992.0)	56	319.4 (95.5–825.4)	0.0311
	Piperaquine	100	45.8 (5.6–185.2)	40	24.0 (2.5–125.7)	0.0002
	Atovaquone	64	0.5 (0.1–8.3)	6	0.8 (0.2–13.8)	0.7510
	Pyronaridine	64	5.4 (0.2–16.5)	6	3.3 (0.7–10.3)	0.1726
	Artesunate	148	2.9 (0.6–8.1)	40	4.0 (1.2–8.7)	0.0003
	DHA	176	2.7 (0.9–9.8)	56	3.0 (0.7–7.9)	0.1120
Preah Vihear	Chloroquine	105	318.4 (16.7–978.5)	17	339.4 (108.7–864.2)	0.5518
	Mefloquine	111	18.4 (3.6–46.6)	18	27.2 (4.3–69.7)	0.0100
	Quinine	113	200.8 (31.4–956.5)	19	297.3 (61.8–933.1)	0.0231
	Piperaquine	107	27.9 (4.9–130.1)	19	25.8 (7.9–66.2)	0.6976
	Atovaquone	27	0.6 (0.1–9.3)	3	0.4 (0.2–1.2)	0.3837
	Pyronaridine	27	4.5 (1.4–9.2)	3	6.6 (3.2–12.8)	0.4265
	Artesunate	111	2.4 (0.4–9.5)	19	3.8 (1.4–10.0)	0.0132
	DHA	111	2.1 (0.5–6.7)	18	2.8 (0.9–7.2)	0.0753
Ratanakiri	Chloroquine	155	175.8 (11.8–933.1)	5	149.6 (39.3–301.3)	0.6912
	Mefloquine	155	18.9 (4.2–64.7)	5	16.9 (9.7–32.5)	0.6554
	Quinine	160	131.8 (10.2–695.8)	4	184.4 (72.8–351.3)	0.3763
	Piperaquine	119	24.1 (5.5–67.1)	5	36.2 (13.6–72.3)	0.1567
	Atovaquone	28	0.4 (0.1–2.4)	0	NA ^c	NA
	Pyronaridine	29	3.6 (1.2–9.8)	0	NA	NA
	Artesunate	119	2.0 (0.9–5.6)	4	2.7 (2.3–4.7)	0.1619
	DHA	161	1.8 (0.5–5.8)	4	1.7 (1.0–2.8)	0.7787

^a GM, geometric mean.

^b *P* values were calculated using the Mann-Whitney test. Boldface indicates a statistically significant difference.

^c NA, not available.

to be 10%, 12%, and 3% in Pursat, Preah Vihear, and Ratanakiri, respectively. Together, these data suggest that MQ resistance is presently decreasing in Pursat and Preah Vihear and remains consistently low in Ratanakiri, likely due to the relatively low prevalence of artemisinin resistance in this province (9, 10). To further investigate these possibilities, we measured the *in vitro* susceptibility of 514/844 (60.9%) parasite isolates to MQ using a SYBR green I-based growth inhibition assay as previously described (17). The 50% inhibitory concentrations (IC_{50} s) of at least four drugs were previously reported for 242 of these isolates (17). Parasites with multiple *pfmdr1* copies had higher geometric mean MQ IC_{50} s than those with one *pfmdr1* copy in Pursat (28.2 versus 14.2 nM, $P < 0.0001$, Mann-Whitney test) and Preah Vihear (27.2 versus 18.4 nM, $P = 0.0100$) (Table 2). Taken together, these genotype and phenotype data suggest that AS+MQ may now be effective for the treatment of malaria in Pursat, Preah Vihear, and Ratanakiri.

We also examined the association of *pfmdr1* copy number and the *in vitro* susceptibility of parasite isolates to chloroquine (CQ; $n = 467$), quinine (QN; $n = 526$), PPQ ($n = 390$), atovaquone (ATV; $n = 128$), pyronaridine (PYN; $n = 129$), AS ($n = 441$), and DHA ($n = 526$). As for MQ, parasites carrying one *pfmdr1* copy were significantly more susceptible to QN and AS in Pursat and Preah Vihear. As observed previously (14, 19–21), parasites harboring one *pfmdr1* copy were significantly less susceptible to CQ and PPQ in Pursat (Table 2), indicating that the mechanisms of resistance to MQ and QN differ from those mediating resistance to CQ and PPQ in this province. In Ratanakiri, *pfmdr1* copy number did not associate with IC_{50} for any of the antimalarial drugs tested, but the number of samples with multiple *pfmdr1* copies were very few (≤ 5) in this province. These data indicate that *pfmdr1* amplification is involved in decreasing parasite susceptibility to MQ and QN but not to CQ and PPQ in Cambodia. Whether the widespread decrease in the use of AS+MQ, increase in the use of DHA-PPQ, or both are driving the deamplification of *pfmdr1* requires further investigation. As expected, parasite isolates are highly susceptible to atovaquone and pyronaridine, which have not been extensively used as ACT partner drugs in Cambodia (12, 22).

In summary, our data indicate that *P. falciparum* is regaining *in vitro* susceptibility to MQ in Pursat and, possibly, in Preah Vihear as well and suggest that AS+MQ may be an effective first-line treatment for *P. falciparum* malaria in Cambodian provinces where DHA-PPQ treatment failures have been documented—a possibility we are presently investigating. In other areas of Southeast Asia where DHA-PPQ treatment failures are suspected, early quantification of the *pfmdr1* copy number and MQ IC_{50} s of contemporary isolates is a useful approach to rapidly investigate the suitability of AS+MQ as an alternative ACT.

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REFERENCES

- Denis MB, Tsuyuoka R, Poravuth Y, Narann TS, Seila S, Lim C, Incardona S, Lim P, Sem R, Socheat D, Christophel EM, Ringwald P. 2006. Surveillance of the efficacy of artesunate and mefloquine combination for the treatment of uncomplicated falciparum malaria in Cambodia. *Trop Med Int Health* 11:1360–1366. <http://dx.doi.org/10.1111/j.1365-3156.2006.01690.x>.
- Denis MB, Tsuyuoka R, Lim P, Lindegardh N, Yi P, Top SN, Socheat D, Fandeur T, Annerberg A, Christophel EM, Ringwald P. 2006. Efficacy of artemether-lumefantrine for the treatment of uncomplicated falciparum malaria in northwest Cambodia. *Trop Med Int Health* 11:1800–1807. <http://dx.doi.org/10.1111/j.1365-3156.2006.01739.x>.
- Lim P, Alker AP, Khim N, Shah NK, Incardona S, Doung S, Yi P, Bouth DM, Bouchier C, Puijalon OM, Meshnick SR, Wongsrichanalai C, Fandeur T, Le Bras J, Ringwald P, Arie F. 2009. Pfm1 copy number and artemisinin derivatives combination therapy failure in falciparum malaria in Cambodia. *Malaria J* 8:11. <http://dx.doi.org/10.1186/1475-2875-8-11>.
- Rogers WO, Sem R, Tero T, Chim P, Lim P, Muth S, Socheat D, Arie F, Wongsrichanalai C. 2009. Failure of artesunate-mefloquine combination therapy for uncomplicated Plasmodium falciparum malaria in southern Cambodia. *Malaria J* 8:10. <http://dx.doi.org/10.1186/1475-2875-8-10>.
- Alker AP, Lim P, Sem R, Shah NK, Yi P, Bouth DM, Tsuyuoka R, Maguire JD, Fandeur T, Arie F, Wongsrichanalai C, Meshnick SR. 2007. Pfm1 and *in vivo* resistance to artesunate-mefloquine in falciparum malaria on the Cambodian-Thai border. *Am J Trop Med Hyg* 76:641–647.
- Lim P, Wongsrichanalai C, Chim P, Khim N, Kim S, Chy S, Sem R, Nhem S, Yi P, Duong S, Bouth DM, Genton B, Beck HP, Gobert JG, Rogers WO, Coppee JY, Fandeur T, Mercereau-Puijalon O, Ringwald P, Le Bras J, Arie F. 2010. Decreased *in vitro* susceptibility of Plasmodium falciparum isolates to artesunate, mefloquine, chloroquine, and quinine in Cambodia from 2001 to 2007. *Antimicrob Agents Chemother* 54:2135–2142. <http://dx.doi.org/10.1128/AAC.01304-09>.
- Leang R, Barrette A, Bouth DM, Menard D, Abdur R, Duong S, Ringwald P. 2013. Efficacy of dihydroartemisinin-piperazine for treatment of uncomplicated Plasmodium falciparum and Plasmodium vivax in Cambodia, 2008 to 2010. *Antimicrob Agents Chemother* 57:818–826. <http://dx.doi.org/10.1128/AAC.00686-12>.
- Saunders DL, Vanachayangkul P, Lon C, U.S. Army Military Malaria Research Program, National Center for Parasitology, Entomology, and Malaria Control (CNM), Royal Cambodian Armed Forces. 2014. Dihydroartemisinin-piperazine failure in Cambodia. *N Engl J Med* 371:484–485. <http://dx.doi.org/10.1056/NEJMc1403007>.
- Arie F, Witkowski B, Amaratunga C, Beghain J, Langlois AC, Khim N, Kim S, Duru V, Bouchier C, Ma L, Lim P, Leang R, Duong S, Sreng S, Suon S, Chuor CM, Bout DM, Menard S, Rogers WO, Genton B, Fandeur T, Miotto O, Ringwald P, Le Bras J, Berry A, Barale JC, Fairhurst RM, Benoit-Vical F, Mercereau-Puijalon O, Menard D. 2014. A molecular marker of artemisinin-resistant Plasmodium falciparum malaria. *Nature* 505:50–55.
- Ashley EA, Dhorda M, Fairhurst RM, Amaratunga C, Lim P, Suon S, Sreng S, Anderson JM, Mao S, Sam B, Sopha C, Chuor CM, Nguon C, Sovannaroeth S, Pukrittayakamee S, Jittamala P, Chotivanich K, Chutasmit K, Suchatsoonthorn C, Runcharoen R, Hien TT, Thuy-Nhien NT, Thanh NV, Phu NH, Htut Y, Han KT, Aye KH, Mokuolu OA, Olaosebikan RR, Folaranmi OO, Mayxay M, Khanthavong M, Hongy-anthong B, Newton PN, Onyamboko MA, Fanello CI, Tshefu AK, Mishra N, Valecha N, Phyo AP, Nosten F, Yi P, Tripura R, Borrmann S, Bashraheil M, Peshu J, Faiz MA, Ghose A, Hossain MA, Samad R, Rahman MR, et al. 2014. Spread of artemisinin resistance in Plasmodium falciparum malaria. *N Engl J Med* 371:411–423. <http://dx.doi.org/10.1056/NEJMoa1314981>.
- White NJ, Pukrittayakamee S, Phyo AP, Rueangweeraayut R, Nosten F, Jittamala P, Jeeyapant A, Jain JP, Lefevre G, Li R, Magnusson B, Diagona TT, Leong FJ. 2014. Spiroindolone KAE609 for falciparum and vivax malaria. *N Engl J Med* 371:403–410. <http://dx.doi.org/10.1056/NEJMoa1315860>.
- Rueangweeraayut R, Phyo AP, Uthaisin C, Poravuth Y, Binh TQ, Tinto H, Penali LK, Valecha N, Tien NT, Abdulla S, Borghini-Fuhrer I, Duparc S, Shin CS, Fleckenstein L, Pyronaridine-Artesunate Study Team. 2012. Pyronaridine-artesunate versus mefloquine plus artesunate

- for malaria. *N Engl J Med* 366:1298–1309. <http://dx.doi.org/10.1056/NEJMoa1007125>.
13. Song J, Socheat D, Tan B, Seila S, Xu Y, Ou F, Sokunthea S, Sophorn L, Zhou C, Deng C, Wang Q, Li G. 2011. Randomized trials of artemisinin-piperaquine, dihydroartemisinin-piperaquine phosphate and artemether-lumefantrine for the treatment of multi-drug resistant falciparum malaria in Cambodia-Thailand border area. *Malaria J* 10:231. <http://dx.doi.org/10.1186/1475-2875-10-231>.
 14. Veiga MI, Ferreira PE, Malmberg M, Jornhagen L, Bjorkman A, Nosten F, Gil JP. 2012. *pfmdr1* amplification is related to increased Plasmodium falciparum in vitro sensitivity to the bisquinoline piperaquine. *Antimicrob Agents Chemother* 56:3615–3619. <http://dx.doi.org/10.1128/AAC.06350-11>.
 15. Eastman RT, Dharia NV, Winzeler EA, Fidock DA. 2011. Piperaquine resistance is associated with a copy number variation on chromosome 5 in drug-pressured Plasmodium falciparum parasites. *Antimicrob Agents Chemother* 55:3908–3916. <http://dx.doi.org/10.1128/AAC.01793-10>.
 16. Imwong M, Dondorp AM, Nosten F, Yi P, Mungthin M, Hanchana S, Das D, Phyo AP, Lwin KM, Pukrittayakamee S, Lee SJ, Saisung S, Koecharoen K, Nguon C, Day NP, Socheat D, White NJ. 2010. Exploring the contribution of candidate genes to artemisinin resistance in Plasmodium falciparum. *Antimicrob Agents Chemother* 54:2886–2892. <http://dx.doi.org/10.1128/AAC.00032-10>.
 17. Lim P, Dek D, Try V, Eastman RT, Chy S, Sreng S, Suon S, Mao S, Sopha C, Sam B, Ashley EA, Miotto O, Dondorp AM, White NJ, Su XZ, Char MC, Anderson JM, Amaratunga C, Menard D, Fairhurst RM. 2013. Ex vivo susceptibility of Plasmodium falciparum to antimalarial drugs in western, northern, and eastern Cambodia, 2011–2012: association with molecular markers. *Antimicrob Agents Chemother* 57:5277–5283. <http://dx.doi.org/10.1128/AAC.00687-13>.
 18. Amaratunga C, Sreng S, Suon S, Phelps ES, Stepniewska K, Lim P, Zhou C, Mao S, Anderson JM, Lindegardh N, Jiang H, Song J, Su XZ, White NJ, Dondorp AM, Anderson TJ, Fay MP, Mu J, Duong S, Fairhurst RM. 2012. Artemisinin-resistant Plasmodium falciparum in Pursat province, western Cambodia: a parasite clearance rate study. *Lancet Infect Dis* 12: 851–858. [http://dx.doi.org/10.1016/S1473-3099\(12\)70181-0](http://dx.doi.org/10.1016/S1473-3099(12)70181-0).
 19. Barnes DA, Foote SJ, Galatis D, Kemp DJ, Cowman AF. 1992. Selection for high-level chloroquine resistance results in deamplification of the *pfmdr1* gene and increased sensitivity to mefloquine in Plasmodium falciparum. *EMBO J* 11:3067–3075.
 20. Duraisingh MT, von Seidlein LV, Jepson A, Jones P, Sambou I, Pinder M, Warhurst DC. 2000. Linkage disequilibrium between two chromosomally distinct loci associated with increased resistance to chloroquine in Plasmodium falciparum. *Parasitology* 121(Pt 1):1–7. <http://dx.doi.org/10.1017/S0031182099006022>.
 21. Pickard AL, Wongsrichanalai C, Purfield A, Kamwendo D, Emery K, Zalewski C, Kawamoto F, Miller RS, Meshnick SR. 2003. Resistance to antimalarials in Southeast Asia and genetic polymorphisms in *pfmdr1*. *Antimicrob Agents Chemother* 47:2418–2423. <http://dx.doi.org/10.1128/AAC.47.8.2418-2423.2003>.
 22. Hoyer S, Nguon S, Kim S, Habib N, Khim N, Sum S, Christophel EM, Borge S, Thomson A, Kheng S, Chea N, Yok S, Top S, Ros S, Sophal U, Thompson MM, Mellor S, Arie F, Witkowski B, Yeang C, Yeung S, Duong S, Newman RD, Menard D. 2012. Focused screening and treatment (FSAT): a PCR-based strategy to detect malaria parasite carriers and contain drug resistant *P. falciparum*, Pailin, Cambodia. *PLoS One* 7:e45797. <http://dx.doi.org/10.1371/journal.pone.0045797>.